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Only for nonprovisional applications under 37 CFR § 1.53(b))

Box Patent Application
Assistant Commissioner for Patent
Washington, D.C. 20231

See MPEP chapter 600 concerning utility patent application contents.

1. ☐ General Authorization Form & Fee Transmittal
(Submit an original and a duplicate for fee processing)
2. ☒ Specification [Total Pages]
(preferred arrangement set forth below)
- Descriptive Title of the Invention
 - Cross References to Related Applications
 - Statement Regarding Fed sponsored R & D
 - Reference to Microfiche Appendix
 - Background of the Invention
 - Brief Summary of the Invention
 - Brief Description of the Drawings (if filed)
 - Detailed Description
 - Claim(s)
 - Abstract of the Disclosure
3. ☒ Drawing(s) (35 USC 113) [Total Sheets]
4. Oath or Declaration [Total Pages]
- a. ☐ Newly executed (original or copy)
- b. ☐ Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 17 completed)
- i. ☐ DELETION OF INVENTOR(S)
Signed statement attached deleting
inventor(s) named in the prior application,
see 37 CFR 1.63(d)(2) and 1.33(b)
5. ☐ Incorporation By Reference (useable if box 4b is checked) The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered to be part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

6. ☐ Microfiche Computer Program (*Appendix*)
7. Nucleotide and Amino Acid Sequence Submission
(*if applicable, all necessary*)
- a. ☒ Computer-Readable Copy
- b. ☒ Paper Copy (identical to computer copy)
- c. ☒ Statement verifying identity of above copies

8. ☐ Assignment Papers (cover sheet & document(s))

9. ☐ 37 CFR 3.73(b) Statement ☐ Power of Attorney
(when there is an assignee)

10. ☐ English Translation Document (if applicable)

11. ☒ Information Disclosure ☐ Copies of IDS
Statement (IDS)/PTO-1449 Citations

12. ☐ Preliminary Amendment

13. ☒ Return Receipt Postcard

14. ☐ Small Entity ☐ Statement filed in prior application,
Statement(s) Status still proper and desired

15. ☐ Certified Copy of Priority Document(s)
(if foreign priority is claimed)

16. ☒ Other: Certificate of Express Mail

17. If a **CONTINUING APPLICATION**, check appropriate box and supply the requisite information below and in a preliminary amendment

☐ Continuation ☐ Divisional ☒ Continuation-In-Part (CIP) of prior Application No.: **09/590,583**

Prior application information: Examiner _____ Group / Art Unit _____

☐ Claims the benefit of Provisional Application No. _____

1. The first step in the process is to identify the problem or issue that needs to be addressed. This involves gathering information and understanding the context of the problem.

2. The second step is to define the objectives and goals of the project. This involves determining what you want to achieve and how you will measure success.

3. The third step is to develop a plan of action. This involves identifying the steps that need to be taken to achieve the objectives and goals.

4. The fourth step is to implement the plan. This involves putting the plan into action and monitoring progress.

5. The fifth step is to evaluate the results. This involves assessing the outcomes of the project and determining whether the objectives and goals have been achieved.

6. The sixth step is to report on the results. This involves communicating the findings of the project to the relevant stakeholders.

7. The seventh step is to review the process. This involves reflecting on the project and identifying areas for improvement.

8. The eighth step is to share the results. This involves disseminating the findings of the project to a wider audience.

9. The ninth step is to celebrate success. This involves acknowledging the achievements of the project and the team.

10. The tenth step is to learn from the experience. This involves reflecting on the project and identifying lessons learned for future projects.

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REGISTRATION NO. 46,209

Date October 26, 2000

COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF BREAST CANCER

CROSS REFERENCE TO RELATED APPLICATIONS

5 This application is a continuation-in-part of U.S. Patent Application No. 09/590,583, filed June 8, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/577,505, filed May 24, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/534,825, filed March 22, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/429,755, filed October 28, 1999, which is a continuation-in-part
10 of U.S. Patent Application No. 09/289,198, filed April 9, 1999, which is a continuation-in-part of U.S. Patent Application No. 09/062,451, filed April 17, 1998, which is a continuation in part of U.S. Patent Application No. 08/991,789, filed December 11, 1997, which is a continuation-in-part of U.S. Patent Application No. 08/838,762, filed April 9, 1997, now abandoned, which claims priority from International Patent Application
15 No. PCT/US97/00485, filed January 10, 1997, and is a continuation-in-part of U.S. Patent Application No. 08/700,014, filed August 20, 1996, which is a continuation-in-part of U.S. Patent Application No. 08/585,392, filed January 11, 1996, now abandoned.

TECHNICAL FIELD OF THE INVENTION

20 The present invention relates generally to therapy and diagnosis of cancer, such as breast cancer. The invention is more specifically related to polypeptides, comprising at least a portion of a breast tumor protein, and to polynucleotides encoding such polypeptides. Such polypeptides and polynucleotides are useful in pharmaceutical compositions, *e.g.*, vaccines, and other compositions for the diagnosis and treatment of breast cancer.

25 BACKGROUND OF THE INVENTION

Breast cancer is a significant health problem for women in the United States and throughout the world. Although advances have been made in detection and treatment

of the disease, breast cancer remains the second leading cause of cancer-related deaths in women, affecting more than 180,000 women in the United States each year. For women in North America, the life-time odds of getting breast cancer are now one in eight.

No vaccine or other universally successful method for the prevention or
 5 treatment of breast cancer is currently available. Management of the disease currently relies on a combination of early diagnosis (through routine breast screening procedures) and aggressive treatment, which may include one or more of a variety of treatments such as surgery, radiotherapy, chemotherapy and hormone therapy. The course of treatment for a particular breast cancer is often selected based on a variety of prognostic parameters,
 10 including an analysis of specific tumor markers. *See, e.g., Porter-Jordan and Lippman, Breast Cancer 8:73-100 (1994).* However, the use of established markers often leads to a result that is difficult to interpret, and the high mortality observed in breast cancer patients indicates that improvements are needed in the treatment, diagnosis and prevention of the disease.

15 Accordingly, there is a need in the art for improved methods for therapy and diagnosis of breast cancer. The present invention fulfills these needs and further provides other related advantages.

SUMMARY OF THE INVENTION

In one aspect, the present invention provides polynucleotide compositions
 20 comprising a sequence selected from the group consisting of:

(a) sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;

(b) complements of the sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;

25 (c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;

(d) sequences that hybridize to a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, under moderately stringent conditions;

(e) sequences having at least 75% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;

(f) sequences having at least 90% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325; and

(g) degenerate variants of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

10

In one preferred embodiment, the polynucleotide compositions of the invention are expressed in at least about 20%, more preferably in at least about 30%, and most preferably in at least about 50% of breast tumors samples tested, at a level that is at least about 2-fold, preferably at least about 5-fold, and most preferably at least about 10-fold higher than that for normal tissues.

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The present invention, in another aspect, provides polypeptide compositions comprising an amino acid sequence that is encoded by a polynucleotide sequence described above.

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The present invention further provides polypeptide compositions comprising an amino acid sequence selected from the group consisting of sequences recited in SEQ ID NO: 299, 300, 304-306, 308-312, 314 and 326.

25

In certain preferred embodiments, the polypeptides and/or polynucleotides of the present invention are immunogenic, *i.e.*, they are capable of eliciting an immune response, particularly a humoral and/or cellular immune response, as further described herein.

The present invention further provides fragments, variants and/or derivatives of the disclosed polypeptide and/or polynucleotide sequences, wherein the fragments, variants and/or derivatives preferably have a level of immunogenic activity of at least about 50%, preferably at least about 70% and more preferably at least about 90% of the level of

immunogenic activity of a polypeptide sequence set forth in SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326 or a polypeptide sequence encoded by a polynucleotide sequence set forth in SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

5 The present invention further provides polynucleotides that encode a polypeptide described above, expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

 Within other aspects, the present invention provides pharmaceutical compositions comprising a polypeptide or polynucleotide as described above and a
10 physiologically acceptable carrier.

 Within a related aspect of the present invention, the pharmaceutical compositions, *e.g.*, vaccine compositions, are provided for prophylactic or therapeutic applications. Such compositions generally comprise an immunogenic polypeptide or polynucleotide of the invention and an immunostimulant, such as an adjuvant.

15 The present invention further provides pharmaceutical compositions that comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to a polypeptide of the present invention, or a fragment thereof; and (b) a physiologically acceptable carrier.

 Within further aspects, the present invention provides pharmaceutical
20 compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) a pharmaceutically acceptable carrier or excipient. Illustrative antigen presenting cells include dendritic cells, macrophages, monocytes, fibroblasts and B cells.

 Within related aspects, pharmaceutical compositions are provided that
25 comprise: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) an immunostimulant.

 The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins, typically in the form of pharmaceutical compositions, *e.g.*, vaccine

compositions, comprising a physiologically acceptable carrier and/or an immunostimulant. The fusions proteins may comprise multiple immunogenic polypeptides or portions/variants thereof, as described herein, and may further comprise one or more polypeptide segments for facilitating the expression, purification and/or immunogenicity of the polypeptide(s).

Within further aspects, the present invention provides methods for stimulating an immune response in a patient, preferably a T cell response in a human patient, comprising administering a pharmaceutical composition described herein. The patient may be afflicted with breast cancer, in which case the methods provide treatment for the disease, or patient considered at risk for such a disease may be treated prophylactically.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition as recited above. The patient may be afflicted with breast cancer, in which case the methods provide treatment for the disease, or patient considered at risk for such a disease may be treated prophylactically.

The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a polypeptide of the present invention, wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the protein from the sample.

Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating and/or expanding T cells specific for a polypeptide of the present invention, comprising contacting T cells with one or more of: (i) a polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or

expansion of T cells. Isolated T cell populations comprising T cells prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the development of a cancer in a patient, comprising the steps of: (a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of polypeptide disclosed herein; (ii) a polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expressed such a polypeptide; and (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

Within further aspects, the present invention provides methods for determining the presence or absence of a cancer, preferably a breast cancer, in a patient comprising: (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent is an antibody, more preferably a monoclonal antibody.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) detecting in the sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one oligonucleotide primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

Within further aspects, the present invention provides antibodies, such as monoclonal antibodies, that bind to a polypeptide as described above, as well as diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references

disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the differential display PCR products, separated by gel electrophoresis, obtained from cDNA prepared from normal breast tissue (lanes 1 and 2) and from cDNA prepared from breast tumor tissue from the same patient (lanes 3 and 4). The arrow indicates the band corresponding to B18Ag1.

Figure 2 is a northern blot comparing the level of B18Ag1 mRNA in breast tumor tissue (lane 1) with the level in normal breast tissue.

Figure 3 shows the level of B18Ag1 mRNA in breast tumor tissue compared to that in various normal and non-breast tumor tissues as determined by RNase protection assays.

Figure 4 is a genomic clone map showing the location of additional retroviral sequences obtained from ends of XbaI restriction digests (provided in SEQ ID NO:3 - SEQ ID NO:10) relative to B18Ag1.

Figures 5A and 5B show the sequencing strategy, genomic organization and predicted open reading frame for the retroviral element containing B18Ag1.

Figure 6 shows the nucleotide sequence of the representative breast tumor-specific cDNA B18Ag1.

Figure 7 shows the nucleotide sequence of the representative breast tumor-specific cDNA B17Ag1.

Figure 8 shows the nucleotide sequence of the representative breast tumor-specific cDNA B17Ag2.

Figure 9 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag2a.

Figure 10 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag1b.

Figure 11 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag1a.

Figure 12 shows the nucleotide sequence of the representative breast tumor-specific cDNA B11Ag1.

5 Figure 13 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA3c.

Figure 14 shows the nucleotide sequence of the representative breast tumor-specific cDNA B9CG1.

10 Figure 15 shows the nucleotide sequence of the representative breast tumor-specific cDNA B9CG3.

Figure 16 shows the nucleotide sequence of the representative breast tumor-specific cDNA B2CA2.

Figure 17 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA1.

15 Figure 18 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA2.

Figure 19 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA3.

20 Figure 20 shows the nucleotide sequence of the representative breast tumor-specific cDNA B4CA1.

Figure 21A depicts RT-PCR analysis of breast tumor genes in breast tumor tissues (lanes 1-8) and normal breast tissues (lanes 9-13) and H₂O (lane 14).

25 Figure 21B depicts RT-PCR analysis of breast tumor genes in prostate tumors (lane 1, 2), colon tumors (lane 3), lung tumor (lane 4), normal prostate (lane 5), normal colon (lane 6), normal kidney (lane 7), normal liver (lane 8), normal lung (lane 9), normal ovary (lanes 10, 18), normal pancreases (lanes 11, 12), normal skeletal muscle (lane 13), normal skin (lane 14), normal stomach (lane 15), normal testes (lane 16), normal small intestine (lane 17), HBL-100 (lane 19), MCF-12A (lane 20), breast tumors (lanes 21-23), H₂O (lane 24), and colon tumor (lane 25).

Figure 22 shows the recognition of a B11Ag1 peptide (referred to as B11-8) by an anti-B11-8 CTL line.

Figure 23 shows the recognition of a cell line transduced with the antigen B11Ag1 by the B11-8 specific clone A1.

5 Figure 24 shows recognition of a lung adenocarcinoma line (LT-140-22) and a breast adenocarcinoma line (CAMA-1) by the B11-8 specific clone A1.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed generally to compositions and their use in the therapy and diagnosis of cancer, particularly breast cancer. As described further below,
10 illustrative compositions of the present invention include, but are not restricted to, polypeptides, particularly immunogenic polypeptides, polynucleotides encoding such polypeptides, antibodies and other binding agents, antigen presenting cells (APCs) and immune system cells (*e.g.*, T cells).

The practice of the present invention will employ, unless indicated
15 specifically to the contrary, conventional methods of virology, immunology, microbiology, molecular biology and recombinant DNA techniques within the skill of the art, many of which are described below for the purpose of illustration. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, et al. *Molecular Cloning: A Laboratory Manual* (2nd Edition, 1989); Maniatis et al. *Molecular Cloning: A Laboratory Manual* (1982);
20 *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., 1984); *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds., 1985); *Transcription and Translation* (B. Hames & S. Higgins, eds., 1984); *Animal Cell Culture* (R. Freshney, ed., 1986); Perbal, *A Practical Guide to Molecular Cloning* (1984).

All publications, patents and patent applications cited herein, whether supra
25 or infra, are hereby incorporated by reference in their entirety.

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates otherwise.

Polypeptide Compositions

As used herein, the term "polypeptide" " is used in its conventional meaning, *i.e.*, as a sequence of amino acids. The polypeptides are not limited to a specific length of the product; thus, peptides, oligopeptides, and proteins are included within the definition of polypeptide, and such terms may be used interchangeably herein unless specifically indicated otherwise. This term also does not refer to or exclude post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like, as well as other modifications known in the art, both naturally occurring and non-naturally occurring. A polypeptide may be an entire protein, or a subsequence thereof. Particular polypeptides of interest in the context of this invention are amino acid subsequences comprising epitopes, *i.e.*, antigenic determinants substantially responsible for the immunogenic properties of a polypeptide and being capable of evoking an immune response.

Particularly illustrative polypeptides of the present invention comprise those encoded by a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, or a sequence that hybridizes under moderately stringent conditions, or, alternatively, under highly stringent conditions, to a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325. Certain other illustrative polypeptides of the invention comprise amino acid sequences as set forth in any one of SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326.

The polypeptides of the present invention are sometimes herein referred to as breast tumor proteins or breast tumor polypeptides, as an indication that their identification has been based at least in part upon their increased levels of expression in breast tumor samples. Thus, a "breast tumor polypeptide" or "breast tumor protein," refers generally to a polypeptide sequence of the present invention, or a polynucleotide sequence encoding such a polypeptide, that is expressed in a substantial proportion of breast tumor samples, for example preferably greater than about 20%, more preferably greater than about 30%, and most preferably greater than about 50% or more of breast

tumor samples tested, at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in normal tissues, as determined using a representative assay provided herein. A breast tumor polypeptide sequence of the invention, based upon its increased level of expression in tumor cells, has particular utility both as a diagnostic marker as well as a therapeutic target, as further described below.

In certain preferred embodiments, the polypeptides of the invention are immunogenic, *i.e.*, they react detectably within an immunoassay (such as an ELISA or T-cell stimulation assay) with antisera and/or T-cells from a patient with breast cancer. Screening for immunogenic activity can be performed using techniques well known to the skilled artisan. For example, such screens can be performed using methods such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In one illustrative example, a polypeptide may be immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies detected using, for example, ¹²⁵I-labeled Protein A.

As would be recognized by the skilled artisan, immunogenic portions of the polypeptides disclosed herein are also encompassed by the present invention. An "immunogenic portion," as used herein, is a fragment of an immunogenic polypeptide of the invention that itself is immunologically reactive (*i.e.*, specifically binds) with the B-cells and/or T-cell surface antigen receptors that recognize the polypeptide. Immunogenic portions may generally be identified using well known techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used herein, antisera and antibodies are "antigen-specific" if they specifically bind to an antigen (*i.e.*, they react with the protein in an ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well-known techniques.

In one preferred embodiment, an immunogenic portion of a polypeptide of the present invention is a portion that reacts with antisera and/or T-cells at a level that is not substantially less than the reactivity of the full-length polypeptide (*e.g.*, in an ELISA and/or T-cell reactivity assay). Preferably, the level of immunogenic activity of the immunogenic portion is at least about 50%, preferably at least about 70% and most preferably greater than about 90% of the immunogenicity for the full-length polypeptide. In some instances, preferred immunogenic portions will be identified that have a level of immunogenic activity greater than that of the corresponding full-length polypeptide, *e.g.*, having greater than about 100% or 150% or more immunogenic activity.

10 In certain other embodiments, illustrative immunogenic portions may include peptides in which an N-terminal leader sequence and/or transmembrane domain have been deleted. Other illustrative immunogenic portions will contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

15 In another embodiment, a polypeptide composition of the invention may also comprise one or more polypeptides that are immunologically reactive with T cells and/or antibodies generated against a polypeptide of the invention, particularly a polypeptide having an amino acid sequence disclosed herein, or to an immunogenic fragment or variant thereof.

20 In another embodiment of the invention, polypeptides are provided that comprise one or more polypeptides that are capable of eliciting T cells and/or antibodies that are immunologically reactive with one or more polypeptides described herein, or one or more polypeptides encoded by contiguous nucleic acid sequences contained in the polynucleotide sequences disclosed herein, or immunogenic fragments or variants thereof,
25 or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency.

The present invention, in another aspect, provides polypeptide fragments comprising at least about 5, 10, 15, 20, 25, 50, or 100 contiguous amino acids, or more, including all intermediate lengths, of a polypeptide compositions set forth herein, such as

those set forth in SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326, or those encoded by a polynucleotide sequence set forth in a sequence of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

In another aspect, the present invention provides variants of the polypeptide compositions described herein. Polypeptide variants generally encompassed by the present invention will typically exhibit at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% or more identity (determined as described below), along its length, to a polypeptide sequences set forth herein.

In one preferred embodiment, the polypeptide fragments and variants provide by the present invention are immunologically reactive with an antibody and/or T-cell that reacts with a full-length polypeptide specifically set forth herein.

In another preferred embodiment, the polypeptide fragments and variants provided by the present invention exhibit a level of immunogenic activity of at least about 50%, preferably at least about 70%, and most preferably at least about 90% or more of that exhibited by a full-length polypeptide sequence specifically set forth herein.

A polypeptide "variant," as the term is used herein, is a polypeptide that typically differs from a polypeptide specifically disclosed herein in one or more substitutions, deletions, additions and/or insertions. Such variants may be naturally occurring or may be synthetically generated, for example, by modifying one or more of the above polypeptide sequences of the invention and evaluating their immunogenic activity as described herein and/or using any of a number of techniques well known in the art.

For example, certain illustrative variants of the polypeptides of the invention include those in which one or more portions, such as an N-terminal leader sequence or transmembrane domain, have been removed. Other illustrative variants include variants in which a small portion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

In many instances, a variant will contain conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would

expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. As described above, modifications may be made in the structure of the polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics, *e.g.*, with immunogenic characteristics. When it is desired to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, immunogenic variant or portion of a polypeptide of the invention, one skilled in the art will typically change one or more of the codons of the encoding DNA sequence according to Table 1.

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

TABLE 1

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes,

substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8);
 5 glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6); histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with
 10 similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101
 15 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0);
 20 threonine (−0.4); proline (−0.5 \pm 1); alanine (−0.5); histidine (−0.5); cysteine (−1.0); methionine (−1.3); valine (−1.5); leucine (−1.8); isoleucine (−1.8); tyrosine (−2.3); phenylalanine (−2.5); tryptophan (−3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically
 25 equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-, methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

Amino acid substitutions may further be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer. Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure and hydropathic nature of the polypeptide.

As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other

sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

When comparing polypeptide sequences, two sequences are said to be
 5 “identical” if the sequence of amino acids in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A “comparison window” as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to
 10 about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment
 15 schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183,
 20 Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J.
 25 (1983) *Proc. Natl. Acad., Sci. USA* 80:726-730.

Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl.*

Acad. Sci. USA 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining
 5 percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides and polypeptides of the invention. Software for
 10 performing BLAST analyses is publicly available through the National Center for Biotechnology Information. For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one
 15 or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment.

In one preferred approach, the “percentage of sequence identity” is determined by comparing two optimally aligned sequences over a window of comparison
 20 of at least 20 positions, wherein the portion of the polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical
 25 amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Within other illustrative embodiments, a polypeptide may be a fusion polypeptide that comprises multiple polypeptides as described herein, or that comprises at least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the polypeptide or to enable the polypeptide to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the polypeptide.

Fusion polypeptides may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion polypeptide is expressed as a recombinant polypeptide, allowing the production of increased levels, relative to a non-fused polypeptide, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion polypeptide that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion polypeptide using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes.

Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea et al., *Gene* 40:39-46, 1985; Murphy et al., *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

The ligated DNA sequences are operably linked to suitable transcriptional or translational regulatory elements. The regulatory elements responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the second polypeptide.

The fusion polypeptide can comprise a polypeptide as described herein together with an unrelated immunogenic protein, such as an immunogenic protein capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (*see*, for example, Stoute et al. *New Engl. J. Med.*, 336:86-91, 1997).

In one preferred embodiment, the immunological fusion partner is derived from a *Mycobacterium* sp., such as a *Mycobacterium tuberculosis*-derived Ra12 fragment. Ra12 compositions and methods for their use in enhancing the expression and/or immunogenicity of heterologous polynucleotide/polypeptide sequences is described in U.S. Patent Application 60/158,585, the disclosure of which is incorporated herein by reference in its entirety. Briefly, Ra12 refers to a polynucleotide region that is a subsequence of a *Mycobacterium tuberculosis* MTB32A nucleic acid. MTB32A is a serine protease of 32 KD molecular weight encoded by a gene in virulent and avirulent strains of *M. tuberculosis*. The nucleotide sequence and amino acid sequence of MTB32A have been described (for example, U.S. Patent Application 60/158,585; *see also*, Skeiky et al., *Infection and Immun.* (1999) 67:3998-4007, incorporated herein by reference). C-terminal fragments of the MTB32A coding sequence express at high levels and remain as a soluble

polypeptides throughout the purification process. Moreover, Ra12 may enhance the immunogenicity of heterologous immunogenic polypeptides with which it is fused. One preferred Ra12 fusion polypeptide comprises a 14 KD C-terminal fragment corresponding to amino acid residues 192 to 323 of MTB32A. Other preferred Ra12 polynucleotides

5 generally comprise at least about 15 consecutive nucleotides, at least about 30 nucleotides, at least about 60 nucleotides, at least about 100 nucleotides, at least about 200 nucleotides, or at least about 300 nucleotides that encode a portion of a Ra12 polypeptide. Ra12 polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a Ra12 polypeptide or a portion thereof) or may comprise a variant of such a

10 sequence. Ra12 polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions such that the biological activity of the encoded fusion polypeptide is not substantially diminished, relative to a fusion polypeptide comprising a native Ra12 polypeptide. Variants preferably exhibit at least about 70% identity, more preferably at least about 80% identity and most preferably at least about 90% identity to a

15 polynucleotide sequence that encodes a native Ra12 polypeptide or a portion thereof.

Within other preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza B* (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (*e.g.*, the first N-terminal 100-110 amino acids), and a protein D

20 derivative may be lipidated. Within certain preferred embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells. Other fusion partners include the non-structural

25 protein from influenzae virus, NS1 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known

as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This property has been exploited for the development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (*see Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of LYTA may be incorporated into a fusion polypeptide. A repeat portion is found in the C-terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

Yet another illustrative embodiment involves fusion polypeptides, and the polynucleotides encoding them, wherein the fusion partner comprises a targeting signal capable of directing a polypeptide to the endosomal/lysosomal compartment, as described in U.S. Patent No. 5,633,234. An immunogenic polypeptide of the invention, when fused with this targeting signal, will associate more efficiently with MHC class II molecules and thereby provide enhanced in vivo stimulation of CD4⁺ T-cells specific for the polypeptide.

Polypeptides of the invention are prepared using any of a variety of well known synthetic and/or recombinant techniques, the latter of which are further described below. Polypeptides, portions and other variants generally less than about 150 amino acids can be generated by synthetic means, using techniques well known to those of ordinary skill in the art. In one illustrative example, such polypeptides are synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. *See Merrifield, J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

In general, polypeptide compositions (including fusion polypeptides) of the invention are isolated. An "isolated" polypeptide is one that is removed from its original

environment. For example, a naturally-occurring protein or polypeptide is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are also purified, *e.g.*, are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure.

5 Polynucleotide Compositions

The present invention, in other aspects, provides polynucleotide compositions. The terms "DNA" and "polynucleotide" are used essentially interchangeably herein to refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. "Isolated," as used herein, means that a
 10 polynucleotide is substantially away from other coding sequences, and that the DNA molecule does not contain large portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA molecule as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

As will be understood by those skilled in the art, the polynucleotide
 15 compositions of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

As will be also recognized by the skilled artisan, polynucleotides of the
 20 invention may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules may include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or
 25 non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

Polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a polypeptide/protein of the invention or a portion thereof) or may comprise a sequence that encodes a variant or derivative, preferably and immunogenic variant or derivative, of such a sequence.

5 Therefore, according to another aspect of the present invention, polynucleotide compositions are provided that comprise some or all of a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, complements of a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, and degenerate
10 variants of a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325. In certain preferred embodiments, the polynucleotide sequences set forth herein encode immunogenic polypeptides, as described above.

In other related embodiments, the present invention provides polynucleotide
15 variants having substantial identity to the sequences disclosed herein in SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, for example those comprising at least 70% sequence identity, preferably at least 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence identity compared to a polynucleotide sequence of this invention using the methods described herein, (*e.g.*, BLAST analysis using standard
20 parameters, as described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like.

Typically, polynucleotide variants will contain one or more substitutions,
25 additions, deletions and/or insertions, preferably such that the immunogenicity of the polypeptide encoded by the variant polynucleotide is not substantially diminished relative to a polypeptide encoded by a polynucleotide sequence specifically set forth herein). The term “variants” should also be understood to encompass homologous genes of xenogenic origin.

In additional embodiments, the present invention provides polynucleotide fragments comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at least about 10, 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 16, 17, 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers through 200-500; 500-1,000, and the like.

In another embodiment of the invention, polynucleotide compositions are provided that are capable of hybridizing under moderate to high stringency conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50°C-60°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS. One skilled in the art will understand that the stringency of hybridization can be readily manipulated, such as by altering the salt content of the hybridization solution and/or the temperature at which the hybridization is performed. For example, in another embodiment, suitable highly stringent hybridization conditions include those described above, with the exception that the temperature of hybridization is increased, *e.g.*, to 60-65°C or 65-70°C.

In certain preferred embodiments, the polynucleotides described above, *e.g.*, polynucleotide variants, fragments and hybridizing sequences, encode polypeptides that are immunologically cross-reactive with a polypeptide sequence specifically set forth herein. In other preferred embodiments, such polynucleotides encode polypeptides that have a level of immunogenic activity of at least about 50%, preferably at least about 70%, and

more preferably at least about 90% of that for a polypeptide sequence specifically set forth herein.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, illustrative polynucleotide segments with total lengths of about 10,000, about 5000, about 3000, about 2,000, about 1,000, about 500, about 200, about 100, about 50 base pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many implementations of this invention.

When comparing polynucleotide sequences, two sequences are said to be “identical” if the sequence of nucleotides in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A “comparison window” as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183,

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 5 *Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J. (1983) *Proc. Natl. Acad., Sci. USA* 80:726-730.

Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.*
 10 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining
 15 percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides of the invention. Software for performing
 20 BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the
 25 quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62

scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments, (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

Preferably, the “percentage of sequence identity” is determined by comparing two optimally aligned sequences over a window of comparison of at least 20
 5 positions, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical
 10 nucleic acid bases occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

It will be appreciated by those of ordinary skill in the art that, as a result of
 15 the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the
 20 scope of the present invention. Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

25 Therefore, in another embodiment of the invention, a mutagenesis approach, such as site-specific mutagenesis, is employed for the preparation of immunogenic variants and/or derivatives of the polypeptides described herein. By this approach, specific modifications in a polypeptide sequence can be made through mutagenesis of the underlying polynucleotides that encode them. These techniques provides a straightforward

approach to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the polynucleotide.

Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Mutations may be employed in a selected polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or more properties of the encoded polypeptide, such as the immunogenicity of a polypeptide vaccine. The techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example, site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so in length is employed, with about 5 to about 10 residues on both sides of the junction of the sequence being altered.

As will be appreciated by those of skill in the art, site-specific mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially-available and their use is generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double-stranded

vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to
 5 complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

10 The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic
 15 agents, such as hydroxylamine, to obtain sequence variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

As used herein, the term “oligonucleotide directed mutagenesis procedure”
 20 refers to template-dependent processes and vector-mediated propagation which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term “oligonucleotide directed mutagenesis procedure” is intended to refer to a process that involves the template-dependent extension of a primer
 25 molecule. The term template dependent process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987). Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the

recovery of the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

In another approach for the production of polypeptide variants of the present invention, recursive sequence recombination, as described in U.S. Patent No. 5,837,458, may be employed. In this approach, iterative cycles of recombination and screening or selection are performed to “evolve” individual polynucleotide variants of the invention having, for example, enhanced immunogenic activity.

In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic acid hybridization. As such, it is contemplated that nucleic acid segments that comprise a sequence region of at least about 15 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000 (including all intermediate lengths) and even up to full length sequences will also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned, such as the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately

depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the
 5 length complementary sequences one wishes to detect.

The use of a hybridization probe of about 15-25 nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 15 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and
 10 thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 25 contiguous nucleotides, or even longer where desired.

Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequences set forth herein,
 15 or to any continuous portion of the sequences, from about 15-25 nucleotides in length up to and including the full length sequence, that one wishes to utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

Small polynucleotide segments or fragments may be readily prepared by, for
 20 example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA
 25 techniques generally known to those of skill in the art of molecular biology.

The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of

probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about
 5 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating related sequences.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template, less
 10 stringent (reduced stringency) hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any
 15 case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

20 According to another embodiment of the present invention, polynucleotide compositions comprising antisense oligonucleotides are provided. Antisense oligonucleotides have been demonstrated to be effective and targeted inhibitors of protein synthesis, and, consequently, provide a therapeutic approach by which a disease can be treated by inhibiting the synthesis of proteins that contribute to the disease. The efficacy of
 25 antisense oligonucleotides for inhibiting protein synthesis is well established. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829). Further, examples of antisense inhibition have been demonstrated with the nuclear protein cyclin, the multiple drug resistance gene

(MDG1), ICAM-1, E-selectin, STK-1, striatal GABA_A receptor and human EGF (Jaskulski *et al.*, Science. 1988 Jun 10;240(4858):1544-6; Vasanthakumar and Ahmed, Cancer Commun. 1989;1(4):225-32; Peris *et al.*, Brain Res Mol Brain Res. 1998 Jun 15;57(2):310-20; U. S. Patent 5,801,154; U.S. Patent 5,789,573; U. S. Patent 5,718,709 and U.S. Patent 5,610,288). Antisense constructs have also been described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683).

Therefore, in certain embodiments, the present invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary, and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein. Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence and determination of secondary structure, T_m , binding energy, and relative stability. Antisense compositions may be selected based upon their relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell. Highly preferred target regions of the mRNA, are those which are at or near the AUG translation initiation codon, and those sequences which are substantially complementary to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations can be performed, for example, using v.4 of the OLIGO primer analysis software and/or the BLASTN 2.0.5 algorithm software (Altschul *et al.*, Nucleic Acids Res. 1997, 25(17):3389-402).

The use of an antisense delivery method employing a short peptide vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, Nucleic Acids Res. 1997 Jul 15;25(14):2730-6). It has been demonstrated that several molecules of the MPG peptide coat the antisense oligonucleotides and can be delivered into cultured mammalian cells in less than 1 hour with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane.

According to another embodiment of the invention, the polynucleotide compositions described herein are used in the design and preparation of ribozyme molecules for inhibiting expression of the tumor polypeptides and proteins of the present invention in tumor cells. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, Proc Natl Acad Sci U S A. 1987 Dec;84(24):8788-92; Forster and Symons, Cell. 1987 Apr 24;49(2):211-20). For example, a large number of ribozymes accelerate phosphoester transfer reactions with a high degree of specificity, often cleaving only one of several phosphoesters in an oligonucleotide substrate (Cech *et al.*, Cell. 1981 Dec;27(3 Pt 2):487-96; Michel and Westhof, J Mol Biol. 1990 Dec 5;216(3):585-610; Reinhold-Hurek and Shub, Nature. 1992 May 14;357(6374):173-6). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide sequence ("IGS") of the ribozyme prior to chemical reaction.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general, enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic

nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it
 5 is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme necessary
 10 to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding to the target RNA, but also on the mechanism of target RNA
 15 cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woolf *et al.*, Proc Natl Acad Sci U S A. 1992 Aug 15;89(16):7305-9). Thus, the specificity of action of a ribozyme is greater than that of an antisense oligonucleotide binding the same RNA site.

20 The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis δ virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* Nucleic Acids Res. 1992 Sep 11;20(17):4559-65. Examples of hairpin motifs are described by Hampel *et al.* (Eur. Pat. Appl. Publ. No. EP 0360257),
 25 Hampel and Tritz, Biochemistry 1989 Jun 13;28(12):4929-33; Hampel *et al.*, Nucleic Acids Res. 1990 Jan 25;18(2):299-304 and U. S. Patent 5,631,359. An example of the hepatitis δ virus motif is described by Perrotta and Been, Biochemistry. 1992 Dec 1;31(47):11843-52; an example of the RNaseP motif is described by Guerrier-Takada *et al.*, Cell. 1983 Dec;35(3 Pt 2):849-57; Neurospora VS RNA ribozyme motif is described by

Collins (Saville and Collins, Cell. 1990 May 18;61(4):685-96; Saville and Collins, Proc Natl Acad Sci U S A. 1991 Oct 1;88(19):8826-30; Collins and Olive, Biochemistry. 1993 Mar 23;32(11):2795-9); and an example of the Group I intron is described in (U. S. Patent 4,987,071). All that is important in an enzymatic nucleic acid molecule of this invention is
 5 that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO
 10 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

15 Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see *e.g.*, Int. Pat. Appl. Publ. No. WO 92/07065; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688,
 20 which describe various chemical modifications that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the
 25 general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex*

vivo to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical, systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

Another means of accumulating high concentrations of a ribozyme(s) within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells. Ribozymes expressed from such promoters have been shown to function in mammalian cells. Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

In another embodiment of the invention, peptide nucleic acids (PNAs) compositions are provided. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, *Antisense Nucleic Acid Drug Dev.* 1997 7(4) 431-37). PNA is able to be utilized in a number methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided

by Corey (*Trends Biotechnol* 1997 Jun;15(6):224-9). As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-specific mRNA, and thereby alter the level of ACE activity in a host cell to which such PNA compositions have been administered.

PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, *Science* 1991 Dec 6;254(5037):1497-500; Hanvey *et al.*, *Science*. 1992 Nov 27;258(5087):1481-5; Hyrup and Nielsen, *Bioorg Med Chem*. 1996 Jan;4(1):5-23). This chemistry has three important consequences: firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral molecules; secondly, PNAs are achiral, which avoids the need to develop a stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc or Fmoc protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used.

PNA monomers or ready-made oligomers are commercially available from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, *Bioorg Med Chem*. 1995 Apr;3(4):437-45). The manual protocol lends itself to the production of chemically modified PNAs or the simultaneous synthesis of families of closely related PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this difficulty, it is suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography, providing yields and purity of product similar to those observed during the synthesis of peptides.

Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The ease with which PNAs can be modified facilitates optimization for better solubility or for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (for example, Norton *et al.*, *Bioorg Med Chem.* 1995 Apr;3(4):437-45; Petersen *et al.*, *J Pept Sci.* 1995 May-Jun;1(3):175-83; Orum *et al.*, *Biotechniques.* 1995 Sep;19(3):472-80; Footer *et al.*, *Biochemistry.* 1996 Aug 20;35(33):10673-9; Griffith *et al.*, *Nucleic Acids Res.* 1995 Aug 11;23(15):3003-8; Pardridge *et al.*, *Proc Natl Acad Sci U S A.* 1995 Jun 6;92(12):5592-6; Boffa *et al.*, *Proc Natl Acad Sci U S A.* 1995 Mar 14;92(6):1901-5; Gambacorti-Passerini *et al.*, *Blood.* 1996 Aug 15;88(4):1411-7; Armitage *et al.*, *Proc Natl Acad Sci U S A.* 1997 Nov 11;94(23):12320-5; Seeger *et al.*, *Biotechniques.* 1997 Sep;23(3):512-7). U.S. Patent No. 5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to therapeutics.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (*Anal Chem.* 1993 Dec 15;65(24):3545-9) and Jensen *et al.* (1997 Apr 22;36(16):5072-7). Rose uses capillary gel electrophoresis to determine binding of PNAs to their complementary oligonucleotide, measuring the relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BIAcore™ technology.

Other applications of PNAs that have been described and will be apparent to the skilled artisan include use in DNA strand invasion, antisense inhibition, mutational analysis, enhancers of transcription, nucleic acid purification, isolation of transcriptionally active genes, blocking of transcription factor binding, genome cleavage, biosensors, *in situ* hybridization, and the like.

Polynucleotide Identification, Characterization and Expression

Polynucleotides compositions of the present invention may be identified, prepared and/or manipulated using any of a variety of well established techniques (see generally, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989, and other like references). For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for tumor-associated expression (*i.e.*, expression that is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using the microarray technology of Affymetrix, Inc. (Santa Clara, CA) according to the manufacturer's instructions (and essentially as described by Schena et al., *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller et al., *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as tumor cells.

Many template dependent processes are available to amplify a target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCRTM) which is described in detail in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCRTM, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCRTM amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Any of a number of other template dependent processes, many of which are variations of the PCR[™] amplification technique, are readily known and available in the art. Illustratively, some such methods include the ligase chain reaction (referred to as LCR), described, for example, in Eur. Pat. Appl. Publ. No. 320,308 and U.S. Patent No. 4,883,750; Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880; Strand Displacement Amplification (SDA) and Repair Chain Reaction (RCR). Still other amplification methods are described in Great Britain Pat. Appl. No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025. Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (PCT Intl. Pat. Appl. Publ. No. WO 88/10315), including nucleic acid sequence based amplification (NASBA) and 3SR. Eur. Pat. Appl. Publ. No. 329,822 describes a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA). PCT Intl. Pat. Appl. Publ. No. WO 89/06700 describes a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. Other amplification methods such as "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) are also well-known to those of skill in the art.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (*e.g.*, a tumor cDNA library) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (*e.g.*, by nick-translation or end-labeling with ³²P) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (*see* Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor

Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences
 5 may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then be assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

10 Alternatively, amplification techniques, such as those described above, can be useful for obtaining a full length coding sequence from a partial cDNA sequence. One such amplification technique is inverse PCR (*see* Triglia et al., *Nucl. Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template
 15 for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known region. A variation on this procedure,
 20 which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Additional techniques include
 25 capture PCR (Lagerstrom et al., *PCR Methods Applic.* 1:111-19, 1991) and walking PCR (Parker et al., *Nucl. Acids. Res.* 19:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that

available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (*e.g.*, NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences may also be obtained by analysis of genomic fragments.

5 In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally
10 equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

 As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or
15 eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

 Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding
20 sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation
25 patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

 In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it

may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the heterologous moiety.

5 Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. et al. (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

15 A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (*e.g.*, Creighton, T. (1983) *Proteins, Structures and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (*e.g.*, the Edman degradation procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

20 In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into appropriate expression vector, *i.e.*, a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Such techniques are described, for example, in Sambrook, J. et al. (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview,

N.Y., and Ausubel, F. M. et al. (1989) Current Protocols in Molecular Biology, John Wiley & Sons, New York. N.Y.

A variety of expression vector/host systems may be utilized to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms
 5 such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (*e.g.*, baculovirus); plant cell systems transformed with virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmids); or animal cell
 10 systems.

The "control elements" or "regulatory sequences" present in an expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the
 15 vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or PSPORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from
 20 mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV may be advantageously used with an appropriate selectable marker.

In bacterial systems, any of a number of expression vectors may be selected
 25 depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used. Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of

interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of .beta.-galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign
 5 polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST
 10 moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel et al. (supra) and Grant et al. (1987) *Methods Enzymol.* 153:516-544.

15 In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of RUBISCO
 20 or heat shock promoters may be used (Coruzzi, G. et al. (1984) *EMBO J.* 3:1671-1680; Broglie, R. et al. (1984) *Science* 224:838-843; and Winter, J. et al. (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, for example, Hobbs, S. or Murry, L. E. in
 25 McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in

Trichoplusia larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein.

- 5 The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or *Trichoplusia* larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. et al. (1994) *Proc. Natl. Acad. Sci.* 91 :3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression
10 vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition,
15 transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the
20 polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure
25 translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, COS, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. et al. (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase (Lowy, I. et al. (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or aprt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. et al. (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. et al (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been

described, for example, *trpB*, which allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). The use of visible markers has gained popularity with such markers as anthocyanins, beta-glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. et al. (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells that contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques which include, for example, membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies specific for the product are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton, R. et al. (1990; Serological

Methods, a Laboratory Manual, APS Press, St Paul. Minn.) and Maddox, D. E. et al. (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for
 5 producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by
 10 addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

15 Host cells transformed with a polynucleotide sequence of interest may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of the invention may
 20 be designed to contain signal sequences which direct secretion of the encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides
 25 such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen. San Diego, Calif.) between the purification domain and the

encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMLAC (immobilized metal ion affinity chromatography) as described in Porath, J. et al. (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase cleavage site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. et al. (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

Antibody Compositions, Fragments Thereof and Other Binding Agents

According to another aspect, the present invention further provides binding agents, such as antibodies and antigen-binding fragments thereof, that exhibit immunological binding to a tumor polypeptide disclosed herein, or to a portion, variant or derivative thereof. An antibody, or antigen-binding fragment thereof, is said to "specifically bind," "immunologically bind," and/or is "immunologically reactive" to a polypeptide of the invention if it reacts at a detectable level (within, for example, an ELISA assay) with the polypeptide, and does not react detectably with unrelated polypeptides under similar conditions.

Immunological binding, as used in this context, generally refers to the non-covalent interactions of the type which occur between an immunoglobulin molecule and an antigen for which the immunoglobulin is specific. The strength, or affinity of immunological binding interactions can be expressed in terms of the dissociation constant

(K_d) of the interaction, wherein a smaller K_d represents a greater affinity. Immunological binding properties of selected polypeptides can be quantified using methods well known in the art. One such method entails measuring the rates of antigen-binding site/antigen complex formation and dissociation, wherein those rates depend on the concentrations of the complex partners, the affinity of the interaction, and on geometric parameters that

5 the complex partners, the affinity of the interaction, and on geometric parameters that equally influence the rate in both directions. Thus, both the "on rate constant" (K_{on}) and the "off rate constant" (K_{off}) can be determined by calculation of the concentrations and the actual rates of association and dissociation. The ratio of K_{off}/K_{on} enables cancellation of all parameters not related to affinity, and is thus equal to the dissociation constant K_d . See,

10 generally, Davies et al. (1990) Annual Rev. Biochem. 59:439-473.

An "antigen-binding site," or "binding portion" of an antibody refers to the part of the immunoglobulin molecule that participates in antigen binding. The antigen binding site is formed by amino acid residues of the N-terminal variable ("V") regions of the heavy ("H") and light ("L") chains. Three highly divergent stretches within the V

15 regions of the heavy and light chains are referred to as "hypervariable regions" which are interposed between more conserved flanking stretches known as "framework regions," or "FRs". Thus the term "FR" refers to amino acid sequences which are naturally found between and adjacent to hypervariable regions in immunoglobulins. In an antibody molecule, the three hypervariable regions of a light chain and the three hypervariable

20 regions of a heavy chain are disposed relative to each other in three dimensional space to form an antigen-binding surface. The antigen-binding surface is complementary to the three-dimensional surface of a bound antigen, and the three hypervariable regions of each of the heavy and light chains are referred to as "complementarity-determining regions," or "CDRs."

25 Binding agents may be further capable of differentiating between patients with and without a cancer, such as breast cancer, using the representative assays provided herein. For example, antibodies or other binding agents that bind to a tumor protein will preferably generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, more preferably at least about 30% of patients. Alternatively, or

in addition, the antibody will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the cancer. To determine whether a binding agent satisfies this requirement, biological samples (*e.g.*, blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. Preferably, a statistically significant number of samples with and without the disease will be assayed. Each binding agent should satisfy the above criteria; however, those of ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

Any agent that satisfies the above requirements may be a binding agent. For example, a binding agent may be a ribosome, with or without a peptide component, an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. *See, e.g.*, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, antibodies can be produced by cell culture techniques, including the generation of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (*e.g.*, mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J. Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the desired specificity (*i.e.*, reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

A number of therapeutically useful molecules are known in the art which comprise antigen-binding sites that are capable of exhibiting immunological binding properties of an antibody molecule. The proteolytic enzyme papain preferentially cleaves IgG molecules to yield several fragments, two of which (the "F(ab)" fragments) each comprise a covalent heterodimer that includes an intact antigen-binding site. The enzyme pepsin is able to cleave IgG molecules to provide several fragments, including the "F(ab')₂"

fragment which comprises both antigen-binding sites. An "Fv" fragment can be produced by preferential proteolytic cleavage of an IgM, and on rare occasions IgG or IgA immunoglobulin molecule. Fv fragments are, however, more commonly derived using recombinant techniques known in the art. The Fv fragment includes a non-covalent $V_H::V_L$ heterodimer including an antigen-binding site which retains much of the antigen recognition and binding capabilities of the native antibody molecule. Inbar et al. (1972) Proc. Nat. Acad. Sci. USA 69:2659-2662; Hochman et al. (1976) Biochem 15:2706-2710; and Ehrlich et al. (1980) Biochem 19:4091-4096.

A single chain Fv ("sFv") polypeptide is a covalently linked $V_H::V_L$ heterodimer which is expressed from a gene fusion including V_H - and V_L -encoding genes linked by a peptide-encoding linker. Huston et al. (1988) Proc. Nat. Acad. Sci. USA 85(16):5879-5883. A number of methods have been described to discern chemical structures for converting the naturally aggregated--but chemically separated--light and heavy polypeptide chains from an antibody V region into an sFv molecule which will fold into a three dimensional structure substantially similar to the structure of an antigen-binding site. See, *e.g.*, U.S. Pat. Nos. 5,091,513 and 5,132,405, to Huston et al.; and U.S. Pat. No. 4,946,778, to Ladner et al.

Each of the above-described molecules includes a heavy chain and a light chain CDR set, respectively interposed between a heavy chain and a light chain FR set which provide support to the CDRs and define the spatial relationship of the CDRs relative to each other. As used herein, the term "CDR set" refers to the three hypervariable regions of a heavy or light chain V region. Proceeding from the N-terminus of a heavy or light chain, these regions are denoted as "CDR1," "CDR2," and "CDR3" respectively. An antigen-binding site, therefore, includes six CDRs, comprising the CDR set from each of a heavy and a light chain V region. A polypeptide comprising a single CDR, (*e.g.*, a CDR1, CDR2 or CDR3) is referred to herein as a "molecular recognition unit." Crystallographic analysis of a number of antigen-antibody complexes has demonstrated that the amino acid residues of CDRs form extensive contact with bound antigen, wherein the most extensive

antigen contact is with the heavy chain CDR3. Thus, the molecular recognition units are primarily responsible for the specificity of an antigen-binding site.

As used herein, the term "FR set" refers to the four flanking amino acid sequences which frame the CDRs of a CDR set of a heavy or light chain V region. Some FR residues may contact bound antigen; however, FRs are primarily responsible for folding the V region into the antigen-binding site, particularly the FR residues directly adjacent to the CDRs. Within FRs, certain amino residues and certain structural features are very highly conserved. In this regard, all V region sequences contain an internal disulfide loop of around 90 amino acid residues. When the V regions fold into a binding-site, the CDRs are displayed as projecting loop motifs which form an antigen-binding surface. It is generally recognized that there are conserved structural regions of FRs which influence the folded shape of the CDR loops into certain "canonical" structures--regardless of the precise CDR amino acid sequence. Further, certain FR residues are known to participate in non-covalent interdomain contacts which stabilize the interaction of the antibody heavy and light chains.

A number of "humanized" antibody molecules comprising an antigen-binding site derived from a non-human immunoglobulin have been described, including chimeric antibodies having rodent V regions and their associated CDRs fused to human constant domains (Winter et al. (1991) *Nature* 349:293-299; Lobuglio et al. (1989) *Proc. Nat. Acad. Sci. USA* 86:4220-4224; Shaw et al. (1987) *J Immunol.* 138:4534-4538; and Brown et al. (1987) *Cancer Res.* 47:3577-3583), rodent CDRs grafted into a human supporting FR prior to fusion with an appropriate human antibody constant domain (Riechmann et al. (1988) *Nature* 332:323-327; Verhoeyen et al. (1988) *Science* 239:1534-1536; and Jones et al. (1986) *Nature* 321:522-525), and rodent CDRs supported by recombinantly veneered rodent FRs (European Patent Publication No. 519,596, published Dec. 23, 1992). These "humanized" molecules are designed to minimize unwanted immunological response toward rodent antihuman antibody molecules which limits the duration and effectiveness of therapeutic applications of those moieties in human recipients.

As used herein, the terms "veneered FRs" and "recombinantly veneered FRs" refer to the selective replacement of FR residues from, *e.g.*, a rodent heavy or light chain V region, with human FR residues in order to provide a xenogeneic molecule comprising an antigen-binding site which retains substantially all of the native FR polypeptide folding structure. Veneering techniques are based on the understanding that the ligand binding characteristics of an antigen-binding site are determined primarily by the structure and relative disposition of the heavy and light chain CDR sets within the antigen-binding surface. Davies et al. (1990) Ann. Rev. Biochem. 59:439-473. Thus, antigen binding specificity can be preserved in a humanized antibody only wherein the CDR structures, their interaction with each other, and their interaction with the rest of the V region domains are carefully maintained. By using veneering techniques, exterior (*e.g.*, solvent-accessible) FR residues which are readily encountered by the immune system are selectively replaced with human residues to provide a hybrid molecule that comprises either a weakly immunogenic, or substantially non-immunogenic veneered surface.

The process of veneering makes use of the available sequence data for human antibody variable domains compiled by Kabat et al., in Sequences of Proteins of Immunological Interest, 4th ed., (U.S. Dept. of Health and Human Services, U.S. Government Printing Office, 1987), updates to the Kabat database, and other accessible U.S. and foreign databases (both nucleic acid and protein). Solvent accessibilities of V region amino acids can be deduced from the known three-dimensional structure for human and murine antibody fragments. There are two general steps in veneering a murine antigen-binding site. Initially, the FRs of the variable domains of an antibody molecule of interest are compared with corresponding FR sequences of human variable domains obtained from the above-identified sources. The most homologous human V regions are then compared residue by residue to corresponding murine amino acids. The residues in the murine FR which differ from the human counterpart are replaced by the residues present in the human moiety using recombinant techniques well known in the art. Residue switching is only carried out with moieties which are at least partially exposed (solvent accessible), and care is exercised in the replacement of amino acid residues which may have a significant effect

on the tertiary structure of V region domains, such as proline, glycine and charged amino acids.

In this manner, the resultant "veneered" murine antigen-binding sites are thus designed to retain the murine CDR residues, the residues substantially adjacent to the CDRs, the residues identified as buried or mostly buried (solvent inaccessible), the residues believed to participate in non-covalent (*e.g.*, electrostatic and hydrophobic) contacts between heavy and light chain domains, and the residues from conserved structural regions of the FRs which are believed to influence the "canonical" tertiary structures of the CDR loops. These design criteria are then used to prepare recombinant nucleotide sequences which combine the CDRs of both the heavy and light chain of a murine antigen-binding site into human-appearing FRs that can be used to transfect mammalian cells for the expression of recombinant human antibodies which exhibit the antigen specificity of the murine antibody molecule.

In another embodiment of the invention, monoclonal antibodies of the present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include ^{90}Y , ^{123}I , ^{125}I , ^{131}I , ^{186}Re , ^{188}Re , ^{211}At , and ^{212}Bi . Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, *Pseudomonas* exotoxin, *Shigella* toxin, and pokeweed antiviral protein.

A therapeutic agent may be coupled (*e.g.*, covalently bonded) to a suitable monoclonal antibody either directly or indirectly (*e.g.*, via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulfhydryl group, on one may be capable of reacting with a carbonyl-containing group, such as an anhydride or an acid halide, or with an alkyl group containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an agent or an antibody, and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described in the catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulfhydryl groups or oxidized carbohydrate residues. There are numerous references describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.

Where a therapeutic agent is more potent when free from the antibody portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitler), by irradiation of a photolabile bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter et al.), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn et al.), by serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler et al.).

It may be desirable to couple more than one agent to an antibody. In one embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent may be prepared in a variety of ways. For example, more than one agent may be coupled

directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as albumins (*e.g.*, U.S. Patent No. 4,507,234, to Kato et al.), peptides and polysaccharides such as aminodextran (*e.g.*, U.S. Patent No. 4,699,784, to Shih et al.). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (*e.g.*, U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide agents include radiohalogenated small molecules and chelating compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison et al. discloses representative chelating compounds and their synthesis.

15 T Cell Compositions

The present invention, in another aspect, provides T cells specific for a tumor polypeptide disclosed herein, or for a variant or derivative thereof. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient, using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or unrelated humans, non-human mammals, cell lines or cultures.

25 T cells may be stimulated with a polypeptide, polynucleotide encoding a polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide of interest. Preferably, a tumor

polypeptide or polynucleotide of the invention is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

T cells are considered to be specific for a polypeptide of the present invention if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen et al., *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with a tumor polypeptide (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days will typically result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN-γ) is indicative of T cell activation (see Coligan et al., Current Protocols in Immunology, vol. 1, Wiley Interscience (Greene 1998)). T cells that have been activated in response to a tumor polypeptide, polynucleotide or polypeptide-expressing APC may be CD4⁺ and/or CD8⁺. Tumor polypeptide-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

For therapeutic purposes, CD4⁺ or CD8⁺ T cells that proliferate in response to a tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a tumor polypeptide, or a short peptide corresponding to an immunogenic portion of such a polypeptide, with or without the

addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize a tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of the tumor polypeptide can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

5 Pharmaceutical Compositions

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable carriers for administration to a cell or an animal, either alone, or in combination with one or more other modalities of therapy.

10 It will be understood that, if desired, a composition as disclosed herein may be administered in combination with other agents as well, such as, *e.g.*, other proteins or polypeptides or various pharmaceutically-active agents. In fact, there is virtually no limit to other components that may also be included, given that the additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The
15 compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesized as described herein. Likewise, such compositions may further comprise substituted or derivatized RNA or DNA compositions.

20 Therefore, in another aspect of the present invention, pharmaceutical compositions are provided comprising one or more of the polynucleotide, polypeptide, antibody, and/or T-cell compositions described herein in combination with a physiologically acceptable carrier. In certain preferred embodiments, the pharmaceutical compositions of the invention comprise immunogenic polynucleotide and/or polypeptide
25 compositions of the invention for use in prophylactic and therapeutic vaccine applications. Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant approach)," Plenum Press (NY, 1995).

Generally, such compositions will comprise one or more polynucleotide and/or polypeptide compositions of the present invention in combination with one or more immunostimulants.

It will be apparent that any of the pharmaceutical compositions described herein can contain pharmaceutically acceptable salts of the polynucleotides and polypeptides of the invention. Such salts can be prepared, for example, from pharmaceutically acceptable non-toxic bases, including organic bases (*e.g.*, salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (*e.g.*, sodium, potassium, lithium, ammonium, calcium and magnesium salts).

In another embodiment, illustrative immunogenic compositions, *e.g.*, vaccine compositions, of the present invention comprise DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the polynucleotide may be administered within any of a variety of delivery systems known to those of ordinary skill in the art. Indeed, numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate polynucleotide expression systems will, of course, contain the necessary regulatory DNA regulatory sequences for expression in a patient (such as a suitable promoter and terminating signal). Alternatively, bacterial delivery systems may involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface or secretes such an epitope.

Therefore, in certain embodiments, polynucleotides encoding immunogenic polypeptides described herein are introduced into suitable mammalian host cells for expression using any of a number of known viral-based systems. In one illustrative embodiment, retroviruses provide a convenient and effective platform for gene delivery systems. A selected nucleotide sequence encoding a polypeptide of the present invention can be inserted into a vector and packaged in retroviral particles using techniques known in the art. The recombinant virus can then be isolated and delivered to a subject. A number of illustrative retroviral systems have been described (*e.g.*, U.S. Pat. No. 5,219,740; Miller and Rosman (1989) *BioTechniques* 7:980-990; Miller, A. D. (1990) *Human Gene Therapy*

1:5-14; Scarpa et al. (1991) *Virology* 180:849-852; Burns et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:8033-8037; and Boris-Lawrie and Temin (1993) *Cur. Opin. Genet. Develop.* 3:102-109.

In addition, a number of illustrative adenovirus-based systems have also
 5 been described. Unlike retroviruses which integrate into the host genome, adenoviruses persist extrachromosomally thus minimizing the risks associated with insertional mutagenesis (Haj-Ahmad and Graham (1986) *J. Virol.* 57:267-274; Bett et al. (1993) *J. Virol.* 67:5911-5921; Mittereder et al. (1994) *Human Gene Therapy* 5:717-729; Seth et al. (1994) *J. Virol.* 68:933-940; Barr et al. (1994) *Gene Therapy* 1:51-58; Berkner, K. L.
 10 (1988) *BioTechniques* 6:616-629; and Rich et al. (1993) *Human Gene Therapy* 4:461-476).

Various adeno-associated virus (AAV) vector systems have also been developed for polynucleotide delivery. AAV vectors can be readily constructed using techniques well known in the art. See, *e.g.*, U.S. Pat. Nos. 5,173,414 and 5,139,941; International Publication Nos. WO 92/01070 and WO 93/03769; Lebkowski et al. (1988)
 15 *Molec. Cell. Biol.* 8:3988-3996; Vincent et al. (1990) *Vaccines* 90 (Cold Spring Harbor Laboratory Press); Carter, B. J. (1992) *Current Opinion in Biotechnology* 3:533-539; Muzyczka, N. (1992) *Current Topics in Microbiol. and Immunol.* 158:97-129; Kotin, R. M. (1994) *Human Gene Therapy* 5:793-801; Shelling and Smith (1994) *Gene Therapy* 1:165-169; and Zhou et al. (1994) *J. Exp. Med.* 179:1867-1875.

20 Additional viral vectors useful for delivering the polynucleotides encoding polypeptides of the present invention by gene transfer include those derived from the pox family of viruses, such as vaccinia virus and avian poxvirus. By way of example, vaccinia virus recombinants expressing the novel molecules can be constructed as follows. The DNA encoding a polypeptide is first inserted into an appropriate vector so that it is adjacent
 25 to a vaccinia promoter and flanking vaccinia DNA sequences, such as the sequence encoding thymidine kinase (TK). This vector is then used to transfect cells which are simultaneously infected with vaccinia. Homologous recombination serves to insert the vaccinia promoter plus the gene encoding the polypeptide of interest into the viral genome.

The resulting TK.sup.(-) recombinant can be selected by culturing the cells in the presence of 5-bromodeoxyuridine and picking viral plaques resistant thereto.

A vaccinia-based infection/transfection system can be conveniently used to provide for inducible, transient expression or coexpression of one or more polypeptides described herein in host cells of an organism. In this particular system, cells are first infected in vitro with a vaccinia virus recombinant that encodes the bacteriophage T7 RNA polymerase. This polymerase displays exquisite specificity in that it only transcribes templates bearing T7 promoters. Following infection, cells are transfected with the polynucleotide or polynucleotides of interest, driven by a T7 promoter. The polymerase expressed in the cytoplasm from the vaccinia virus recombinant transcribes the transfected DNA into RNA which is then translated into polypeptide by the host translational machinery. The method provides for high level, transient, cytoplasmic production of large quantities of RNA and its translation products. See, *e.g.*, Elroy-Stein and Moss, Proc. Natl. Acad. Sci. USA (1990) 87:6743-6747; Fuerst et al. Proc. Natl. Acad. Sci. USA (1986) 83:8122-8126.

Alternatively, avipoxviruses, such as the fowlpox and canarypox viruses, can also be used to deliver the coding sequences of interest. Recombinant avipox viruses, expressing immunogens from mammalian pathogens, are known to confer protective immunity when administered to non-avian species. The use of an Avipox vector is particularly desirable in human and other mammalian species since members of the Avipox genus can only productively replicate in susceptible avian species and therefore are not infective in mammalian cells. Methods for producing recombinant Avipoxviruses are known in the art and employ genetic recombination, as described above with respect to the production of vaccinia viruses. See, *e.g.*, WO 91/12882; WO 89/03429; and WO 92/03545.

Any of a number of alphavirus vectors can also be used for delivery of polynucleotide compositions of the present invention, such as those vectors described in U.S. Patent Nos. 5,843,723; 6,015,686; 6,008,035 and 6,015,694. Certain vectors based on Venezuelan Equine Encephalitis (VEE) can also be used, illustrative examples of which can be found in U.S. Patent Nos. 5,505,947 and 5,643,576.

Moreover, molecular conjugate vectors, such as the adenovirus chimeric vectors described in Michael et al. *J. Biol. Chem.* (1993) 268:6866-6869 and Wagner et al. *Proc. Natl. Acad. Sci. USA* (1992) 89:6099-6103, can also be used for gene delivery under the invention.

5 Additional illustrative information on these and other known viral-based delivery systems can be found, for example, in Fisher-Hoch et al., *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner et al., *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner et al., *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; 10 Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld et al., *Science* 252:431-434, 1991; Kolls et al., *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler et al., *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman et al., *Circulation* 88:2838-2848, 1993; and Guzman et al., *Cir. Res.* 73:1202-1207, 1993.

In certain embodiments, a polynucleotide may be integrated into the genome 15 of a target cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the polynucleotide may be stably maintained in the cell as a separate, episomal segment of DNA. Such polynucleotide segments or "episomes" encode sequences sufficient to permit maintenance 20 and replication independent of or in synchronization with the host cell cycle. The manner in which the expression construct is delivered to a cell and where in the cell the polynucleotide remains is dependent on the type of expression construct employed.

In another embodiment of the invention, a polynucleotide is administered/delivered as "naked" DNA, for example as described in Ulmer et al., *Science* 25 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

In still another embodiment, a composition of the present invention can be delivered via a particle bombardment approach, many of which have been described. In

one illustrative example, gas-driven particle acceleration can be achieved with devices such as those manufactured by Powderject Pharmaceuticals PLC (Oxford, UK) and Powderject Vaccines Inc. (Madison, WI), some examples of which are described in U.S. Patent Nos. 5,846,796; 6,010,478; 5,865,796; 5,584,807; and EP Patent No. 0500 799. This approach
 5 offers a needle-free delivery approach wherein a dry powder formulation of microscopic particles, such as polynucleotide or polypeptide particles, are accelerated to high speed within a helium gas jet generated by a hand held device, propelling the particles into a target tissue of interest.

In a related embodiment, other devices and methods that may be useful for
 10 gas-driven needle-less injection of compositions of the present invention include those provided by Bioject, Inc. (Portland, OR), some examples of which are described in U.S. Patent Nos. 4,790,824; 5,064,413; 5,312,335; 5,383,851; 5,399,163; 5,520,639 and 5,993,412.

According to another embodiment, the pharmaceutical compositions
 15 described herein will comprise one or more immunostimulants in addition to the immunogenic polynucleotide, polypeptide, antibody, T-cell and/or APC compositions of this invention. An immunostimulant refers to essentially any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. One preferred type of immunostimulant comprises an adjuvant. Many adjuvants contain a
 20 substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins. Certain adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company,
 25 Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA); aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres;

monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF, interleukin-2, -7, -12, and other like growth factors, may also be used as adjuvants.

Within certain embodiments of the invention, the adjuvant composition is preferably one that induces an immune response predominantly of the Th1 type. High levels of Th1-type cytokines (*e.g.*, IFN- γ , TNF α , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast, high levels of Th2-type cytokines (*e.g.*, IL-4, IL-5, IL-6 and IL-10) tend to favor the induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-type responses.

Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.

Certain preferred adjuvants for eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-de-O-acylated monophosphoryl lipid A, together with an aluminum salt. MPL[®] adjuvants are available from Corixa Corporation (Seattle, WA; *see*, for example, US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato et al., *Science* 273:352, 1996. Another preferred adjuvant comprises a saponin, such as Quil A, or derivatives thereof, including QS21 and QS7 (Aquila Biopharmaceuticals Inc., Framingham, MA); Escin; Digitonin; or *Gypsophila* or *Chenopodium quinoa* saponins. Other preferred formulations include more than one saponin in the adjuvant combinations of the present invention, for example combinations of at least two of the following group comprising QS21, QS7, Quil A, β -escin, or digitonin.

Alternatively the saponin formulations may be combined with vaccine vehicles composed of chitosan or other polycationic polymers, polylactide and polylactide-co-glycolide particles, poly-N-acetyl glucosamine-based polymer matrix, particles composed of polysaccharides or chemically modified polysaccharides, liposomes and lipid-based particles, particles composed of glycerol monoesters, etc. The saponins may also be formulated in the presence of cholesterol to form particulate structures such as liposomes or ISCOMs. Furthermore, the saponins may be formulated together with a polyoxyethylene ether or ester, in either a non-particulate solution or suspension, or in a particulate structure such as a paucilamellar liposome or ISCOM. The saponins may also be formulated with excipients such as Carbopol[®] to increase viscosity, or may be formulated in a dry powder form with a powder excipient such as lactose.

In one preferred embodiment, the adjuvant system includes the combination of a monophosphoryl lipid A and a saponin derivative, such as the combination of QS21 and 3D-MPL[®] adjuvant, as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO 96/33739. Other preferred formulations comprise an oil-in-water emulsion and tocopherol. Another particularly preferred adjuvant formulation employing QS21, 3D-MPL[®] adjuvant and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

Another enhanced adjuvant system involves the combination of a CpG-containing oligonucleotide and a saponin derivative particularly the combination of CpG and QS21 is disclosed in WO 00/09159. Preferably the formulation additionally comprises an oil in water emulsion and tocopherol.

Additional illustrative adjuvants for use in the pharmaceutical compositions of the invention include Montanide ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (*e.g.*, SBAS-2 or SBAS-4, available from SmithKline Beecham, Rixensart, Belgium), Detox (Enhanzyn[®]) (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures of which are

incorporated herein by reference in their entireties, and polyoxyethylene ether adjuvants such as those described in WO 99/52549A1.

Other preferred adjuvants include adjuvant molecules of the general formula
(I): $\text{HO}(\text{CH}_2\text{CH}_2\text{O})_n\text{-A-R}$,

5 wherein, n is 1-50, A is a bond or $-\text{C}(\text{O})-$, R is C_{1-50} alkyl or Phenyl C_{1-50} alkyl.

One embodiment of the present invention consists of a vaccine formulation comprising a polyoxyethylene ether of general formula (I), wherein n is between 1 and 50, preferably 4-24, most preferably 9; the R component is C_{1-50} , preferably $\text{C}_4\text{-C}_{20}$ alkyl and most preferably C_{12} alkyl, and A is a bond. The concentration of the polyoxyethylene ethers
10 should be in the range 0.1-20%, preferably from 0.1-10%, and most preferably in the range 0.1-1%. Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether, polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether. Polyoxyethylene ethers such as polyoxyethylene lauryl
15 ether are described in the Merck index (12th edition: entry 7717). These adjuvant molecules are described in WO 99/52549.

The polyoxyethylene ether according to the general formula (I) above may, if desired, be combined with another adjuvant. For example, a preferred adjuvant combination is preferably with CpG as described in the pending UK patent application GB
20 9820956.2.

According to another embodiment of this invention, an immunogenic composition described herein is delivered to a host via antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to
25 increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (see Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In general, dendritic cells may be identified based on their typical shape (stellate *in situ*, with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (see Zitvogel et al., *Nature Med.* 4:594-600, 1998).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF α to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF α , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc γ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface

molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide of the invention
 5 (or portion or other variant thereof) such that the encoded polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a pharmaceutical composition comprising such transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient,
 10 resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi et al., *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the tumor polypeptide, DNA
 15 (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the
 20 polypeptide.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier will typically vary depending on the mode of administration. Compositions of the present invention may be formulated for any appropriate manner of administration, including for
 25 example, topical, oral, nasal, mucosal, intravenous, intracranial, intraperitoneal, subcutaneous and intramuscular administration.

Carriers for use within such pharmaceutical compositions are biocompatible, and may also be biodegradable. In certain embodiments, the formulation preferably provides a relatively constant level of active component release. In other embodiments,

however, a more rapid rate of release immediately upon administration may be desired. The formulation of such compositions is well within the level of ordinary skill in the art using known techniques. Illustrative carriers useful in this regard include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like.

5 Other illustrative delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained
10 within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

In another illustrative embodiment, biodegradable microspheres (*e.g.*, polylactate polyglycolate) are employed as carriers for the compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent
15 Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344, 5,407,609 and 5,942,252. Modified hepatitis B core protein carrier systems, such as described in WO/99 40934, and references cited therein, will also be useful for many applications. Another illustrative carrier/delivery system employs a carrier comprising particulate-protein complexes, such as those described in U.S. Patent No. 5,928,647, which
20 are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

The pharmaceutical compositions of the invention will often further comprise one or more buffers (*e.g.*, neutral buffered saline or phosphate buffered saline), carbohydrates (*e.g.*, glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents
25 such as EDTA or glutathione, adjuvants (*e.g.*, aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate.

The pharmaceutical compositions described herein may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are typically sealed in such a way to preserve the sterility and stability of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, a pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

The development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation, is well known in the art, some of which are briefly discussed below for general purposes of illustration.

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (see, for example, Mathiowitz *et al.*, Nature 1997 Mar 27;386(6623):410-4; Hwang *et al.*, Crit Rev Ther Drug Carrier Syst 1998;15(3):243-84; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451). Tablets, troches, pills, capsules and the like may also contain any of a variety of additional components, for example, a binder, such as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be

present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may
5 be incorporated into sustained-release preparation and formulations.

Typically, these formulations will contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active
10 compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and
15 treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. Alternatively, the active ingredient may be incorporated into an oral solution such as one
20 containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

25 In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally. Such approaches are well known to the skilled artisan, some of which are further described, for example, in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363. In certain embodiments, solutions of the active compounds as free base

or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations generally will contain a preservative to prevent the growth of microorganisms.

Illustrative pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (for example, see U. S. Patent 5,466,468). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and/or by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

In one embodiment, for parenteral administration in an aqueous solution, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or

injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. Moreover, for human administration, preparations will of course preferably meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

In another embodiment of the invention, the compositions disclosed herein may be formulated in a neutral or salt form. Illustrative pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective.

The carriers can further comprise any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions. The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a human.

In certain embodiments, the pharmaceutical compositions may be delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the lungs *via* nasal aerosol sprays has been described, *e.g.*, in U. S. Patent 5,756,353 and U. S. Patent

5,804,212. Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, J Controlled Release 1998 Mar 2;52(1-2):81-7) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871) are also well-known in the pharmaceutical arts. Likewise, illustrative transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045.

In certain embodiments, liposomes, nanocapsules, microparticles, lipid particles, vesicles, and the like, are used for the introduction of the compositions of the present invention into suitable host cells/organisms. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like. Alternatively, compositions of the present invention can be bound, either covalently or non-covalently, to the surface of such carrier vehicles.

The formation and use of liposome and liposome-like preparations as potential drug carriers is generally known to those of skill in the art (see for example, Lasic, Trends Biotechnol 1998 Jul;16(7):307-21; Takakura, Nippon Rinsho 1998 Mar;56(3):691-5; Chandran *et al.*, Indian J Exp Biol. 1997 Aug;35(8):801-9; Margalit, Crit Rev Ther Drug Carrier Syst. 1995;12(2-3):233-61; U.S. Patent 5,567,434; U.S. Patent 5,552,157; U.S. Patent 5,565,213; U.S. Patent 5,738,868 and U.S. Patent 5,795,587, each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that are normally difficult to transfect by other procedures, including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, J Biol Chem. 1990 Sep 25;265(27):16337-42; Muller *et al.*, DNA Cell Biol. 1990 Apr;9(3):221-9). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, various drugs, radiotherapeutic agents, enzymes, viruses, transcription factors, allosteric effectors and the like, into a variety of cultured cell lines and animals. Furthermore, the use of liposomes does not appear to be associated with autoimmune responses or unacceptable toxicity after systemic delivery.

In certain embodiments, liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs)).

Alternatively, in other embodiments, the invention provides for
 5 pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (see, for example, Quintanar-Guerrero *et al.*, Drug Dev Ind Pharm. 1998 Dec;24(12):1113-28). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) may be designed using polymers able to be degraded *in*
 10 *vivo*. Such particles can be made as described, for example, by Couvreur *et al.*, Crit Rev Ther Drug Carrier Syst. 1988;5(1):1-20; zur Muhlen *et al.*, Eur J Pharm Biopharm. 1998 Mar;45(2):149-55; Zambaux *et al.* J Controlled Release. 1998 Jan 2;50(1-3):31-40; and U. S. Patent 5,145,684.

Cancer Therapeutic Methods

15 In further aspects of the present invention, the pharmaceutical compositions described herein may be used for the treatment of cancer, particularly for the immunotherapy of breast cancer. Within such methods, the pharmaceutical compositions described herein are administered to a patient, typically a warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer. Accordingly, the above
 20 pharmaceutical compositions may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. As discussed above, administration of the pharmaceutical compositions may be by any
 25 suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous host

immune system to react against tumors with the administration of immune response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established tumor-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8⁺ cytotoxic T lymphocytes and CD4⁺ T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-activated killer cells), B cells and antigen-presenting cells (such as dendritic cells and macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic antibodies (as described above and in U.S. Patent No. 4,918,164) for passive immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture conditions typically use intermittent stimulation with antigen, often in the presence of cytokines (such as IL-2) and non-dividing feeder cells. As noted above, immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage, monocyte, fibroblast and/or B cells, may be pulsed with immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a recombinant virus or other expression system. Cultured effector cells for use in therapy must be able to grow and distribute widely, and to survive long term *in vivo*.

Studies have shown that cultured effector cells can be induced to grow *in vivo* and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (*see, for example, Cheever et al., Immunological Reviews 157:177, 1997*).

5 Alternatively, a vector expressing a polypeptide recited herein may be introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitary, intraperitoneal or intratumor administration.

10 Routes and frequency of administration of the therapeutic compositions described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably,
 15 between 1 and 10 doses may be administered over a 52 week period. Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the
 20 basal (*i.e.*, untreated) level. Such response can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor cells *in vitro*. Such vaccines should also be capable of causing an immune response that leads to an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial or longer disease-free survival) in vaccinated
 25 patients as compared to non-vaccinated patients. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 25 µg to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit. Such a response can be monitored by establishing an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial, or longer disease-free survival) in treated patients as compared to non-treated patients. Increases in preexisting immune responses to a tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using standard proliferation, cytotoxicity or cytokine assays, which may be performed using samples obtained from a patient before and after treatment.

Cancer Detection and Diagnostic Compositions, Methods and Kits

In general, a cancer may be detected in a patient based on the presence of one or more breast tumor proteins and/or polynucleotides encoding such proteins in a biological sample (for example, blood, sera, sputum urine and/or tumor biopsies) obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as breast cancer. In addition, such proteins may be useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the biological sample. Polynucleotide primers and probes may be used to detect the level of mRNA encoding a tumor protein, which is also indicative of the presence or absence of a cancer. In general, a breast tumor sequence should be present at a level that is at least three fold higher in tumor tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in the art for using a binding agent to detect polypeptide markers in a sample. *See, e.g.*, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, the presence or absence of a cancer in a patient may be determined by (a) contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c) comparing the level of polypeptide with a predetermined cut-off value.

In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding agent/polypeptide complex.

5 Such detection reagents may comprise, for example, a binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized binding agent after incubation of the binding agent
10 with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length breast tumor proteins and polypeptide portions thereof to which the binding agent binds, as described above.

15 The solid support may be any material known to those of ordinary skill in the art to which the tumor protein may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic
20 particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681. The binding agent may be immobilized on the solid support using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which
25 may be a direct linkage between the agent and functional groups on the support or may be a linkage by way of a cross-linking agent). Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1

hour and about 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10 μ g, and preferably about 100 ng to about 1 μ g, is sufficient to immobilize an adequate amount of binding agent.

5 Covalent attachment of binding agent to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group
10 on the support with an amine and an active hydrogen on the binding partner (*see, e.g.*, Pierce Immunotechnology Catalog and Handbook, 1991, at A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the sample, such that
15 polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a method appropriate for the
20 specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20™ (Sigma Chemical Co., St. Louis, MO). The immobilized
25 antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with breast cancer. Preferably, the contact time is

sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20™. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide. An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

To determine the presence or absence of a cancer, such as breast cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value for the detection of a cancer is the average mean signal obtained when the immobilized antibody is incubated with samples from patients without the cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve,

according to the method of Sackett et al., *Clinical Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985, p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%-specificity) that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the immobilized binding agent as the sample passes through the membrane. A second, labeled binding agent then binds to the binding agent-polypeptide complex as a solution containing the second binding agent flows through the membrane. The detection of bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the sample. The sample migrates along the membrane through a region containing second binding agent and to the area of immobilized binding agent. Concentration of second binding agent at the area of immobilized antibody indicates the presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of binding agent immobilized on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a positive signal in the two-antibody sandwich assay, in the format discussed above. Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of

antibody immobilized on the membrane ranges from about 25 ng to about 1µg, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological sample.

Of course, numerous other assay protocols exist that are suitable for use with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such tumor protein specific antibodies may correlate with the presence of a cancer.

A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with a tumor protein in a biological sample. Within certain methods, a biological sample comprising CD4⁺ and/or CD8⁺ T cells isolated from a patient is incubated with a tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For example, T cells may be isolated from a patient by routine techniques (such as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (*e.g.*, 5 - 25 µg/ml). It may be desirable to incubate another aliquot of a T cell sample in the absence of tumor polypeptide to serve as a control. For CD4⁺ T cells, activation is preferably detected by evaluating proliferation of the T cells. For CD8⁺ T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on the level of mRNA encoding a tumor protein in a biological sample. For example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of a tumor cDNA derived from a biological sample, wherein at

least one of the oligonucleotide primers is specific for (*i.e.*, hybridizes to) a polynucleotide encoding the tumor protein. The amplified cDNA is then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a polynucleotide encoding a tumor protein may be
 5 used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers and probes should comprise an oligonucleotide sequence that has at least about 60%, preferably at least about 75% and more preferably at least about 90%, identity to a portion of a
 10 polynucleotide encoding a tumor protein of the invention that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a polypeptide described herein under moderately stringent conditions, as defined above. Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are
 15 at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule having a sequence as disclosed herein. Techniques for both PCR based assays and hybridization assays are well known in the art (*see*, for example, Mullis et al., *Cold Spring Harbor Symp. Quant. Biol.*, 51:263, 1987; Erlich ed.,
 20 *PCR Technology*, Stockton Press, NY, 1989).

One preferred assay employs RT-PCR, in which PCR is applied in conjunction with reverse transcription. Typically, RNA is extracted from a biological sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be
 25 separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and from an individual who is not afflicted with a cancer. The amplification reaction may be performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in

expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered positive.

In another embodiment, the compositions described herein may be used as markers for the progression of cancer. In this embodiment, assays as described above for the diagnosis of a cancer may be performed over time, and the change in the level of reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter performed as needed. In general, a cancer is progressing in those patients in whom the level of polypeptide or polynucleotide detected increases over time. In contrast, the cancer is not progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

Certain *in vivo* diagnostic assays may be performed directly on a tumor. One such assay involves contacting tumor cells with a binding agent. The bound binding agent may then be detected directly or indirectly via a reporter group. Such binding agents may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

As noted above, to improve sensitivity, multiple tumor protein markers may be assayed within a given sample. It will be apparent that binding agents specific for different proteins provided herein may be combined within a single assay. Further, multiple primers or probes may be used concurrently. The selection of tumor protein markers may be based on routine experiments to determine combinations that results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

The present invention further provides kits for use within any of the above diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment. For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to a tumor protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or

more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

5 Alternatively, a kit may be designed to detect the level of mRNA encoding a tumor protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridizes to a polynucleotide encoding a tumor protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that may be present within such kits
10 include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a tumor protein.

The following Examples are offered by way of illustration and not by way of limitation.

15 EXAMPLE 1

PREPARATION OF BREAST TUMOR-SPECIFIC cDNAs USING

DIFFERENTIAL DISPLAY RT-PCR

This Example illustrates the preparation of cDNA molecules encoding
20 breast tumor-specific polypeptides using a differential display screen.

A. Preparation of B18Ag1 cDNA and Characterization of mRNA Expression

Tissue samples were prepared from breast tumor and normal tissue of a patient with breast cancer that was confirmed by pathology after removal from the patient. Normal RNA and tumor RNA was extracted from the samples and mRNA was isolated and
25 converted into cDNA using a (dT)₁₂AG (SEQ ID NO:130) anchored 3' primer. Differential display PCR was then executed using a randomly chosen primer (CTTCAACCTC) (SEQ ID NO:103). Amplification conditions were standard buffer containing 1.5 mM MgCl₂, 20 pmol of primer, 500 pmol dNTP, and 1 unit of *Taq* DNA polymerase (Perkin-Elmer,

Branchburg, NJ). Forty cycles of amplification were performed using 94°C denaturation for 30 seconds, 42°C annealing for 1 minute, and 72°C extension for 30 seconds. An RNA fingerprint containing 76 amplified products was obtained. Although the RNA fingerprint of breast tumor tissue was over 98% identical to that of the normal breast tissue, a band was repeatedly observed to be specific to the RNA fingerprint pattern of the tumor. This band was cut out of a silver stained gel, subcloned into the T-vector (Novagen, Madison, WI) and sequenced.

The sequence of the cDNA, referred to as B18Ag1, is provided in SEQ ID NO:1. A database search of GENBANK and EMBL revealed that the B18Ag1 fragment initially cloned is 77% identical to the endogenous human retroviral element S71, which is a truncated retroviral element homologous to the Simian Sarcoma Virus (SSV). S71 contains an incomplete *gag* gene, a portion of the *pol* gene and an LTR-like structure at the 3' terminus (*see* Werner et al., *Virology* 174:225-238 (1990)). B18Ag1 is also 64% identical to SSV in the region corresponding to the P30 (*gag*) locus. B18Ag1 contains three separate and incomplete reading frames covering a region which shares considerable homology to a wide variety of *gag* proteins of retroviruses which infect mammals. In addition, the homology to S71 is not just within the *gag* gene, but spans several kb of sequence including an LTR.

B18Ag1-specific PCR primers were synthesized using computer analysis guidelines. RT-PCR amplification (94°C, 30 seconds; 60°C → 42°C, 30 seconds; 72°C, 30 seconds for 40 cycles) confirmed that B18Ag1 represents an actual mRNA sequence present at relatively high levels in the patient's breast tumor tissue. The primers used in amplification were B18Ag1-1 (CTG CCT GAG CCA CAA ATG) (SEQ ID NO:128) and B18Ag1-4 (CCG GAG GAG GAA GCT AGA GGA ATA) (SEQ ID NO:129) at a 3.5 mM magnesium concentration and a pH of 8.5, and B18Ag1-2 (ATG GCT ATT TTC GGG GCC TGA CA) (SEQ ID NO:126) and B18Ag1-3 (CCG GTA TCT CCT CGT GGG TAT T) (SEQ ID NO:127) at 2 mM magnesium at pH 9.5. The same experiments showed exceedingly low to nonexistent levels of expression in this patient's normal breast tissue (*see* Figure 1). RT-PCR experiments were then used to show that B18Ag1 mRNA is

present in nine other breast tumor samples (from Brazilian and American patients) but absent in, or at exceedingly low levels in, the normal breast tissue corresponding to each cancer patient. RT-PCR analysis has also shown that the B18Ag1 transcript is not present in various normal tissues (including lymph node, myocardium and liver) and present at
 5 relatively low levels in PBMC and lung tissue. The presence of B18Ag1 mRNA in breast tumor samples, and its absence from normal breast tissue, has been confirmed by Northern blot analysis, as shown in Figure 2.

The differential expression of B18Ag1 in breast tumor tissue was also confirmed by RNase protection assays. Figure 3 shows the level of B18Ag1 mRNA in
 10 various tissue types as determined in four different RNase protection assays. Lanes 1-12 represent various normal breast tissue samples, lanes 13-25 represent various breast tumor samples; lanes 26-27 represent normal prostate samples; lanes 28-29 represent prostate tumor samples; lanes 30-32 represent colon tumor samples; lane 33 represents normal aorta; lane 34 represents normal small intestine; lane 35 represents normal skin, lane 36
 15 represents normal lymph node; lane 37 represents normal ovary; lane 38 represents normal liver; lane 39 represents normal skeletal muscle; lane 40 represents a first normal stomach sample, lane 41 represents a second normal stomach sample; lane 42 represents a normal lung; lane 43 represents normal kidney; and lane 44 represents normal pancreas. Interexperimental comparison was facilitated by including a positive control RNA of
 20 known β -actin message abundance in each assay and normalizing the results of the different assays with respect to this positive control.

RT-PCR and Southern Blot analysis has shown the B18Ag1 locus to be present in human genomic DNA as a single copy endogenous retroviral element. A genomic clone of approximately 12-18 kb was isolated using the initial B18Ag1 sequence
 25 as a probe. Four additional subclones were also isolated by XbaI digestion. Additional retroviral sequences obtained from the ends of the XbaI digests of these clones (located as shown in Figure 4) are shown as SEQ ID NO:3 - SEQ ID NO:10, where SEQ ID NO:3 shows the location of the sequence labeled 10 in Figure 4, SEQ ID NO:4 shows the location of the sequence labeled 11-29, SEQ ID NO:5 shows the location of the sequence

labeled 3, SEQ ID NO:6 shows the location of the sequence labeled 6, SEQ ID NO:7 shows the location of the sequence labeled 12, SEQ ID NO:8 shows the location of the sequence labeled 13, SEQ ID NO:9 shows the location of the sequence labeled 14 and SEQ ID NO:10 shows the location of the sequence labeled 11-22.

5 Subsequent studies demonstrated that the 12-18 kb genomic clone contains a retroviral element of about 7.75 kb, as shown in Figures 5A and 5B. The sequence of this retroviral element is shown in SEQ ID NO:141. The numbered line at the top of Figure 5A represents the sense strand sequence of the retroviral genomic clone. The box below this line shows the position of selected restriction sites. The arrows depict the different
10 overlapping clones used to sequence the retroviral element. The direction of the arrow shows whether the single-pass subclone sequence corresponded to the sense or anti-sense strand. Figure 5B is a schematic diagram of the retroviral element containing B18Ag1 depicting the organization of viral genes within the element. The open boxes correspond to predicted reading frames, starting with a methionine, found throughout the element. Each
15 of the six likely reading frames is shown, as indicated to the left of the boxes, with frames 1-3 corresponding to those found on the sense strand.

Using the cDNA of SEQ ID NO:1 as a probe, a longer cDNA was obtained (SEQ ID NO:227) which contains minor nucleotide differences (less than 1%) compared to the genomic sequence shown in SEQ ID NO:141.

20 B. Preparation of cDNA Molecules Encoding Other Breast Tumor-Specific Polypeptides

Normal RNA and tumor RNA was prepared and mRNA was isolated and converted into cDNA using a (dT)₁₂AG anchored 3' primer, as described above. Differential display PCR was then executed using the randomly chosen primers of SEQ ID
25 NOs:87-125. Amplification conditions were as noted above, and bands observed to be specific to the RNA fingerprint pattern of the tumor were cut out of a silver stained gel, subcloned into either the T-vector (Novagen, Madison, WI) or the pCRII vector (Invitrogen, San Diego, CA) and sequenced. The sequences are provided in SEQ ID

NO:11 - SEQ ID NO:86. Of the 79 sequences isolated, 67 were found to be novel (SEQ ID NOs:11-26 and 28-77) (*see also* Figures 6-20).

An extended DNA sequence (SEQ ID NO:290) for the antigen B15Ag1 (originally identified partial sequence provided in SEQ ID NO:27) was obtained in further studies. Comparison of the sequence of SEQ ID NO:290 with those in the gene bank as described above, revealed homology to the known human β -A activin gene. Further studies led to the isolation of the full-length cDNA sequence for the antigen B21GT2 (also referred to as B311D; originally identified partial cDNA sequence provided in SEQ ID NOs:56). The full-length sequence is provided in SEQ ID NO:307, with the corresponding amino acid sequence being provided in SEQ ID NO:308. Further studies led to the isolation of a splice variant of B311D. The B311D clone of SEQ ID NO:316 was sequenced and a XhoI/NotI fragment from this clone was gel purified and ³²P-cDTP labeled by random priming for use as a probe for further screening to obtain additional B311D gene sequence. Two fractions of a human breast tumor cDNA bacterial library were screened using standard techniques. One of the clones isolated in this manner yielded additional sequence which includes a poly A⁺ tail. The determined cDNA sequence of this clone (referred to as B311D_BT1_1A) is provided in SEQ ID NO:317. The sequences of SEQ ID NOs:316 and 317 were found to share identity over a 464 bp region, with the sequences diverging near the poly A⁺ sequence of SEQ ID NO:317.

Subsequent studies identified an additional 146 sequences (SEQ ID NOs:142-289), of which 115 appeared to be novel (SEQ ID NOs:142, 143, 146-152, 154-166, 168-176, 178-192, 194-198, 200-204, 206, 207, 209-214, 216, 218, 219, 221-240, 243-245, 247, 250, 251, 253, 255, 257-266, 268, 269, 271-273, 275, 276, 278, 280, 281, 284, 288 and 291). To the best of the inventors' knowledge none of the previously identified sequences have heretofore been shown to be expressed at a greater level in human breast tumor tissue than in normal breast tissue.

In further studies, several different splice forms of the antigen B11Ag1 (also referred to as B305D) were isolated, with each of the various splice forms containing slightly different versions of the B11Ag1 coding frame. Splice junction sequences define individual exons

which, in various patterns and arrangements, make up the various splice forms. Primers were designed to examine the expression pattern of each of the exons using RT-PCR as described below. Each exon was found to show the same expression pattern as the original B11Ag1 clone, with expression being breast tumor-, normal prostate- and normal testis-specific. The determined cDNA sequences for the isolated protein coding exons are provided in SEQ ID NOs:292-298, respectively. The predicted amino acid sequences corresponding to the sequences of SEQ ID NOs:292 and 298 are provided in SEQ ID NOs:299 and 300. Additional studies using rapid amplification of cDNA ends (RACE), a 5' specific primer to one of the splice forms of B11Ag1 provided above and a breast adenocarcinoma, led to the isolation of three additional, related, splice forms referred to as isoforms B11C-15, B11C-8 and B11C-9,16. The determined cDNA sequences for these isoforms are provided in SEQ ID NO: 301-303, with the corresponding predicted amino acid sequences being provided in SEQ ID NOs:304-306.

The protein coding region of B11C-15 (SEQ ID NO: 301; also referred to as B305D isoform C) was used as a query sequence in a BLASTN search of the Genbank DNA database. A match was found to a genomic clone from chromosome 21 (Accession no. AP001465). The pairwise alignments provided in the BLASTN output were used to identify the putative exon, or coding, sequence of the chromosome 21 sequence that corresponds to the B305D sequence. Based on the BlastN pairwise alignments, the following pieces of GenBank record AP001465 were put together: base pairs 67978-68499, 72870-72987, 73144-73335, 76085-76206, 77905-78085, 80520-80624, 87602-87633. This sequence was then aligned with the B305D isoform C sequence using the DNA Star Seqman program and excess sequence was deleted in such a way as to maintain the sequence most similar to B305D. The final edited form of the chromosome 21 sequence was 96.5% identical to B305D. This resulting edited sequence from chromosome 21 was then translated and found to contain no stop codons other than the final stop codon in the same position as that for B305D. As with B305D, the chromosome 21 sequence (provided in SEQ ID NO: 325) encoded a protein (SEQ ID NO: 326) with 384 amino acids. An

alignment of this protein with the B305D isoform C protein (SEQ ID NO: 304) showed 90% amino acid identity.

In subsequent studies on B305D isoform A (cDNA sequence provided in SEQ ID NO:292), the cDNA sequence (provided in SEQ ID NO:313) was found to contain an additional guanine residue at position 884, leading to a frameshift in the open reading frame. The determined DNA sequence of this ORF is provided in SEQ ID NO:314. This frameshift generates a protein sequence (provided in SEQ ID NO:315) of 293 amino acids that contains the C-terminal domain common to the other isoforms of B305D but that differs in the N-terminal region.

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EXAMPLE 2

PREPARATION OF B18Ag1 DNA FROM HUMAN GENOMIC DNA

This Example illustrates the preparation of B18Ag1 DNA by amplification from human genomic DNA.

B18Ag1 DNA may be prepared from 250 ng human genomic DNA using 20 pmol of B18Ag1 specific primers, 500 pmol dNTPS and 1 unit of *Taq* DNA polymerase (Perkin Elmer, Branchburg, NJ) using the following amplification parameters: 94°C for 30 seconds denaturing, 30 seconds 60°C to 42°C touchdown annealing in 2°C increments every two cycles and 72°C extension for 30 seconds. The last increment (a 42°C annealing temperature) should cycle 25 times. Primers were selected using computer analysis. Primers synthesized were B18Ag1-1, B18Ag1-2, B18Ag1-3, and B18Ag1-4. Primer pairs that may be used are 1+3, 1+4, 2+3, and 2+4.

Following gel electrophoresis, the band corresponding to B18Ag1 DNA may be excised and cloned into a suitable vector.

EXAMPLE 3

PREPARATION OF B18Ag1 DNA FROM BREAST TUMOR cDNA

This Example illustrates the preparation of B18Ag1 DNA by amplification
 5 from human breast tumor cDNA.

First strand cDNA is synthesized from RNA prepared from human breast tumor tissue in a reaction mixture containing 500 ng poly A+ RNA, 200 pmol of the primer (T)₁₂AG (*i.e.*, TTT TTT TTT TTT AG) (SEQ ID NO:130), 1X first strand reverse transcriptase buffer, 6.7 mM DTT, 500 mmol dNTPs, and 1 unit AMV or MMLV reverse
 10 transcriptase (from any supplier, such as Gibco-BRL (Grand Island, NY)) in a final volume of 30 μ l. After first strand synthesis, the cDNA is diluted approximately 25 fold and 1 μ l is used for amplification as described in Example 2. While some primer pairs can result in a heterogeneous population of transcripts, the primers B18Ag1-2 (5'ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:126) and B18Ag1-3 (5'CCG GTA TCT CCT CGT
 15 GGG TAT T) (SEQ ID NO:127) yield a single 151 bp amplification product.

EXAMPLE 4

IDENTIFICATION OF B-CELL AND T-CELL EPITOPES OF B18Ag1

20 This Example illustrates the identification of B18Ag1 epitopes.

The B18Ag1 sequence can be screened using a variety of computer algorithms. To determine B-cell epitopes, the sequence can be screened for hydrophobicity and hydrophilicity values using the method of Hopp, *Prog. Clin. Biol. Res.* 172B:367-77 (1985) or, alternatively, Cease et al., *J. Exp. Med.* 164:1779-84 (1986) or Spouge et al., *J.*
 25 *Immunol.* 138:204-12 (1987). Additional Class II MHC (antibody or B-cell) epitopes can be predicted using programs such as AMPHI (*e.g.*, Margalit et al., *J. Immunol.* 138:2213 (1987)) or the methods of Rothbard and Taylor (*e.g.*, *EMBO J.* 7:93 (1988)).

Once peptides (15-20 amino acids long) are identified using these techniques, individual peptides can be synthesized using automated peptide synthesis

equipment (available from manufacturers such as Perkin Elmer/Applied Biosystems Division, Foster City, CA) and techniques such as Merrifield synthesis. Following synthesis, the peptides can be used to screen sera harvested from either normal or breast cancer patients to determine whether patients with breast cancer possess antibodies reactive with the peptides. Presence of such antibodies in breast cancer patient would confirm the immunogenicity of the specific B-cell epitope in question. The peptides can also be tested for their ability to generate a serologic or humoral immune response in animals (mice, rats, rabbits, chimps etc.) following immunization *in vivo*. Generation of a peptide-specific antiserum following such immunization further confirms the immunogenicity of the specific B-cell epitope in question.

To identify T-cell epitopes, the B18Ag1 sequence can be screened using different computer algorithms which are useful in identifying 8-10 amino acid motifs within the B18Ag1 sequence which are capable of binding to HLA Class I MHC molecules. (*see, e.g., Rammensee et al., Immunogenetics 41:178-228 (1995)*). Following synthesis such peptides can be tested for their ability to bind to class I MHC using standard binding assays (*e.g., Sette et al., J. Immunol. 153:5586-92 (1994)*) and more importantly can be tested for their ability to generate antigen reactive cytotoxic T-cells following *in vitro* stimulation of patient or normal peripheral mononuclear cells using, for example, the methods of Bakker et al., *Cancer Res. 55:5330-34 (1995)*; Visseren et al., *J. Immunol. 154:3991-98 (1995)*; Kawakami et al., *J. Immunol. 154:3961-68 (1995)*; and Kast et al., *J. Immunol. 152:3904-12 (1994)*. Successful *in vitro* generation of T-cells capable of killing autologous (bearing the same Class I MHC molecules) tumor cells following *in vitro* peptide stimulation further confirms the immunogenicity of the B18Ag1 antigen. Furthermore, such peptides may be used to generate murine peptide and B18Ag1 reactive cytotoxic T-cells following *in vivo* immunization in mice rendered transgenic for expression of a particular human MHC Class I haplotype (Vitiello et al., *J. Exp. Med. 173:1007-15 (1991)*).

A representative list of predicted B18Ag1 B-cell and T-cell epitopes, broken down according to predicted HLA Class I MHC binding antigen, is shown below:

Predicted Th Motifs (B-cell epitopes) (SEQ ID NOS.: 131-133)

SSGGRTFDDFHRYLLVGI
 QGAAQKPINLSKXIEVVQGHDE
 SPGVFLEHLQEAYRIYTPFDLSA

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Predicted HLA A2.1 Motifs (T-cell epitopes) (SEQ ID NOS.: 134-140)

YLLVGIQGA
 GAAQKPINL
 NLSKXIEVV
 EVVQGHDES
 HLQEAYRIY
 NLAQVAQAA
 FVAQAAPDS

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EXAMPLE 5

IDENTIFICATION OF T-CELL EPITOPES OF B11Ag1

This Example illustrates the identification of B11Ag1 (also referred to as B305D) epitopes. Four peptides, referred to as B11-8, B11-1, B11-5 and B11-12 (SEQ ID NOS:309-312, respectfully) were derived from the B11Ag1 gene.

Human CD8 T cells were primed *in vitro* to the peptide B11-8 using dendritic cells according to the protocol of Van Tsai et al. (*Critical Reviews in Immunology* 18:65-75, 1998). The resulting CD8 T cell cultures were tested for their ability to recognize the B11-8 peptide or a negative control peptide, presented by the B-LCL line, JY. Briefly, T cells were incubated with autologous monocytes in the presence of 10 ug/ml peptide, 10 ng/ml IL-7 and 10 ug/ml IL-2, and assayed for their ability to specifically lyse target cells in a standard 51-Cr release assay. As shown in Fig. 22, the bulk culture line demonstrated strong recognition of the B11-8 peptide with weaker recognition of the peptide B11-1.

A clone from this CTL line was isolated following rapid expansion using the monoclonal antibody OKT3 and human IL-2. As shown in Fig. 23, this clone (referred to as A1), in addition to being able to recognize specific peptide, recognized JY LCL transduced with the B11Ag1 gene. This data demonstrates that B11-8 is a naturally
 5 processed epitope of the B11Ag1 gene. In addition these T cells were further found to recognize and lyse, in an HLA-A2 restricted manner, an established tumor cell line naturally expressing B11Ag1 (Fig. 24). The T cells strongly recognize a lung adenocarcinoma (LT-140-22) naturally expressing B11Ag1 transduced with HLA-A2, as well as an A2+ breast carcinoma (CAMA-1) transduced with B11Ag1, but not
 10 untransduced lines or another negative tumor line (SW620).

These data clearly demonstrate that these human T cells recognize not only B11-specific peptides but also transduced cells, as well as naturally expressing tumor lines.

CTL lines raised against the antigens B11-5 and B11-12, using the procedures described above, were found to recognize corresponding peptide-coated targets.
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EXAMPLE 6

CHARACTERIZATION OF BREAST TUMOR GENES DISCOVERED BY DIFFERENTIAL DISPLAY PCR

5 The specificity and sensitivity of the breast tumor genes discovered by differential display PCR were determined using RT-PCR. This procedure enabled the rapid evaluation of breast tumor gene mRNA expression semiquantitatively without using large amounts of RNA. Using gene specific primers, mRNA expression levels in a variety of tissues were examined, including 8 breast tumors, 5 normal breasts, 2 prostate tumors, 2
10 colon tumors, 1 lung tumor, and 14 other normal adult human tissues, including normal prostate, colon, kidney, liver, lung, ovary, pancreas, skeletal muscle, skin, stomach and testes.

To ensure the semiquantitative nature of the RT-PCR, β -actin was used as internal control for each of the tissues examined. Serial dilutions of the first strand cDNAs
15 were prepared and RT-PCR assays performed using β -actin specific primers. A dilution was then selected that enabled the linear range amplification of β -actin template, and which was sensitive enough to reflect the difference in the initial copy number. Using this condition, the β -actin levels were determined for each reverse transcription reaction from each tissue. DNA contamination was minimized by DNase treatment and by assuring a
20 negative result when using first strand cDNA that was prepared without adding reverse transcriptase.

Using gene specific primers, the mRNA expression levels were determined in a variety of tissues. To date, 38 genes have been successfully examined by RT-PCR, five of which exhibit good specificity and sensitivity for breast tumors (B15AG-1,
25 B31GA1b, B38GA2a, B11A1a and B18AG1a). Figures 21A and 21B depict the results for three of these genes: B15AG-1 (SEQ ID NO:27), B31GA1b (SEQ ID NO:148) and B38GA2a (SEQ ID NO:157). Table I summarizes the expression level of all the genes tested in normal breast tissue and breast tumors, and also in other tissues.

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The breast tumor antigen expressed in an *E. coli* recombinant expression system was grown overnight in LB broth with the appropriate antibiotics at 37 °C in a shaking incubator. The next morning, 10 ml of the overnight culture was added to 500 ml to 2x YT plus appropriate antibiotics in a 2L-baffled Erlenmeyer flask. When the Optical Density (at 560 nm) of the culture reached 0.4-0.6, the cells were induced with IPTG (1 mM). Four hours after induction with IPTG, the cells were harvested by centrifugation. The cells were then washed with phosphate buffered saline and centrifuged again. The supernatant was discarded and the cells were either frozen for future use or immediately processed. Twenty ml of lysis buffer was added to the cell pellets and vortexed. To break

open the *E. coli* cells, this mixture was then run through the French Press at a pressure of 16,000 psi. The cells were then centrifuged again and the supernatant and pellet were checked by SDS-PAGE for the partitioning of the recombinant protein. For proteins that localized to the cell pellet, the pellet was resuspended in 10 mM Tris pH 8.0, 1% CHAPS and the inclusion body pellet was washed and centrifuged again. This procedure was repeated twice more. The washed inclusion body pellet was solubilized with either 8 M urea or 6 M guanidine HCl containing 10 mM Tris pH 8.0 plus 10 mM imidazole. The solubilized protein was added to 5 ml of nickel-chelate resin (Qiagen) and incubated for 45 min to 1 hour at room temperature with continuous agitation. After incubation, the resin and protein mixture were poured through a disposable column and the flow through was collected. The column was then washed with 10-20 column volumes of the solubilization buffer. The antigen was then eluted from the column using 8M urea, 10 mM Tris pH 8.0 and 300 mM imidazole and collected in 3 ml fractions. A SDS-PAGE gel was run to determine which fractions to pool for further purification.

As a final purification step, a strong anion exchange resin such as HiPrepQ (Biorad) was equilibrated with the appropriate buffer and the pooled fractions from above were loaded onto the column. Antigen was eluted off the column with a increasing salt gradient. Fractions were collected as the column was run and another SDS-PAGE gel was run to determine which fractions from the column to pool. The pooled fractions were dialyzed against 10 mM Tris pH 8.0. The protein was then vialled after filtration through a 0.22 micron filter and the antigens were frozen until needed for immunization.

Four hundred micrograms of B305D antigen was combined with 100 micrograms of muramyl dipeptide (MDP). Every four weeks rabbits were boosted with 100 micrograms mixed with an equal volume of Incomplete Freund's Adjuvant (IFA). Seven days following each boost, the animal was bled. Sera was generated by incubating the blood at 4 °C for 12-24 hours followed by centrifugation.

Ninety-six well plates were coated with B305D antigen by incubating with 50 microliters (typically 1 microgram) of recombinant protein at 4 °C for 20 hours. 250 microliters of BSA blocking buffer was added to the wells and incubated at room

temperature for 2 hours. Plates were washed 6 times with PBS/0.01% Tween. Rabbit sera was diluted in PBS. Fifty microliters of diluted sera was added to each well and incubated at room temperature for 30 min. Plates were washed as described above before 50 microliters of goat anti-rabbit horse radish peroxidase (HRP) at a 1:10000 dilution was added and incubated at room temperature for 30 min. Plates were again washed as described above and 100 microliters of TMB microwell peroxidase substrate was added to each well. Following a 15 min incubation in the dark at room temperature, the colorimetric reaction was stopped with 100 microliters of 1N H₂SO₄ and read immediately at 450 nm. The polyclonal antibodies showed immunoreactivity to B305D.

- 10 Immunohistochemical (IHC) analysis of B305D expression in breast cancer and normal breast specimens was performed as follows. Paraffin-embedded formal fixed tissue was sliced into 8 micron sections. Steam heat induced epitope retrieval (SHIER) in 0.1 M sodium citrate buffer (pH 6.0) was used for optimal staining conditions. Sections were incubated with 10% serum/PBS for 5 minutes. Primary antibody was added to each section for 25 min at indicated concentrations followed by a 25 min incubation with either an anti-rabbit or anti-mouse biotinylated antibody. Endogenous peroxidase activity was blocked by three 1.5 min incubations with hydrogen peroxide. The avidin biotin complex/horseradish peroxidase (ABC/HRP) systems was used along with DAB chromagen to visualize antigen expression. Slides were counterstained with hematoxylin.
- 15 B305D expression was detected in both breast tumor and normal breast tissue. However, the intensity of staining was much less in normal samples than in tumor samples and surface expression of B305D was observed only in breast tumor tissues.
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- A summary of real-time PCR and immunohistochemical analysis of B305D expression in an extensive panel of normal tissues is presented in Table II below. These results demonstrate minimal expression of B305D in testis, inconclusive results in gall bladder, and no detection in all other tissues tested.
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TABLE II

mRNA	IHC staining	Tissue type	Summary
Moderately positive	Positive	Testis	Nuclear staining of small minority of spermatids; spermatozoa negative; seminoma negative
Negative	Negative	Thymus	No expression
N/A	Negative	Artery	No expression
Negative	Negative	Skeletal muscle	No expression
Negative	Positive (weak staining)	Small bowel	No expression
Negative	Positive (weak staining)	Ovary	No expression
Negative		Pituitary	No expression
Negative	Positive (weak staining)	Stomach	No expression
Negative	Negative	Spinal cord	No expression
Negative	Negative	Spleen	No expression
Negative	Negative	Ureter	No expression
N/A	Negative	Gall bladder	Inconclusive
N/A	Negative	Placenta	No expression
Negative	Negative	Thyroid	No expression
Negative	Negative	Heart	No expression
Negative	Negative	Kidney	No expression
Negative	Negative	Liver	No expression
Negative	Negative	Brain-cerebellum	No expression
Negative	Negative	Colon	No expression
Negative	Negative	Skin	No expression
Negative	Negative	Bone marrow	No expression
N/A	Negative	Parathyroid	No expression
Negative	Negative	Lung	No expression
Negative	Negative	Esophagus	No expression
Negative	Positive (weak staining)	Uterus	No expression
Negative	Negative	Adrenal	No expression
Negative	Negative	Pancreas	No expression
N/A	Negative	Lymph node	No expression
Negative	Negative	Brain-cortex	No expression
N/A	Negative	Fallopian tube	No expression
Negative	Positive (weak staining)	Bladder	No expression
Negative	N/A	Bone	No expression
Negative	N/A	Salivary gland	No expression
Negative	N/A	Activated PBMC	No expression
Negative	N/A	Resting PBMC	No expression

Negative	N/A	Trachea	No expression
Negative	N/A	Vena cava	No expression
Negative	N/A	Retina	No expression
Negative	N/A	Cartilage	No expression

EXAMPLE 8

PROTEIN EXPRESSION OF BREAST TUMOR ANTIGENS

5 This example describes the expression and purification of the breast tumor antigen B305D in *E. coli* and in mammalian cells.

Expression of B305D isoform C-15 (SEQ ID NO:301; translated to 384 amino acids) in *E. coli* was achieved by cloning the open reading frame of B305D isoform C-15 downstream of the first 30 amino acids of the *M. tuberculosis* antigen Ra12 (SEQ ID NO:318) in pET17b. First, the internal EcoRI site in the B305D ORF was mutated without changing the protein sequence so that the gene could be cloned at the EcoRI site with Ra12. The PCR primers used for site-directed mutagenesis are shown in SEQ ID NO:319 (referred to as AW012) and SEQ ID NO:320 (referred to as AW013). The ORF of EcoRI site-modified B305D was then amplified by PCR using the primers AW014 (SEQ ID NO:321) and AW015 (SEQ ID NO:322). The PCR product was digested with EcoRI and ligated to the Ra12/pET17b vector at the EcoRI site. The sequence of the resulting fusion construct (referred to as Ra12mB11C) was confirmed by DNA sequencing. The determined cDNA sequence for the fusion construct is provided in SEQ ID NO:323, with the amino acid sequence being provided in SEQ ID NO:324.

20 The fusion construct was transformed into BL21(DE3)CodonPlus-RIL *E. coli* (Stratagene) and grown overnight in LB broth with kanamycin. The resulting culture was induced with IPTG. Protein was transferred to PVDF membrane and blocked with 5% non-fat milk (in PBS-Tween buffer), washed three times and incubated with mouse anti-His tag antibody (Clontech) for 1 hour. The membrane was washed 3 times and probed with HRP-Protein A (Zymed) for 30 min. Finally, the membrane was washed 3 times and developed with ECL (Amersham). Expression was detected by Western blot.

For recombinant expression in mammalian cells, B305D isoform C-15 (SEQ ID NO:301; translated to 384 amino acids) was subcloned into the mammalian expression vectors pCEP4 and pcDNA3.1 (Invitrogen). These constructs were transfected into HEK293 cells (ATCC) using Fugene 6 reagent (Roche). Briefly, the HEK cells were
 5 plated at a density of 100,000 cells/ml in DMEM (Gibco) containing 10% FBS (Hyclone) and grown overnight. The following day, 2 ul of Fugene 6 was added to 100 ul of DMEM containing no FBS and incubated for 15 minutes at room temperature. The Fugene 6/DMEM mixture was added to 1 ug of B305D/pCEP4 or B305D/pcDNA plasmid DNA and incubated for 15 minutes at room temperature. The Fugene/DNA mix was then added
 10 to the HEK293 cells and incubated for 48-72 hours at 37 °C with 7% CO₂. Cells were rinsed with PBS, the collected and pelleted by centrifugation.

For Western blot analysis, whole cell lysates were generated by incubating the cells in Triton-X100 containing lysis buffer for 30 minutes on ice. Lysates were then cleared by centrifugation at 10,000 rpm for 5 minutes at 4 °C. Samples were diluted with
 15 SDS_PAGE loading buffer containing beta-mercaptoethanol, and boiled for 10 minutes prior to loading the SDS_PAGE gel. Proteins were transferred to nitrocellulose and probed using Protein A purified anti-B305D rabbit polyclonal sera (prepared as described above) at a concentration of 1 ug/ml. The blot was revealed with a goat anti-rabbit Ig coupled to HRP followed by incubation in ECL substrate. Expression of B305D was detected in the
 20 the HEK293 lysates transfected with B305D, but not in control HEK293 cells transfected with vector alone.

For FACS analysis, cells were washed further with ice cold staining buffer and then incubated with a 1:100 dilution of a goat anti-rabbit Ig (H+L)-FITC reagent (Southern Biotechnology) for 30 minutes on ice. Following 3 washes, the cells were
 25 resuspended in staining buffer containing Propidium Iodide (PI), a vital stain that allows for identification of permeable cells, and then analyzed by FACS. The FACS analysis showed surface expression of B305D protein.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

CLAIMS

What is Claimed:

1. An isolated polynucleotide comprising a sequence selected from the group consisting of:
 - (a) sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (b) complements of the sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (d) sequences that hybridize to a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, under moderately stringent conditions;
 - (e) sequences having at least 75% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (f) sequences having at least 90% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325; and
 - (g) degenerate variants of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

2. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:
 - (a) sequences encoded by a polynucleotide of claim 1; and
 - (b) sequences having at least 70% identity to a sequence encoded by a polynucleotide of claim 1; and

(c) sequences having at least 90% identity to a sequence encoded by a polynucleotide of claim 1.

3. An expression vector comprising a polynucleotide of claim 1 operably linked to an expression control sequence.

4. A host cell transformed or transfected with an expression vector according to claim 3.

5. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a polypeptide of claim 2.

6. A method for detecting the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with a binding agent that binds to a polypeptide of claim 2;
- (c) detecting in the sample an amount of polypeptide that binds to the binding agent; and
- (d) comparing the amount of polypeptide to a predetermined cut-off value and therefrom determining the presence of a cancer in the patient.

7. A fusion protein comprising at least one polypeptide according to claim 2.

8. An oligonucleotide that hybridizes to a sequence recited in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325 under moderately stringent conditions.

9. A method for stimulating and/or expanding T cells specific for a tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1; and
- (c) antigen-presenting cells that express a polypeptide according to claim 1,

under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

10. An isolated T cell population, comprising T cells prepared according to the method of claim 9.

11. A composition comprising a first component selected from the group consisting of physiologically acceptable carriers and immunostimulants, and a second component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1;
- (c) antibodies according to claim 5;
- (d) fusion proteins according to claim 7;
- (e) T cell populations according to claim 10; and
- (f) antigen presenting cells that express a polypeptide according to claim 2.

12. A method for stimulating an immune response in a patient, comprising administering to the patient a composition of claim 11.

13. A method for the treatment of a cancer in a patient, comprising administering to the patient a composition of claim 11.

14. A method for determining the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with an oligonucleotide according to claim 8;
- (c) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and
- (d) compare the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence of the cancer in the patient.

15. A diagnostic kit comprising at least one oligonucleotide according to claim 8.

16. A diagnostic kit comprising at least one antibody according to claim 5 and a detection reagent, wherein the detection reagent comprises a reporter group.

17. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

- (a) incubating CD4+ and/or CD8+ T cells isolated from a patient with at least one component selected from the group consisting of: (i) polypeptides according to claim 2; (ii) polynucleotides according to claim 1; and (iii) antigen presenting cells that express a polypeptide of claim 2, such that T cell proliferate;
- (b) administering to the patient an effective amount of the proliferated T cells,

and thereby inhibiting the development of a cancer in the patient.

COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF
BREAST CANCER

ABSTRACT OF THE DISCLOSURE

Compositions and methods for the therapy and diagnosis of cancer, particularly breast cancer, are disclosed. Illustrative compositions comprise one or more breast tumor polypeptides, immunogenic portions thereof, polynucleotides that encode such polypeptides, antigen presenting cell that expresses such polypeptides, and T cells that are specific for cells expressing such polypeptides. The disclosed compositions are useful, for example, in the diagnosis, prevention and/or treatment of diseases, particularly breast cancer.

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cDNA PREPARED FROM
NORMAL BREAST TISSUE
FROM THE SAME PATIENT

cDNA PREPARED
FROM BREAST TUMOR



Fig. 1

BREAST TUMOR mRNA
NORMAL BREAST TISSUE mRNA

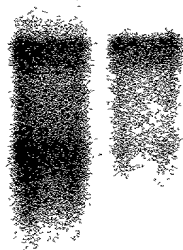


Fig. 2

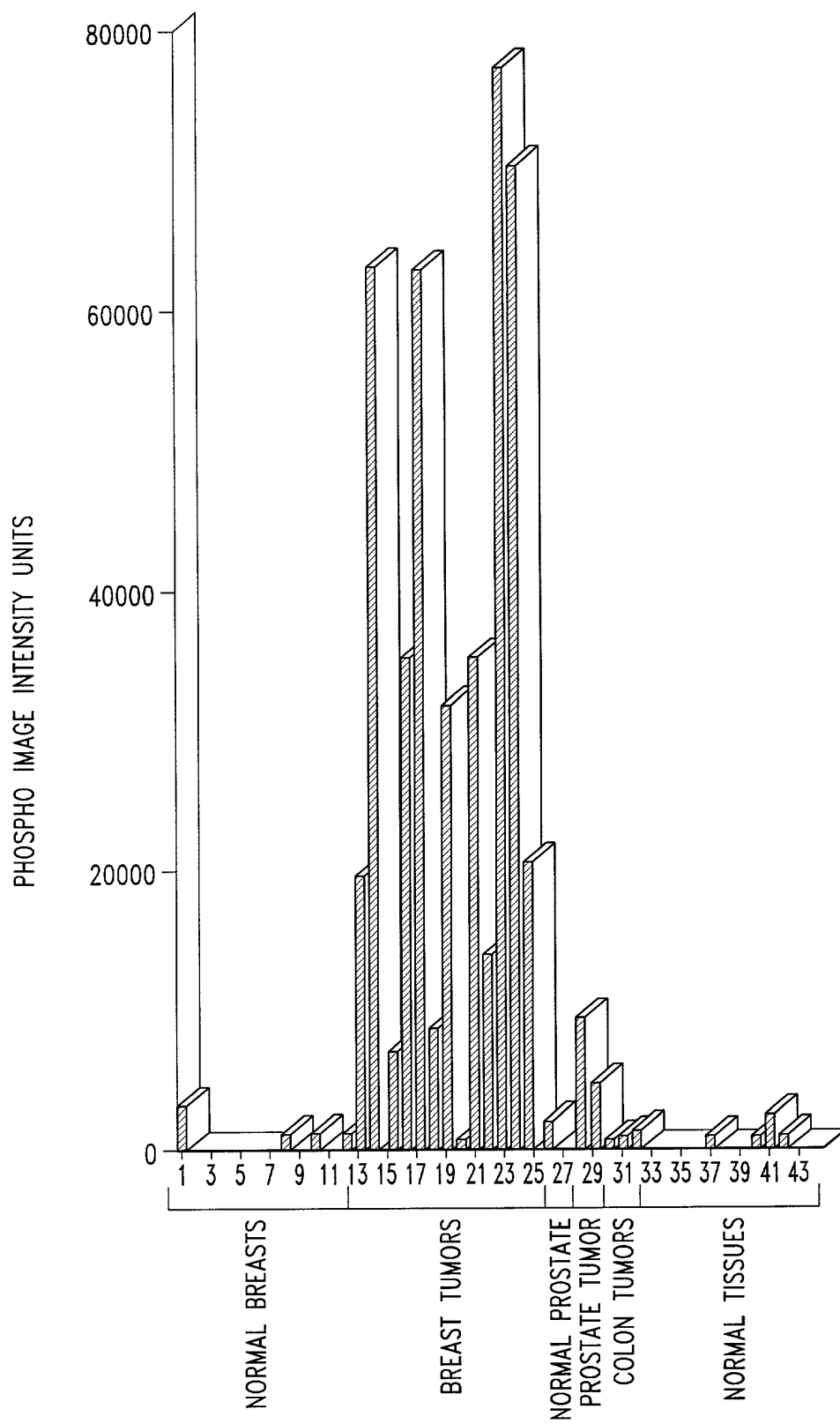


Fig. 3

GENOMIC CLONE MAP

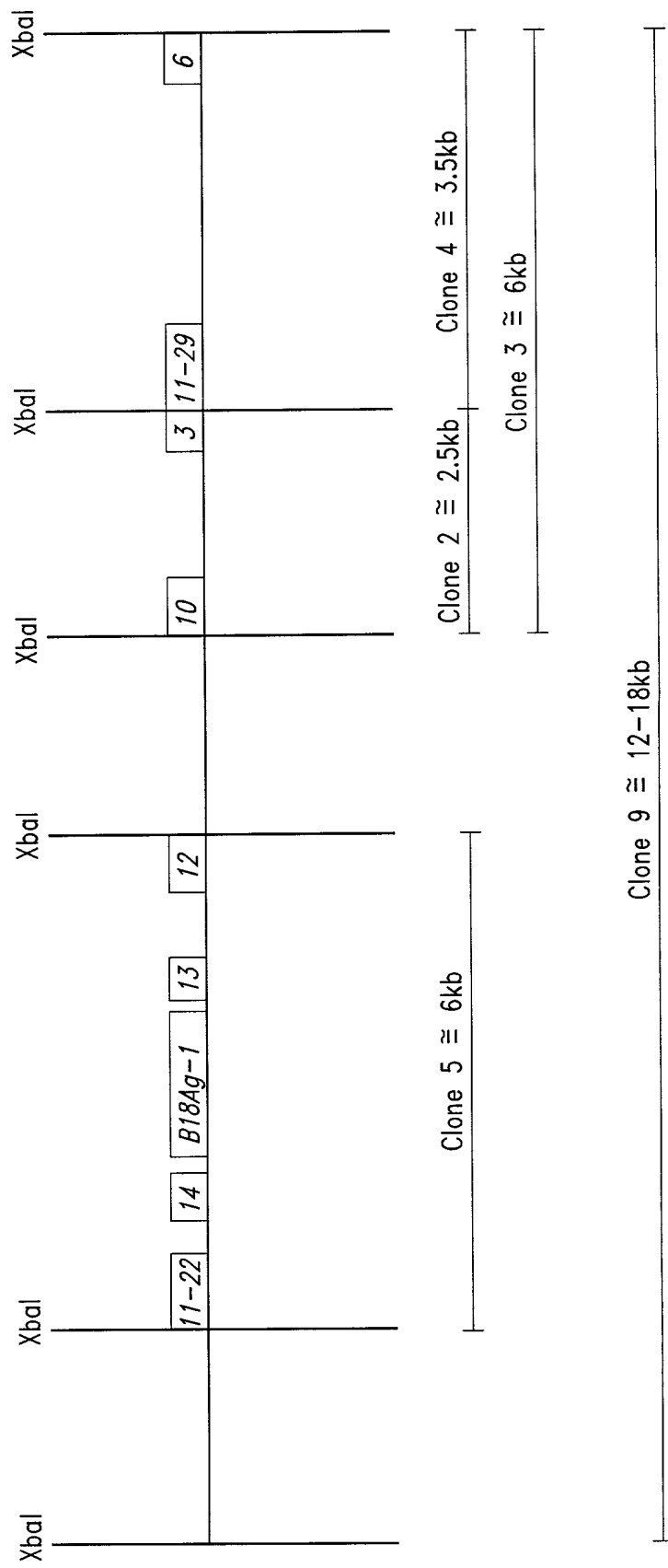


Fig. 4

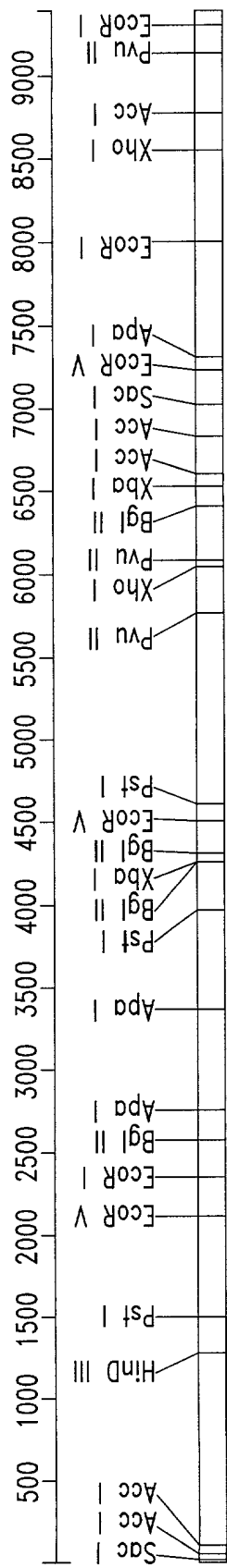


Fig. 5A

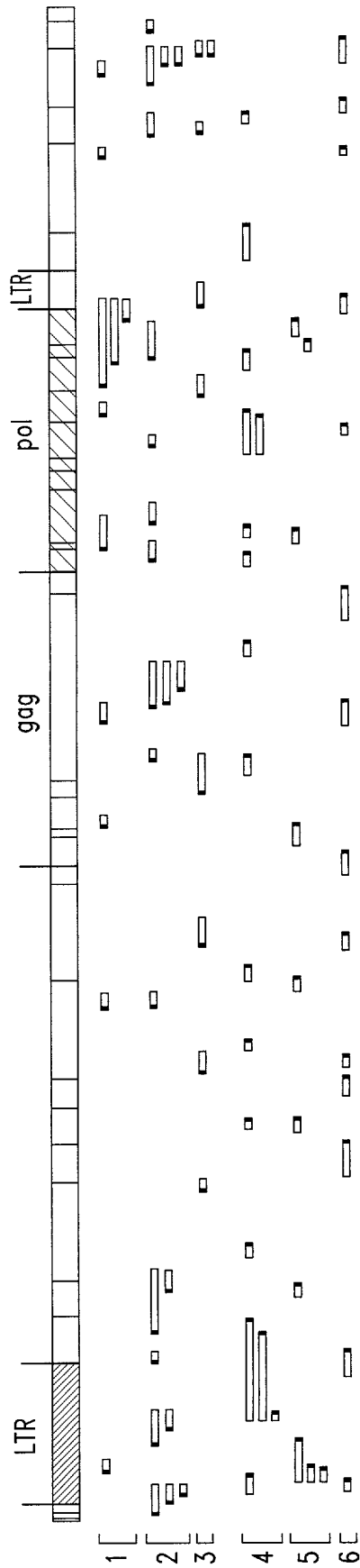


Fig. 5B

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B18Ag1

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1				5					10					15		
GGG	AGA	ACT	TTT	GAC	GAT	TTC	CAC	CGG	TAT	CTC	CTC	GTG	GGT	ATT	CAG	96
Gly	Arg	Thr	Phe	Asp	Asp	Phe	His	Arg	Tyr	Leu	Leu	Val	Gly	Ile	Gln	
			20					25					30			
GGA	GCT	GCC	CAG	AAA	CCT	ATA	AAC	TTG	TCT	AAG	GCG	ATT	GAA	GTC	GTC	144
Gly	Ala	Ala	Gln	Lys	Pro	Ile	Asn	Leu	Ser	Lys	Ala	Ile	Glu	Val	Val	
		35					40					45				
CAG	GGG	CAT	GAT	GAG	TCA	CCA	GGA	GTG	TTT	TTA	GAG	CAC	CTC	CAG	GAG	192
Gln	Gly	His	Asp	Glu	Ser	Pro	Gly	Val	Phe	Leu	Glu	His	Leu	Gln	Glu	
	50					55					60					
GCT	TAT	CGG	ATT	TAC	ACC	CCT	TTT	GAC	CTG	GCA	GCC	CCC	GAA	AAT	AGC	240
Ala	Tyr	Arg	Ile	Tyr	Thr	Pro	Phe	Asp	Leu	Ala	Ala	Pro	Glu	Asn	Ser	
65					70				75					80		
CAT	GCT	CTT	AAT	TTG	GCA	TTT	GTG	GCT	CAG	GCA	GCC	CCA	GAT	AGT	AAA	288
His	Ala	Leu	Asn	Leu	Ala	Phe	Val	Ala	Gln	Ala	Ala	Pro	Asp	Ser	Lys	
			85						90					95		
AGG	AAA	CTC	CAA	AAA	CTA	GAG	GGA	TTT	TGC	TGG	AAT	GAA	TAC	CAG	TCA	336
Arg	Lys	Leu	Gln	Lys	Leu	Glu	Gly	Phe	Cys	Trp	Asn	Glu	Tyr	Gln	Ser	
			100					105					110			
GCT	TTT	AGA	GAT	AGC	CTA	AAA	GGT	TTT								363
Ala	Phe	Arg	Asp	Ser	Leu	Lys	Gly	Phe								
		115					120									

Fig. 6

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B17Ag1

GC TGGGCACAGT GGCTCATACC TGTAATCCTG ACCGTTTCAG AGGCTCAGGT	60
CG CTTGAGCCCA AGATTTC AAG ACTAGTCTGG GTAACATAGT GAGACCCTAT	120
AA AAATAAAAAA ATGAGCCTGG TGTAGTGGCA CACACCAGCT GAGGAGGGAG	180
CT AGGAGA	196

Fig. 7

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B17Ag2

GC TTGGGGGCTC TGACTAGAAA TTCAAGGAAC CTGGGATTCA AGTCCAACTG	60
AC TTACACTGTG GNCTCCAATA AACTGCTTCT TTCCTATTCC CTCTCTATTA	120
AA GGAAAACGAT GTCTGTGTAT AGCCAAGTCA GNTATCCTAA AAGGAGATAC	180
AT TAAATATCAG AATGTAAAAC CTGGGAACCA GGTTCCTCAGC CTGGGATTAA	240
CA AGAAGACTGA ACAGTACTAC TGTGAAAAGC CCGAAGNGGC AATATGTTCA	300
TT GAAGGATGGC TGGGAGAATG AATGCTCTGT CCCCCAGTCC CAAGCTCACT	360
CT CCTTTATAGC CTAGGAGA	388

Fig. 8

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag2a

GC CTATAATCAT GTTTCTCATT ATTTTCACAT TTTATTAACC AATTTCTGTT	60
AA AATATGAGGG AAATATATGA AACAGGGAGG CAATGTTTCAG ATAATTGATC	120
TG ATTTCTACAT CAGATGCTCT TTCCTTTCCT GTTTATTTCC TTTTATTTTC	180
GG TCGAATGTAA TAGCTTTGTT TCAAGAGAGA GTTTTGGCAG TTTCTGTAGC	240
CT GCTCATGTCT CCAGGCATCT ATTTGCACTT TAGGAGGTGT CGTGGGAGAC	300
CT ATTTTTTCCA TATTTGGGCA ACTACTA	337

Fig. 9

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag1b

GC CATACAGTGC CTTTCCATTT ATTTAACCCC CACCTGAACG GCATAAACTG	60
GC TGGTGTTTTT TACTGTAAAC AATAAGGAGA CTTTGCTCTT CATTTAAACC	120
AT TTCATATTTT ACGCTCGAGG GTTTTTACCG GTTCCTTTTT ACACTCCTTA	180
TT TAAGTCGTTT GGAACAAGAT ATTTTTTCTT TCCTGGCAGC TTTTAACATT	240
TT TGTGTCTGGG GGACTGCTGG TCACTGTTTC TCACAGTTGC AAATCAAGGC	300
CC AAGAAAAAAAA AATTTTTTTG TTTTATTTGA AACTGGACCG GATAAACGGT	360
CG GCTGCTGTAT ATAGTTTTAA ATGGTTTATT GCACCTCCTT AAGTTGCACT	420
GG GGGGNTTTTG NATAGAAAGT NTTTANTCAC ANAGTCACAG GGACTTTTNT	480
NA CTGAGCTAAA AAGGGCTGNT TTTCGGGTGG GGGCAGATGA AGGCTCACAG	540
TC TCTTAGAGGG GGGAACTNCT A	571

Fig. 10

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag1a

TA ATAACTTAAA TATATTTTGA TCACCCACTG GGGTGATAAG ACAATAGATA	60
TT TCCAAAAAGC ATAAAACCAA AGTATCATAC CAAACCAAAT TCATACTGCT	120
CC GCACTGAAAC TTCACCTTCT AACTGTCTAC CTAACCAAAT TCTACCCTTC	180
GG TGC GTGCTCA C TACTCTTTT TTTTTTTTTT TTTNTTTTGG AGATGGAGTC	240
CA GCCCAGGGGT GGAGTACAAT GGCACAACCT CAGCTCACTG NAACCTCCGC	300
TT CATGAGATTC TCCTGNTTCA GCCTTCCCAG TAGCTGGGAC TACAGGTGTG	360
TG CCTGGNTAAT CTTTTTTNGT TTTNGGGTAG AGATGGGGGT TTTACATGTT	420
TG GTNTCGAACT CCTGACCTCA AGTGATCCAC CCACCTCAGG CTCCCAAAGT	480
TA CAGACATGAG CCACTGNGCC CAGNCCTGGT GCATGCTCAC TTCTCTAGGC	540
	548

Fig. 11

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B11Ag1

TG CACATGCAGA ATATTCTATC GGTACTTCAG CTATTACTCA TTTTGATGGC	60
AG CCTATCCTCA AGATGAGTAT TTAGAAAGAA TTGATTTAGC GATAGACCAA	120
GC ACTCTGACTA CACGAAATTG TTCAGATGTG ATGGATTTAT GACAGTTGAT	180
GA GATTATTAAG TGATTATTTT AAAGGGAATC CATTAAATTCC AGAATATCTT	240
TC AAGATGATAT AGAAATAGAA CAGAAAGAGA CTACAAATGA AGATGTATCA	300
TA TTGAAGAGCC TATAGTAGAA AATGAATTAG CTGCATTTAT TAGCCTTACA	360
TT TTCCTGATGA ATCTTATATT CAGCCATCGA CATAGCATTG CCTGATGGGC	420
GA ATAATAGAAA CTGGGTGCGG GGCTATTGAT GAATTCATCC NCAGTAAATT	480
AC AAAATATAAC TCGATTGCAT TTGGATGATG GAATACTAAA TCTGGCAAAA	540
GG AGCTACTAGT AACCTCTCTT TTTGAGATGC AAAATTTTCT TTTAGGGTTT	600
CT ACTTTACGGA TATTGGAGCA TAACGGGA	638

Fig. 12

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA3c

ACTGATGGAT GTCGCCGGAG GCGAGGGGCC TTATCTGATG CTCGGCTGCC TGTTCGTGAT 60
GTGCGCGGCG ATTGGGCTGT TTATCTCAAA CACCGCCACG GCGGTGCTGA TGGCGCCTAT 120
TGCCTTAGCG GCGGCGAAGT CAATGGGCGT CTCACCCTAT CCTTTTGCCA TGGTGGTGGC 180
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TGGCCCTGGC AAGTACTCAT TTAGCGATTT TGTCAAATA GGCGTG 286

Fig. 13

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B9CG1

AG CAGCCCCTTC TTCTCAATTT CATCTGTCAC TACCCTGGTG TAGTATCTCA	60
CA TTTTATAGC CTCCTCCCTG GTCTGTCTTT TGATTTTCCT GCCTGTAATC	120
AC ATAAGTGCAA GTAAACATTT CTAAAGTGTG GTTATGCTCA TGTCACCTCCT	180
AA ATAGTTTCCA TTACCGTCTT AATAAAATTC GGATTTGTTC TTTNCTATTN	240
CA CCTATGACCG AA	262

Fig. 14

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B9CG3

AG CAAAGCCAGT GGTTTGAGCT CTCTACTGTG TAAACTCCTA AACCAAGGCC	60
TA AATGGTGGCA GGATTTTAT TATAACATG TACCCATGCA AATTTCTAT	120
GA TATATTCTTC TACATTTAAA CAATAAAAAT AATCTATTTT TAAAAGCCTA	180
AG TTAGGTAAGA GTGTTTAATG AGAGGGTATA AGGTATAAAT CACCAGTCAA	240
TG CCTATGACCG A	261

Fig. 15

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B2CA2

GG GCATGGACGC AGACGCCTGA CGTTTGGCTG AAAATCTTTC ATTGATTCGT	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCTGT TGCTCGCCAG TTTTNTGTT	120
GG ANAAGGCAAN GAGCTCTTCA GACTATTGGN ATTNTCGTTC GGTCTTCTGC	180
CG NCTTGCAANG ATCTTCAT	208

Fig. 16

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA1

GG GCATGGACGC AGACGCCTGA CGTTTGGCTG AAAATCTTTC ATTGATTCGT	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCTGT TGCTCGCCAG TTTTNTGTT	120
GG ANAAGGCAAN GAGCTCTTCA GACTATTGGN ATTNTCGTTC GGTCTTCTGC	180
CG NCTTGCNANG ATCTTCAT	208

Fig. 17

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA2

GG GCATGGACGC AGACGCCTGA CGTTTGGCTG AAAATCTTTC ATTGATTCTG	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCTGT TGCTCGCCAG TTTTNTGTT	120
GG ANAAGGCAAN GAGCTCTTCA GACTATTGGN ATTNTCGTTC GGTCTTCTGC	180
CG NCTTGCAANG ATCTTCAT	208

Fig. 18

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA3

AG GGAGCAAGGA GAAGGCATGG AGAGGCTCAN GCTGGTCCTG GCCTACGACT	60
CT GTCGCCGGGG ATGGTGGAGA ACTGAAGCGG GACCTCCTCG AGGTCCTCCG	120
TC NCCGTCCAGG AGGAGGGTCT TTCCGTGGTC TNGGAGGAGC GGGGGGAGAA	180
TC ATGGTCNACA TCCC	204

Fig. 19

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B4CA1

TC AGGAGCGGGT AGAGTGGCAC CATTGAGGGG ATATTCAAAA ATATTATTTT	60
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CC AATCGCATGG ACATGTTAGA CTTATTTTCT GTTAATGATT NCTATTTTTA	180
GA TTTGAGAAAT TGGTTNTTAT TATATCAATT TTTGGTATTT GTTGAGTTTG	240
GC TTAGTATGTG ACCA	264

Fig. 20

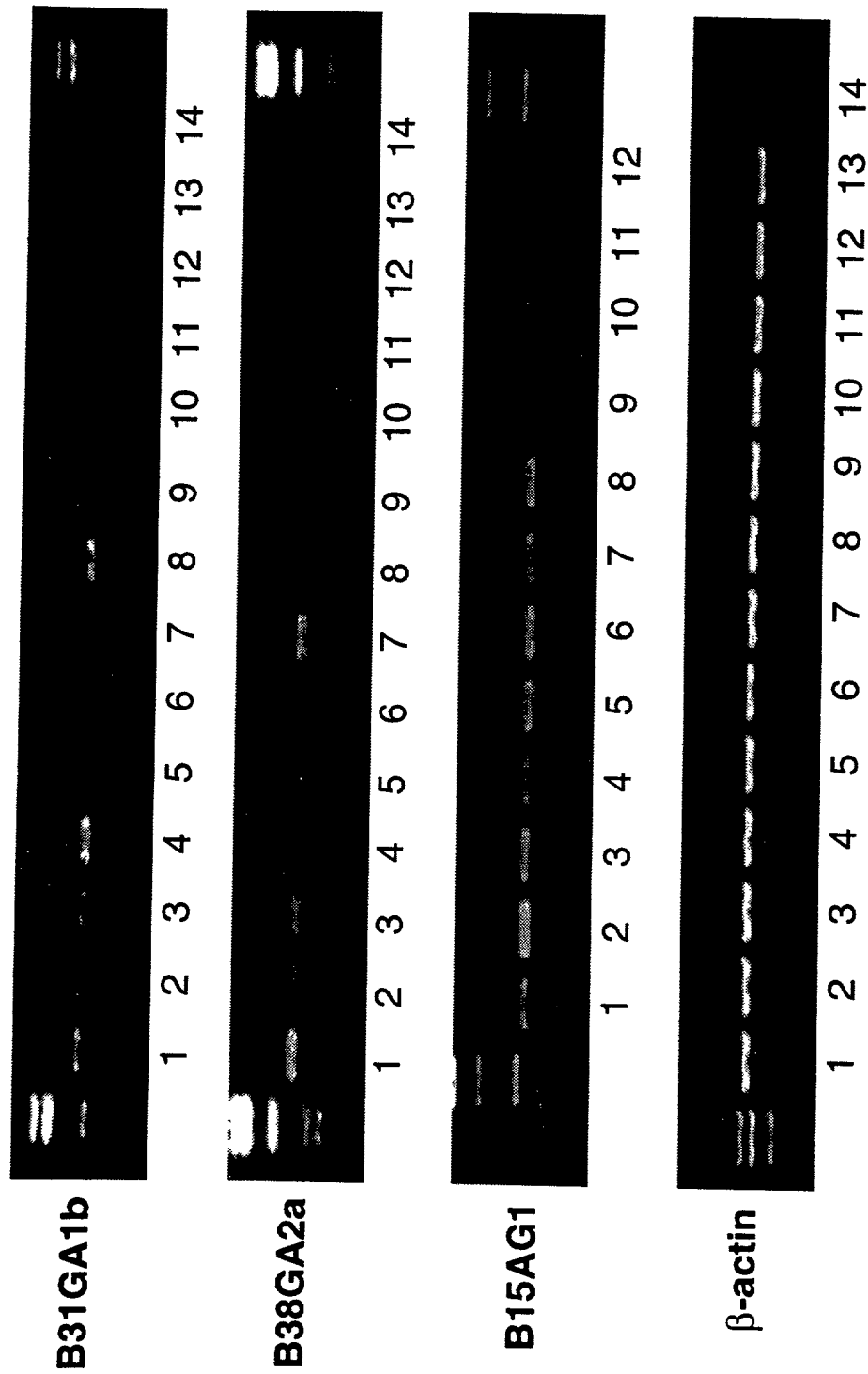


Fig. 21A

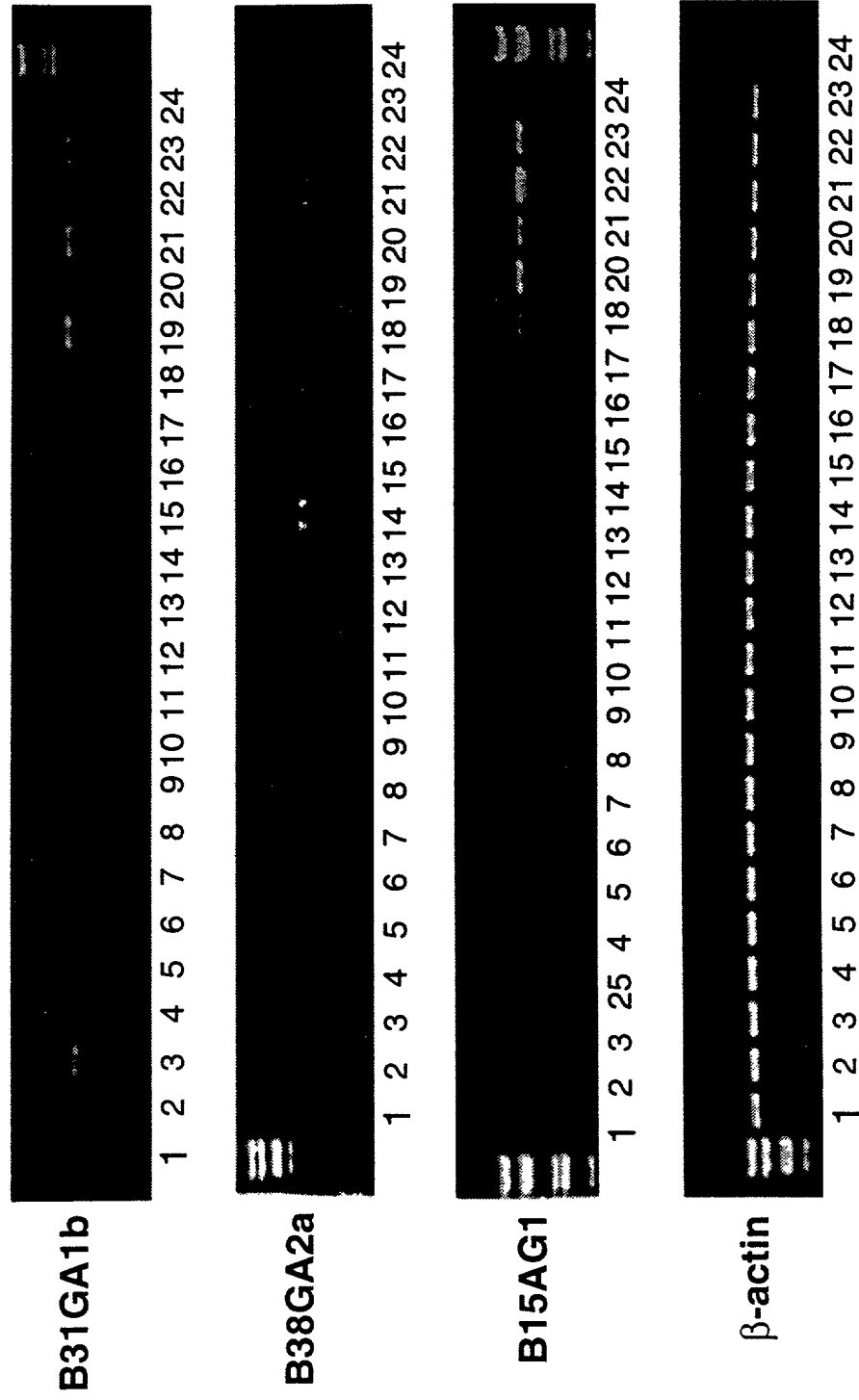


Fig. 21B

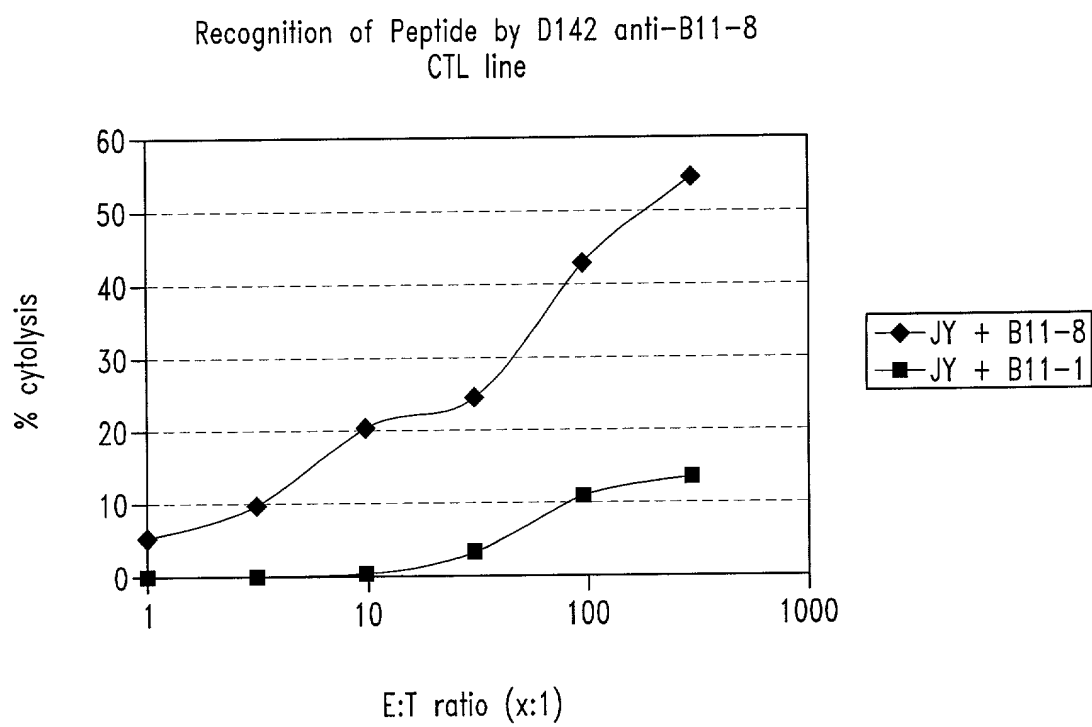


Fig. 22

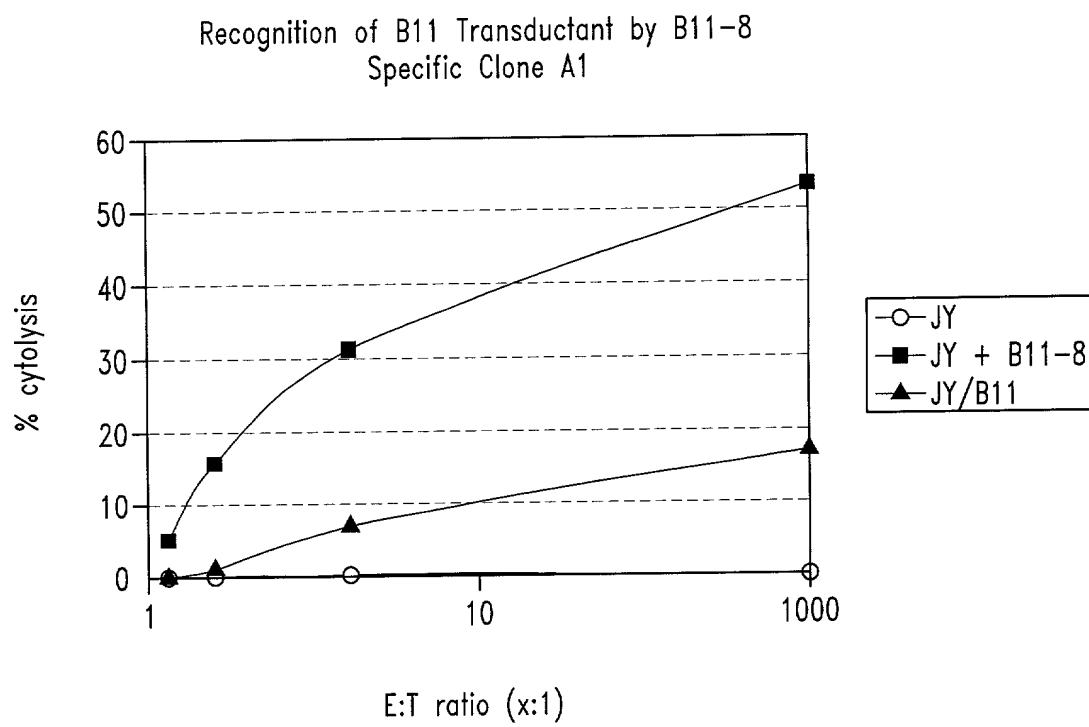


Fig. 23

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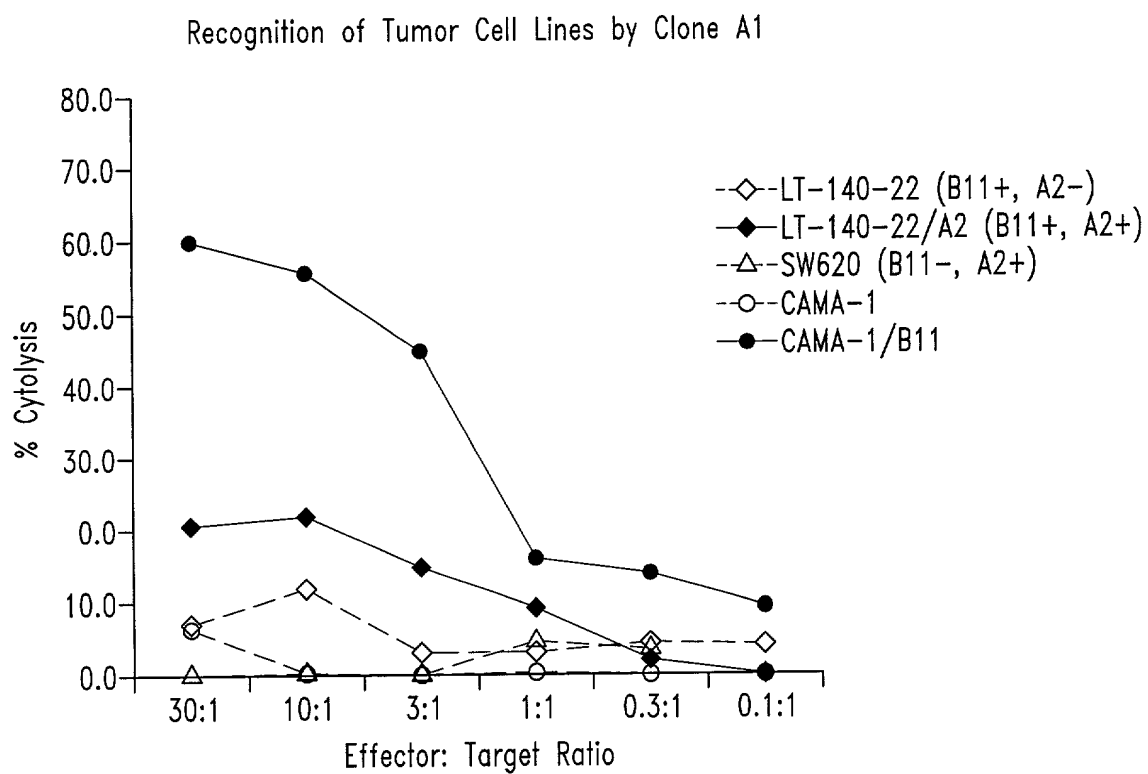


Fig. 24

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants : Tony N. Frudakis et al.
Filed : October 26, 2000
For : COMPOSITIONS AND METHODS FOR THE THERAPY AND
DIAGNOSIS OF BREAST CANCER

Docket No. : 210121.419C10

Date : October 26, 2000

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

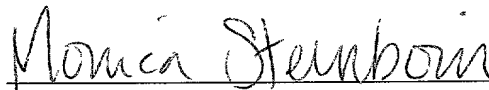
DECLARATION

Sir:

I, Monica Steinborn, in accordance with 37 C.F.R. § 1.821(f) do hereby declare that, to the best of my knowledge, the content of the paper entitled "Sequence Listing" and the computer readable copy contained within the floppy disk are the same.

I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Dated this 26th day of October, 2000.



Monica Steinborn
Biotechnology Paralegal

701 Fifth Avenue, Suite 6300
Seattle, WA 98104-7092
(206) 622-4900
FAX (206) 682-6031

SEQUENCE LISTING

<110> Frudakis, Tony N.
 Reed, Steven G.
 Smith, John M.
 Misher, Linda E.
 Dillon, Davin C.
 Retter, Marc W.
 Wang, Aijun
 Skeiky, Yasir A.W.
 Harlocker, Susan L.

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 Ala Tyr Arg Ile Tyr Thr Pro Phe Asp Leu Ala Ala Pro Glu Asn Ser
 65 70 75 80
 His Ala Leu Asn Leu Ala Phe Val Ala Gln Ala Ala Pro Asp Ser Lys
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 <223> n = A,T,C or G

<400> 4
 tctagagctg cgcttgatc ccgccacagt gaggagacct gaagaccaga gaaaacacag 60
 caagtaggcc ctttaaacta ctcacctgtg ttgtcttcta atttattctg ttttattttg 120

```

tttccatcat ttttaaggggt taaaatcatc ttgttcagac ctcagcatat aaaatgaccc 180
atctgtagac ctcaggctcc aaccataccc caagagttgt ctgggtttgt ttaaattact 240
gccaggtttc agctgcagat atccctggaa ggaatattcc agattccctg agtagtttcc 300
aggttaaaat cctataggct tcttctgttt tgaggaagag ttctgtcag agaaaaacat 360
gattttggat ttttaacttt aatgcttggt aaacgctata aaaaaaattt tctacccta 420
gctttaaagt actgttagtg agaaattaaa attccttcag gaggattaaa ctgccatttc 480
agttacccta attccaaatg ttttggtggt tagaatcttc tttaatgttc ttgaagaagt 540
gttttatatt ttcccatcna gataaattct ctcnncctt nntttntnt ctntttttt 600
aaaacggant cttgctccgt tgtccanct gggaattttt ttttggccaa tctccgctnc 660
cttgcaanaa tncgtcntcc caaaattacc ncctttttcc cactccacc ccnnggaatt 720
acctggaatt anaggccccc ncccccccc cggttaattt gtttttgtt ttagtaaaaa 780
acgggtttcc tgttttagtt aggatggccc anntctgacc cctnatctt cccctcngc 840
cctnaatnt tngnntang gcttaccccc ccnngngtt tttctccat tnaaattttc 900
tntggantct tgaatnncgg gttttccctt ttaaaccnat ttttttttt nnnccccan 960
ttttcctcc cccntntnta angggggtt cccaanccgg gtcnccccc angtcccaa 1020
tttttctccc cccccctctt ttttcttnc cccaaaantc ctatcttttc cttnaaatat 1080
cnantnt 1087

```

```

<210> 5
<211> 1010
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(1010)
<223> n = A,T,C or G

```

```

<400> 5
tctagaccaa gaaatgggag gatttttagag tgactgatga tttctctatc atctgcagtt 60
agtaaacatt ctccacagtt tatgcaaaaa gtaacaaaac cactgcagat gacaaacact 120
aggtaacaca catactatct cccaaatacc taccacaaag ctcaacaatt ttaactggt 180
aggatcactg gctctaata ccatgacatg aggtcaccac caaaccatca agcgctaaac 240
agacagaatg tttccactcc tgatccactg tgtgggaaga agcaccgaac ttaccactg 300
gggggcctgc ntcanaanaa aagcccatgc ccccggtnt ncctttnaac cggaacgaat 360
naaccaccca tccccacanc tctctgttc ntgggccctg catcttggg cctcntntnc 420
tttnggggan acntggggaa ggtaccccat ttcnttgacc cncnanaaaa acccngtgg 480
ccctttgccc tgattcncnt gggccttttc tcttttccct tttgggttgt taaattccc 540
aatgtcccn gaacctctc cntnctgcc aaaacctacc taaattntc nctangnnt 600
ttcttgggtg tncctttcaa aggnaccct ncctgttcan nccnacnaa aattnttcc 660
ntatnntggn cccnaaaaaa nnnatcnnc cnaattgcc gaattggttn ggttttccct 720
netgggggaa accctttaaa tttccccctt ggccggcccc ccttttttcc ccccttnga 780
aggcaggngg ttcttccoga acttccaatt ncaacagecn tgccattgn tgaaacctt 840
ttcctaaaat taaaaaatan cgggttnngg nnggcctct tcccctcng gnggngng 900
aaantcctta cccnaaaaaa ggttgcttag ccccngtcc cactcccc nggaaaaatn 960
aaccttttcn aaaaaaggaa tataantttt ccactccttn gttctcttcc 1010

```

```

<210> 6
<211> 950
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature

```

<222> (1)...(950)

<223> n = A,T,C or G

<400> 6

tctagagctc	gcggccgcga	gctctaatac	gactcactat	agggcgctga	ctcgatctca	60
gctcactgca	atctctgccc	ccgggggcat	gcgattctcc	tgcctcagcc	ttccaagtag	120
ctgggattac	agggcgtgcaa	caccacaccc	ggctaatttt	gtatttttaa	tagagatggg	180
gttttccctt	gttggccann	atggtctcna	acccctgacc	tcnngtgatc	ccccncccn	240
nganctenna	ctgctgggga	tnnccgnnnn	nnccctcccn	ncncnnnnnn	ncncnnntcn	300
tnntccttnc	tennnnnnnn	cnntcnntcc	nncttctcnc	cnntntttnt	cnncnnccnn	360
cnncnccnt	neccnennnt	tenentcnnn	tnccnnnnnn	ntcnncnnnn	cnnnnctntn	420
ccnntacntc	ntnnnnnnnt	centctntnn	cctcnnnnnt	cnctnccnt	tnctctctcn	480
ntnnnnnnct	ccnnnnntct	cntcncnnnn	tnccctnnnn	ncncncccc	ncctcncnn	540
ctnnntttnn	cnncnnntcc	ntnccnttcn	nnccnnntnn	cnncntcnnc	nnctttnttc	600
ccnccnnttc	cttnccntnn	nnntntcnnn	cnctcnnntc	ntttntctct	nnntcccnnc	660
tcnnttcncc	cnntccncc	ccccnccnt	ctctcncncc	nnnnnnntnn	nnnntccncc	720
tnctnccnttc	ntcnntnccnt	tnctntcnnc	nnnnntnccn	tnccntntnt	ctnnntcnnc	780
tnctntntcn	ccntccnttn	ctntctctct	tnctctctcc	ctcncctnct	cnctcncncc	840
ccnntntntn	tnnccnccnt	ncnnnnccnn	cntcnnnttc	tctctnctnn	nnntnnccct	900
nnccnctncc	ctnnntnccnt	ncnnntacnn	tnctnctcnn	tcttcccttc		950

<210> 7

<211> 1086

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1086)

<223> n = A,T,C or G

<400> 7

tctagagctc	gcggccgcga	gctcaattaa	ccctcactaa	agggagtcga	ctcgatcaga	60
ctgttactgt	gtctatgtag	aaagaagtag	acataagaga	ttccattttg	ttctgtacta	120
agaaaaattc	ttctgccttg	agatgctgtt	aatctgtaac	cctagcccca	accctgtgct	180
cacagagaca	tgtgctgtgt	tgactcaagg	ttcaatggat	ttagggctat	gctttgttaa	240
aaaagtgcct	gaagataata	tgcttggtta	aagtcacac	cattctctaa	tctcaagtac	300
ccaggacac	aatacactgc	ggaaggccgc	aggacctct	gtctaggaaa	gccaggatt	360
gtccaagatt	tctccccatg	tgatagcctg	agatatggcc	tcatgggaag	ggtaagacct	420
gactgtcccc	cagcccgaca	tccccagcc	cgacatcccc	cagcccgaca	cccgaagagg	480
gtctgtgctg	aggaagatta	ntaaaagagg	aaggctcttt	gcattgaagt	aagaagaagg	540
ctctgtctcc	tgctcgctcc	tgggcaataa	aatgtcttgg	tgttaaacct	gaatgtatgt	600
tctacttact	gagaatagga	gaaaacatcc	ttagggctgg	aggtgagaca	ccctggcggc	660
atactgctct	ttaatgcacg	agatgtttgt	ntaattgcca	tccagggcc	ccccctttcc	720
ttaacttttt	atganacaaa	aactttgttc	ntttttctct	cgaacctctc	cccctattan	780
cctattggcc	tgcccatccc	ctccccaaan	ggtgaaaana	tgttcntaaa	tncgagggaa	840
tccaaaacnt	tttcccggtg	gtcccccttc	caaccccgct	cctgggcnnn	tttccctccc	900
aaentgtccc	ggntcccttcn	tcccccccc	cttcccngan	aaaaaacccc	gtnrganggn	960
gccccctcaa	attataacct	tccnaaaca	aannggttcn	aagggtggtt	gnttccgggtg	1020
cgctggtcct	tgagggtccc	cctncacccc	aatttggaan	cnngtttttt	ttattgccc	1080
ntcccc						1086

<210> 8

<211> 1177

<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(1177)
<223> n = A,T,C or G

<400> 8

nccnttttaga	tggtgacaan	ntaaacaagc	ngctcaggca	gctgaaaaaa	gccactgata	60
aagcatcctg	gagtatcaga	gtttactggt	agatcagcct	catttgactt	cccctcccac	120
atggtgttta	aatccagcta	cactacttcc	tgactcaaac	tccactattc	ctgttcatga	180
ctgtcaggaa	ctgttggaag	ctactgaaac	tggtccgacct	gatcttcaaa	atgtgcccct	240
aggaaagctg	gatgccaccg	tggtcacaga	cagtaccncc	ttcctcgaga	agggactacg	300
aggggccggg	gcancgtgta	ccaaggagac	tnatgtgttg	tggtgctcagg	ctttaccanc	360
aaacacctca	ncncnnaagg	ctgaattgat	cgccctcact	caggctctcg	gatggggtaa	420
gggatattaa	cgtaaacact	gacagcagg	acgcctttgc	tactgtgcat	gtacgtggag	480
ccatctacca	ggagcgtggg	ctactcactc	ggcagggtgg	tgtnatccac	tgtaaangga	540
catcaaaagg	aaaacnnggc	tggtgcccgt	ggtaaccana	aanctgaten	ncagctcnaa	600
gatgetgtgt	tgactttcac	tcncncctct	taaacttgct	gcccacantc	tcctttccca	660
accagatctg	cctgacaatc	cccatactca	aaaaaaaaan	aanactggcc	ccgaaccna	720
accaataaaa	acggggangg	tnggtnganc	nncctgaccc	aaaaataatg	gatcccccg	780
gctgcaggaa	ttcaattcan	ccttatcnat	acccccaacn	ngnggggggg	ggcngtncc	840
cattncacct	ntattnattc	tttncccccc	ccccgggcnt	cctttttnaa	ctcgtgaaag	900
ggaaaacctg	ncttaccaan	ttatcncctg	gaacntcccc	ttcnccggtg	gnntanaaaa	960
aaaagccnc	antccntcc	naaatttgca	cngaaaggna	aggaatttaa	cctttatttt	1020
ttnttccttt	antttgtnnn	ccccctttta	cccaggcgaa	cngccatcnt	ttaanaaaaa	1080
aaanagaang	tttatttttc	cttngaacca	tcccaatana	aancaccgc	nggggaacgg	1140
gngngnaggc	cntcaccccc	cttntgtng	gnnggnc			1177

<210> 9
<211> 1146
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(1146)
<223> n = A,T,C or G

<400> 9

nccnnttnt	gatgtgtct	ttttggcctc	tctttggata	ctttccctct	cttcagaggt	60
gaaaagggtc	aaaaggagct	gttgacagtc	atcccagggtg	ggccaatgtg	tccagagtac	120
agactccatc	agtgaggtca	aagcctgggg	cttttcagag	aaggaggat	tatgggtttt	180
ccaattatac	aagtcagaag	tagaaagaag	ggacataaac	caggaagggg	gtggagcact	240
catcacccag	agggacttgt	gcctctctca	gtggtagtag	aggggctact	tcctcccacc	300
acggttgcaa	ccaagaggca	atgggtgatg	agcctacagg	ggacatancc	gaggagacat	360
gggatgacct	taaggagta	ggctgggttt	aaggcggttg	gactgggtga	gggaaactct	420
cctcttcttc	agagagaagc	agtacagggc	gagctgaacc	ggctgaagg	cgaggcgaaa	480
acacggtctg	gctcaggaag	accttggaag	taaaattatg	aatgggtgcat	gaatggagcc	540
atggaagggg	tgctcctgac	caaactcagc	cattgatcaa	tgtagggaa	actgatcagg	600
gaagccggga	atttcattaa	caaccggcca	cacagcttga	acattgtgag	gttcagtgc	660
ccttcaagg	gccactccac	tccaactttg	gccattctac	tttgcnaaat	ttccaaaact	720
tcctttttta	aggccgaatc	cntantccct	naaaaacnaa	aaaaaatctg	cncctattct	780


```

ggaaaaggcc cancccttac caggctggaa gaaattttnc cttttttttt tttttgaagg      840
cntttnttaa attgaacctn aattcncccc cccaaaaaaa aaccncncng gggggcggat      900
ttccaaaaac naattccctt accaaaaaac aaaaaccnc ccttnttccc ttcnccctn      960
ttcttttaat tagggagaga tnaagcccc caatttccng gnetngatnn gtttcccccc 1020
ccccatttt ccnaaacctt tcccanchna ggaanccnc ctttttttng gtcngattna 1080
ncaaccttcc aaaccatttt tccnnaaaaa ntttgntngg ngggaaaaan acctnntttt 1140
atagan                                     1146

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```

<210> 10
<211> 545
<212> DNA
<213> Homo sapien

```

```

<400> 10
cttcattggg tacgggcccc ctcgaggctg acggtatcga taagcttgat atcgaattcc      60
tgcagcccg gggatccact agttctagag tcaggaagaa ccaccaacct tcctgatttt 120
tattggctct gagttctgag gccagttttt ttcttctgtt gattatgcgg gattgtcagg 180
cagatctggc tgtggaaagg agactgtggg cagcaagttt agaggcgtga ctgaaagtca 240
cactgcatct tgagctgctg aatcagcttt ctggttacca cgggcaacag ccgtgttttc 300
cttttgatgt cctttacagt ggattacagc cacctgctga ggtgagtagc ccacgctcct 360
ggtagatggc tccacgtaca tgcacagtag caaaggcgta cctgctgtca gtgttaacgt 420
taatattcct accccatcgg agagcctgag tgaggcgcat caattcagcc cttttgtgct 480
gagtggtttg ctggttaagc cctgaacca caacacatct gctccatgg taacagctgc 540
accg                                           545

```

```

<210> 11
<211> 196
<212> DNA
<213> Homo sapien

```

```

<400> 11
tctcctaggc tgggcacagt ggctcatacc tgtaatcctg accgtttcag aggtcaggt      60
ggggggatcg cttgagccca agatttcaag actagtctgg gtaacatagt gagaccctat 120
ctctacgaaa aaataaaaaa atgagcctgg tgtagtggca cacaccagct gaggagggag 180
aatcgagcct aggaga                             196

```

```

<210> 12
<211> 388
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(388)
<223> n = A,T,C or G

```

```

<400> 12
tctcctaggc ttgggggctc tgactagaaa ttcaaggaa cttgggattca agtccaactg      60
tgacaccaac ttacactgtg gnetccaata aactgcttct ttcctattcc ctctctatta 120
aataaaataa ggaaaacgat gtctgtgtat agccaagtca gntatcctaa aaggagatac 180
taagtacat taaatatcag aatgtaaaac ctgggaacca ggttcccagc ctgggattaa 240
actgacagca agaagactga acagtactac tgtgaaaagc ccgaagnggc aatatgttca 300
ctctaccgtt gaaggatggc tgggagaatg aatgctctgt cccccagtcc caagctcact 360
tactatacct cctttatagc ctaggaga                             388

```

<210> 13
 <211> 337
 <212> DNA
 <213> Homo sapien

<400> 13
 tagtagttgc ctataatcat gtttctcatt attttcacat tttattaacc aatttctggt 60
 taccctgaaa aatatgaggg aaatatatga aacagggagg caatgttcag ataattgatc 120
 acaagatatg atttctacat cagatgctct ttcttttccg gtttatttcc tttttatttc 180
 gggtgtgggg tgaatgtaa tagctttggt tcaagagaga gttttggcag tttctgtagc 240
 ttctgacact gctcatgtct ccaggcatct atttgactt taggaggtgt cgtgggagac 300
 tgagaggtct attttttcca tatttgggca actacta 337

<210> 14
 <211> 571
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(571)
 <223> n = A,T,C or G

<400> 14
 tagtagttgc catacagtgc ctttccattt atttaacccc cacctgaacg gcataaaactg 60
 agtgttcagc tgggtgtttt tactgtaaaac aataaggaga ctttgccttt catttaaacc 120
 aaaatcatat ttcatatttt acgctcgagg gtttttaccg gttccttttt aacttcctta 180
 aaacagtttt taagtcgttt ggaacaagat attttttctt tcctggcagc ttttaacatt 240
 atagcaaatt tgtgtctggg ggactgctgg tcaactgttc tcacagtgc aaatcaaggc 300
 atttgcaacc aagaaaaaaa aatttttttg ttttatttga aactggaccg gataaacggt 360
 gtttgagcgg gctgctgtat atagttttaa atgggtttatt gcacctcctt aagttgcact 420
 tatgtggggg ggggnttttg natagaaagt ntttantcac anagtcacag ggacttttnt 480
 cttttgggna ctgagctaaa aagggtgnt tttcgggtgg gggcagatga aggctcacag 540
 gaggcctttc tottagaggg gggaactnct a 571

<210> 15
 <211> 548
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(548)
 <223> n = A,T,C or G

<400> 15
 tatatatatta ataacttaaa tatattttga tcaccactg ggggtgataag acaatagata 60
 taaaagtatt tccaaaaagc ataaaaccaa agtatcatc caaaccaaat tcatactgct 120
 tccccacccc gcaactgaaac ttacacctct aactgtctac ctaaccaaat tctacccttc 180
 aagtcttttg tgcgtgctca ctactctttt tttttttttt tttnttttgg agatggagtc 240
 tggctgtgca gccagggggt ggagtacaat ggcacaacct cagctcactg naacctccgc 300
 ctcccaggtt catgagattc tcctgnttca gccttccag tagctgggac tacaggtgtg 360
 catcaccatg cctggntaat cttttttngt tttngggtag agatgggggt tttacatggt 420

```

ggccaggntg gntcgaact cctgacctca agtgateccac ccacctcagg ctcccaaagt 480
gctaggatta cagacatgag ccaactgngcc cagnccgtgt gcatgctcac ttctctaggg 540
aactacta 548

```

```

<210> 16
<211> 638
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(638)
<223> n = A,T,C or G

```

```

<400> 16
ttccgttatg cacatgcaga atattctatc ggtacttcag ctattaactca ttttgatggc 60
gcaatccgag cctatcctca agatgagtat tttagaaagaa ttgatttagc gatagaccaa 120
gctggttaagc actctgacta caagaaattg ttcagatgtg atggatttat gacagttgat 180
ctttggaaga gattattaag tgattatatt aaaggggaatc cattaattcc agaatatctt 240
ggtttagctc aagatgatat agaaatagaa cagaaagaga ctacaaatga agatgtatca 300
ccaactgata ttgaagagcc tatagtagaa aatgaattag ctgcatttat tagccttaca 360
catagcgatt ttctgatga atcttatatt cagccatcga catagcatta cctgatgggc 420
aaccttacga ataatagaaa ctgggtgagg ggctattgat gaattcatcc ncagtaaatt 480
tggatatnac aaaatataac tcgattgcat ttggatgatg gaataactaaa tctggcaaaa 540
gtaactttgg agctactagt aacctctctt tttgagatgc aaaattttct tttaggggtt 600
cttattctct actttacgga tattggagca taacggga 638

```

```

<210> 17
<211> 286
<212> DNA
<213> Homo sapien

```

```

<400> 17
actgatggat gtcgccggag ggcagggggc ttatctgatg ctgggctgcc tgttcgtgat 60
gtgcgcggcg attgggctgt ttatctcaaa caccgccacg gcgggtgctga tggcgccctat 120
tgcccttagcg gcggcggaagt caatgggcgt ctcacctat ccttttgcca tgggtggtggc 180
gatggcggct tcggcgcggt ttatgacccc ggtctctctg ccgggttaaca ccctgggtgct 240
tggccctggc aagtactcat tttagcgatt tgtcaaaata ggcgtg 286

```

```

<210> 18
<211> 262
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(262)
<223> n = A,T,C or G

```

```

<400> 18
tcggctcatg cagccccctt ttctcaattt catctgtcac taccctgggt tagtatctca 60
tagccttaca tttttatagc ctctccctg gtctgtcttt tgattttcct gctgtaaatc 120
catatcacac ataactgcaa gtaaacattt ctaaagtgtg gttatgtca tgtcactcct 180
gtgncaagaa atagtttcca ttaccgtctt aataaaattc ggatttggtc ttttctattn 240

```

tcactcttca cctatgaccg aa

262

<210> 19

<211> 261

<212> DNA

<213> Homo sapien

<400> 19

tcggtcatag	caaagccagt	ggtttgagct	ctctactgtg	taaactccta	aaccaaggcc	60
atztatgata	aatggtggca	ggatttttat	tataaacatg	tacccatgca	aatttcctat	120
aactctgaga	tatattcttc	tacattttaa	caataaaaat	aatctatttt	taaaagccta	180
atttgcgtag	ttaggtaaga	gtgtttaatg	agaggggtata	aggtataaat	caccagtcaa	240
cgtttctctg	cctatgaccg	a				261

<210> 20

<211> 294

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(294)

<223> n = A,T,C or G

<400> 20

tacaacgagg	cgacgtcggg	aaaatcggac	atgaagccac	cgctggtctt	ttcgtccgag	60
cgataggcgc	cggccagcca	gcggaacggg	tgcccggatg	gcgaagcgag	ccggagttct	120
tcggactgag	tatgaatctt	gttgtgaaaa	tactcgccgc	cttcgttcga	cgacgtcgcg	180
tcgaaatctt	cgancctcct	acgatcgaag	tcttcgtggg	cgacgatcgc	ggtcagttcc	240
gccccaccga	aatcatgggt	gagccggatg	ctgnccccga	agnccctcgt	tgtn	294

<210> 21

<211> 208

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(208)

<223> n = A,T,C or G

<400> 21

ttggtaaagg	gcatggacgc	agacgcctga	cgtttggtctg	aaaatctttc	attgattcgt	60
atcaatgaat	aggaaaattc	ccaaagaggg	aatgtcctgt	tgctcgccag	ttttntgtt	120
gttctcatgg	anaaggcaan	gagctcttca	gactattggn	attntcgttc	ggctctctgc	180
caactagtcg	ncttgcnang	atcttcat				208

<210> 22

<211> 287

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(287)

<223> n = A,T,C or G

<400> 22

```
nccnttgagc tgagtgattg agatntgtaa tggttgtaag ggtgattcag gcggattagg      60
gtggcggggc acccggcagt gggctctccc acaggccagc aggatttggg gcagggtacgg      120
ngtgcgcata gctcgactat atgctatggc aggcgagccg tggaaaggngg atcaggtcac      180
ggcgctggag ctttccacgg tccatgnatt gngatggctg ttctaggcgg ctgttgccaa      240
gcgtgatggt acgctggctg gagcattgat ttctggtgcc aaggtgg      287
```

<210> 23

<211> 204

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(204)

<223> n = A,T,C or G

<400> 23

```
ttgggtaaag ggagcaagga gaaggcatgg agaggctcan gctggctctg gcctacgact      60
gggccaagct gtcgcggggg atggtggaga actgaagcgg gacctctctg aggtcctccg      120
ncgttacttc nccgtccagg aggagggctct ttccgtggctc tnggaggagc ggggggagaa      180
gatnctcttc atggttonaca tccc      204
```

<210> 24

<211> 264

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(264)

<223> n = A,T,C or G

<400> 24

```
tggattggtc aggagcgggt agagtggcac cattgagggg atattcaaaa atattatatt      60
gtcctaaatg atagtgtctg agtttttctt tgacctatga gttatattgg agtttatatt      120
ttaactttcc aatcgcatgg acatgttaga cttattttct gttaatgatt nctatatttta      180
ttaaattgga tttgagaaat tggttnttat tatatcaatt tttggtatatt gttgagtttg      240
acattatagc ttagtatgtg acca      264
```

<210> 25

<211> 376

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(376)

<223> n = A,T,C or G

<400> 25

```
<210> 26
<211> 372
<212> DNA
<213> Homo sapien
```

<400>	26								
acgag	gggaaactcc	gtctctacaa	aaattaaaaa	attagccagg	tgtggtggtg				60
ctgta	atcccagcta	cttggggcggc	tgagacacaa	gaaccaccta	aatgtggggag				120
aggtt	gcattgagtca	tgatcgcgcc	actgcactcc	agcctgggtg	acagactgag				180
gcctc	aaaagaaaaa	gaataggaag	ttcagaaacc	ctgggtgtgg	ngcccagcaa				240
attta	aacaatccct	gcaggcaatg	ctgatgcagc	ctaagttcaa	gagctgctgt				300
aggca	gnagtaaggg	cttccatcca	gcatacaggc	caacactgca	aaagcacctg				360
gttgg	ta								372

<400>	27									
tccac	atctacaagt	tttattttatt	ttgtggggtt	tcagggtgac	taagtttttc					60
attga	aaagagaagt	tgctaaaagg	tgcacaggaa	atcatttttt	taagtgaata					120
atatg	ggtcogtgct	taatacaact	gagacatatt	tgttctctgt	tttttagag					180
tctta	aagccaatc	ccacaatggt	gaaaaaaaaa	tagaaagtat	ttgttctacc					240
ggaga	ctgcagggat	tctcettgaa	aacggagtat	ggaatcaatc	ttaaataaat					300
attgg	ttggtcttct	gggataagaa	attcccaact	cagtggtctg	aaattcacct					360
ttttt	gggaaaaaat	agtcgaaaat	gtcaatttgg	tccataaaat	acatgttact					420
aaqat	attttaagac	aaattctttc	agagctctaa	gattggtgtg	gacagua					477

```
<220>
<221> misc_feature
<222> (1)...(438)
<223> n = A,T,C or G
```

<400> 28
tctncaacct cttagantgtc aaaaaccttn taggcctatct ctaaaagctg actggtattc 60

```
<210> 29
<211> 620
<212> DNA
<213> Homo sapien
```

<400> 29						
ggtac	cagccccaag	ccttgacaac	ttccataggg	tgtcaagcct	gtgggtgcac	60
tcaaa	aattgagttt	tgggatcctc	agcctagatt	tcagaggata	taaagaaaca	120
cacct	agatattcag	acaaaagttt	actacaggga	tgaagctttc	acggaaaacc	180
tagga	aagtacagaa	gagaaatgtg	ggtttgagc	cccaaacag	aatccccctc	240
actgc	ctaataaaac	tgtgagaaga	tggccactgt	catccagaca	ccagaatgat	300
cacca	aaaacttatg	ccatattgcc	tataaaacct	acagacactc	aatgccagcc	360
aaaaa	aaaactgaga	agaagactgt	nccctacaat	gccaccggag	cagaactgcc	420
ccatg	gaagcacagc	tcttatatca	atgtgacctg	gatgttgaga	catggaatcc	480
aatcn	ttttaanact	tccacggttt	aatgactgcc	ctattanatt	cngaacttan	540
ggcct	gtgacctctt	tgctttggcc	attccccctt	tttggaatgg	ctnttttttt	600
gcctg	tnccctctta					620

```
<210> 30
<211> 100
<212> DNA
<213> Homo sapien
```

```

<400> 30
ttacaacgag ggggtcaatg tcataaatgt cacaataaaa caatctcttc tttttttttt      60
tttttttttt tttttttttt tttttttttt tttttttttt                                100

```

```
<210> 31
<211> 762
<212> DNA
<213> Homo sapien
```

```
<220>
<221> misc_feature
<222> (1)...(762)
<223> n = A,T,C or G
```

```

<400> 31
tagtctatgc gccggacaga gcagaattaa attggaagtt gccctccgga cttttctaccc      60
acactcttcc tgaaaagaga aagaaaagag gcaggaaaga ggttaggatt tcattttcaa      120
gagtcagcta attaggagag cagagtttag acagcagtag gcaccccatg atacaaacca      180

```

tgacaaagt	cctgttttag	taactgccag	acatgatcct	gctcaggttt	tgaaatctct	240
ctgcccataa	aagatggaga	gcaggagtgc	catccacatc	aacacgtgtc	caagaaagag	300
tctcagggag	acaaggggat	caaaaaacaa	gattcttaat	gggaaggaaa	tcaaaccaaa	360
aaattagatt	tttctctaca	tatatataat	atacagatat	ttaacacatt	attccagagg	420
tggctccagt	ccttggggct	tgagagatgg	tgaaaacttt	tgttccacat	taacttctgc	480
tctcaaattc	tgaagtatat	cagaatggga	caggcaatgt	tttgctccac	actggggcac	540
agacccaaat	ggttctgtgc	ccgaagaaga	gaagcccgaa	agacatgaag	gatgcttaag	600
gggggttggg	aaagccaaat	tggtantatc	ttttcctcct	gcctgtgttc	cngaagtctc	660
cncatgaagga	attcttaaaa	ccctttgtga	ggaaatgcc	ccttaccatg	acaantggtc	720
ccattgcttt	tagggngatg	gaaacaccaa	gggttttgat	cc		762

<210> 32

<211> 276

<212> DNA

<213> Homo sapien

<400> 32

tagtctatgc	gtgtattaac	ctccccctcc	tcagtaacaa	ccaaagaggc	aggagctggt	60
attaccaacc	ccattttaca	gatgcatcaa	taatgacaga	gaagtgaagt	gacttgcgca	120
cacaaccagt	aaattggcag	agtcagatgt	gaatccatgg	agtctggtct	gcactttcaa	180
tcaccgaata	ccctttctaa	gaaacgtgtg	ctgaatgagt	gcatggataa	atcagtgtct	240
actcaacatc	tttgccctaga	tatcccgcac	agacta			276

<210> 33

<211> 477

<212> DNA

<213> Homo sapien

<400> 33

tagtagttgc	caaatatttg	aaaatttacc	cagaagtgat	tgaaaacttt	ttggaaacaa	60
aaacaaataa	agccaaaagg	taaaataaaa	atatctttgc	actctcgtaa	ttacctatcc	120
ataacttttt	caccgtaagc	tctcctgctt	gttagtgtag	tgtggttata	ttaaactttt	180
tagttattat	tttttattca	cttttccact	agaaagtcat	tattgattta	gcacacatgt	240
tgatctcatt	tcattttttc	tttttatagg	caaaatttga	tgctatgcaa	caaaaatact	300
caagcccatt	atcttttttc	cccccgaaat	ctgaaaattg	caggggacag	aggggaagtta	360
tcccattaaa	aaattgtaaa	tatgttcagt	ttatgtttta	aaatgcacaa	aacataagaa	420
aattgtgttt	acttgagctg	ctgattgtaa	gcagttttat	ctcaggggca	actacta	477

<210> 34

<211> 631

<212> DNA

<213> Homo sapien

<400> 34

tagtagttgc	caattcagat	gatcagaaat	gctgctttcc	tcagcattgt	cttgttaaac	60
cgcattgccat	ttggaacttt	ggcagtgaga	agccaaaagg	aagaggtgaa	tgacatatat	120
atatatatat	attcaatgaa	agtaaaatgt	atatgctcat	atactttcta	gttatcagaa	180
tgagttaaag	tttatgccat	tgggctgctg	catattttta	tcagaagata	aaagaaaatc	240
tgggcatttt	tagaatgtga	tacatgtttt	tttaaaactg	ttaaatatta	tttcgatatt	300
tgtctaagaa	ccggaatggt	cttaaaattt	actaaaacag	tattgtttga	ggaagagaaa	360
actgtactgt	ttgccattat	tacagtcgta	caagtgcatt	tcaagtcacc	cactctctca	420
ggcatcagta	tccacctcat	agctttacac	atthtgacgg	ggaatattgc	agcatcctca	480
ggcctgacat	ctgggaaagg	ctcagatcca	cctactgctc	cttgctcggt	gatttgtttt	540
aaaatattgt	gcctgggtgtc	actttttaagc	cacagccctg	cctaaaagcc	agcagagAAC	600

agaacccgca ccattctata ggcaactact a

631

<210> 35
 <211> 578
 <212> DNA
 <213> Homo sapien

<400> 35
 tagtagttgc catcccatat tacagaaggc tctgtataca tgaattattht ggaagtgatc 60
 tgtttttctct ccaaaacccat ttatcgtaat ttcaccagtc ttggatcaat cttgggtttcc 120
 actgatacca tgaaacctac ttggagcaga cattgcacag ttttctgtgg taaaaactaa 180
 aggtttattht gctaagctgt catcttatgc ttagtattht ttttttacag tggggaattg 240
 ctgagattac atthttgttat tcattagata ctttgggata acttgacact gtcttcttht 300
 tttcgcttht aattgtctac atcatgcttht tgaacaaga acacattagt cctcaagtat 360
 tacataagct tgcttggttac gcttggtggt ttaaaggact atctttggcc tcagggtcac 420
 aagaatgggc aaagtgttht cttatgttct gtagttctca ataaaagatt gccaggggccc 480
 gggtagctgt gctgcactg taatcccagc actttgggaa gctgaggctg gcggatcatg 540
 ttagggcagg tggtcgaaac cagcctgggc aactacta 578

<210> 36
 <211> 583
 <212> DNA
 <213> Homo sapien

<400> 36
 tagtagttgc ctgtaatccc agcaactcag gaggctgggg caggagaatc agttgaacct 60
 gggaggcaga agttgtaatt agcaaagatc gcaccattgc acttcagcct gggcaacaag 120
 agtgagattc catctcaaaa acaaaaaaaaaa gaaaaagaaa agaaaaggaa aaaacgtata 180
 aaccagcca aaacaaaatg atcattcttht taataagcaa gactaattta atgtgttht 240
 ttaatcaaag cagttgaatc ttctgagtta ttggtgaaaa taccatgta gtttaatttag 300
 ggttcttact tgggtgaacg tttgatgttc acaggttata aaatggttaa caaggaaaat 360
 gatgcataaa gaatcttata aactactaaa aataaataaa atataaatgg atagggtgcta 420
 tggatggagt ttttgtgtaa tttaaaatct tgaagtcatt ttggatgctc attggttgctc 480
 tggtaattht cattaggaaa aggttatgat atggggaaac tgtttctgga aattgcggaa 540
 tgtttctcat ctgtaaaatg ctagtatctc agggcaacta cta 583

<210> 37
 <211> 716
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(716)
 <223> n = A,T,C or G

<400> 37
 gatctactag toatntggat tctatocatt gcagctaagc ctttctgaat ggattctact 60
 gctttcttgt tctttaatcc agacccttat atatgttht gttcacaggc agggcaatgt 120
 ttagtgaaaa caattctaaa ttttttattht tgcattthtca tgctaatttc cgtcacactc 180
 cagcaggctt cctgggagaa taaggagaaa tacagctaaa gacattgtcc ctgcttactt 240
 acagcctaatt ggtatgcaaa accacttcaa taaagtaaca ggaaaagtac taaccaggta 300
 gaatggacca aaactgatat agaaaaatca gaggaagaga ggaacaaata tttactgagt 360
 cctagaatgt acaaggcttht ttaattacat atthttatgta aggcctgcaa aaaacagggtg 420

```

agtaatcaac atttgtccca ttttacatat aaggaaactg aagcttaaata tgaataatatt 480
aatgcataga ttttatagtt agaccatggt cagggtcccta tgttatactt actagctgta 540
tgaatatgag aaaataatatt tgttatattt ttggcatcag tatttttcac tgcaaaataa 600
agctaaagtt atttagcaaa cagtcagcat agtgcctgat acatagtagg tgctccaac 660
atgattacnc tantattngg tattanaaaa atccaatata ggcntggata aaaccg 716

```

```

<210> 38
<211> 688
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(688)
<223> n = A,T,C or G

```

```

<400> 38
ttctgtccac atatcatccc actttaattg ttaatcagca aaactttcaa tgaaaaatca 60
tccattttta ccaggatcac accaggaaac tgaagggtgta ttttttttta ccttaaaaaa 120
aaaaaaaaa accaaacaaa ccaaaacaga ttaacagcaa agagttctaa aaaatttaca 180
tttctcttac aactgtcatt cagagaacaa tagttcttaa gtctgttaaa tcttggcatt 240
aacagagaaa cttgatgaan agttgtactt ggaatattgt ggattttttt ttttgtctaa 300
tctcccccta ttgttttgcc aacagtaatt taagtttgtg tggaacatcc ccgtagttga 360
agtgtaaaca atgtatagga aggaatatat gataagatga tgcatacat atgcattaca 420
tgtagggacc ttcacaactt catgcactca gaaaacatgc ttgaagagga ggagaggacg 480
gccaggggc accatccagg tgccttgagg acagagaatg cagaagtggc actgttgaaa 540
tttagaagac catgtgtgaa tggtttcagg cctgggatgt ttgccaccaa gaagtgcctc 600
cgagaaattt ctttcccatt tggaatacag ggtggcttga tgggtacggt ggggtgacca 660
acgaagaaaa tgaaattctg ccctttcc 688

```

```

<210> 39
<211> 585
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(585)
<223> n = A,T,C or G

```

```

<400> 39
tagtagttgc cgcnnaccta aaanttggaa agcatgatgt ctaggaaaca tantaaaata 60
gggtatgcct atgtgctaca gagagatggt agcattttaa gtgcatantt ttatgtattt 120
tgacaaatgc atatncctct ataatccaca actgattacg aagctattac aattaaaaag 180
tttggccggg cgtgggtggg ggtggctgac gcctgtaatc ccagcacttt gggaggccga 240
ggcacgcgga tcacgagggtc gggagttcaa gaccatcctg gctaacacgg tgaaagtcca 300
tctctactaa aaatacgaaa aaattacccc ggcgtgggtg cgggcgcctg tagtcccagc 360
tactccggag gctgaggcag gagaatggcg tgaaccagg acacggagct tgcagtgtgc 420
caacatcacg tcaactgcct ccagcctggg ggacaggaac aagantcccg tctctanaaa 480
agaaaaatac tactnatant ttcnacttta ttttaantta cacagaactn cctcttggtg 540
cccccttacc attcatctca cccacctcct atagggcacn nctaa 585

```

```

<210> 40
<211> 475

```

<212> DNA

<213> Homo sapien

<400> 40

tctgtccaca	ccaatcttag	aagctctgaa	aagaatttgt	ctttaaata	cttttaata	60
taacatgtat	tttatggacc	aaattgacat	tttcgactgt	tttttccaaa	aaagtcaggt	120
gaatttcagc	acactgagtt	gggaatttct	tatcccagaa	gaccaaccaa	tttcataatt	180
atttaagatt	gattccatac	tccgttttca	aggagaatcc	ctgcagtctc	cttaaaggta	240
gaacaaatac	ttcctatatt	tttttcacca	ttgtgggatt	ggactttaag	aggtgactct	300
aaaaaacag	agaacaaata	tgtctcagtt	gtattaagca	cggacccata	ttatcatatt	360
cacttaaaaa	aatgatttcc	tgtgcacctt	ttggcaactt	ctcttttcaa	tgtagggaaa	420
aacttagtca	ccctgaaaac	ccacaaaata	aataaaactt	gtagatgtgg	acaga	475

<210> 41

<211> 423

<212> DNA

<213> Homo sapien

<400> 41

taagagggta	catcggttaa	gaacgtaggc	acatctagag	cttagagaag	tctggggtag	60
gaaaaaaatc	taagtattta	taagggtata	ggtaacattt	aaaagtaggg	ctagctgaca	120
ttatttagaa	agaacacata	cggagagata	agggcaaagg	actaagacca	gaggaacact	180
aatatttagt	gatcacttcc	attcttggtt	aaaatagtaa	cttttaagtt	agcttcaagg	240
aagatttttg	gccatgatta	gttgtcaaaa	gttagttctc	ttgggtttat	attactaatt	300
ttgttttaag	atccttggtt	gtgctttaat	aaagtcatgt	tatatcaaac	gctctaaaac	360
attgtagcat	gttaaatgtc	acaatatact	taccatttgt	tgtatatggc	tgtaccctct	420
cta						423

<210> 42

<211> 527

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(527)

<223> n = A,T,C or G

<400> 42

tctcctaggc	taatgtgtgt	gtttctgtta	aagtaaaaag	ttaaaaattt	taaaaataga	60
aaaaagctta	tagaataaga	atatgaagaa	agaaaatatt	tttgtacatt	tgacacatga	120
gtttatgttt	taagctaagt	gttattacaa	aagagccaaa	aaggttttta	aaattaaaaac	180
gtttgtaaag	ttacagtacc	cttatgttaa	tttataattg	aagaaagaaa	aacttttttt	240
tataaatgta	gtgtagccta	agcatacagt	atttataaag	tctggcagtg	ttcaataatg	300
tcttaggcct	tcacattcac	tactgactc	acccagagca	acttccagtc	ctgtaagctc	360
cattcgtggg	aagtgcctta	tacaggtgca	ccatttattt	tacagtattt	ttactgtacc	420
ttctctatgt	ttccatatgt	ttcgatatac	aaataccact	ggttactatn	gcccnacagg	480
taattccagt	aacacggcct	gtatacgtct	ggtancccta	ngnaaga		527

<210> 43

<211> 331

<212> DNA

<213> Homo sapien

<400> 43

tottcaacct	cgtaggacaa	ctctcatatg	cctgggcact	atTTTTtaggt	tactaccttg	60
gctgcccttc	tttaagaaaa	aaaaaagaag	aaaaaagaac	ttttccacaa	gtttctcttc	120
ctctagttgg	aaaattagag	aatcatgtt	tttaattttg	tgttatttca	gatcacaat	180
tcaaacactt	gtaaacatta	agcttctgtt	caatccctg	ggaagaggat	tcattctgat	240
atttacgggt	caaaagaagt	tgtaatatg	tgcttgaac	acagagaacc	agttattaac	300
ttcctactac	tattatataa	taaataataa	c			331

<210> 44

<211> 592

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(592)

<223> n = A,T,C or G

<400> 44

ggcttagtag	ttgccaggca	aaatarcgtt	gattctcttc	aggagccacc	cccaacaccc	60
ctgtttgctt	ctagacctat	acctagacta	aagtcccagc	agacccttag	agggtgagggt	120
cagagtgacc	cttgaggaga	tgtgctacac	tagaaaagaa	ctgcttgagt	tttctaattt	180
atataagcag	aaatctggag	aagagtcata	ggaatggata	ttaaggggtg	gagataatgg	240
cgaaggaat	atagagttgg	atcaggctgg	acttattgat	ttgaacccac	taagtagaga	300
ttctgctttt	gatgttgcag	ctcaggagg	taaaaaagg	tttaatgggt	ctaatagttt	360
atttgcttgg	ttagctgaaa	tatggataaa	agatggccca	ctgtgagcaa	gctggaaatg	420
cctgatctct	ctcagtttaa	tgtagaggaa	gggatccaaa	agtttaggga	ganttggtatg	480
ctggraktgg	attgggtcact	ttgrgacct	ccwtccag	ctgggagggt	ccagaagata	540
cacccttgac	caacgctttg	cgaaatggat	ttgtgatggc	ggcaactact	aa	592

<210> 45

<211> 567

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(567)

<223> n = A,T,C or G

<400> 45

ggcttagtag	ttgccattgc	gagtgttgc	tcaacgagcg	ttgaacatgg	cggattgtot	60
agattcaacg	gatttgagtt	ttaccagcaa	agcgaaccaa	gcgcggccca	gagaattatg	120
ggttggttgg	ctttgaaaag	atggaaatcc	tgtaggccta	gtcagaaaag	ccttcttgca	180
gaacagttgg	ttctcgggcg	aacgctcatc	aagatgccca	ttggaaaggc	tagcgtgtat	240
ttgggagagc	ctgatagcgt	gtcttctgat	gatgtttgtg	cttggacagt	gacaaaagat	300
atgcaaagca	agtcogaact	agacgtcaag	cttcgtgagc	aaattattgt	agactcctac	360
ttatactgtg	aggaatgata	gccaagggtg	gggacttta	gactaagggtg	gtttgtactt	420
gcgcgatga	tcccaggcag	aaagamctga	tcgctagttt	tatacgggca	actactaagc	480
cgaattccag	cacactggcg	gccgttacta	attggatccg	anctcggtag	cagcttgatg	540
catascctga	gttwtctata	ntgtcnc				567

<210> 46

<211> 908

1000
 900
 800
 700
 600
 500
 400
 300
 200
 100
 0

<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(908)
<223> n = A,T,C or G

<400> 46

gagcgaaaga	ccgagggcag	ngnntangng	cgangaagcg	gagagggcca	aaaagcaacc	60
gctttccccg	gggggtgccg	attcattaag	gcaggtggag	gacaggtttc	ccgatggaag	120
gcggcagggg	cgcaagcaat	taatgtgagt	aggccattca	ttagcaccog	ggcttaacat	180
ttaagcttcg	ggttggtatg	tggtgggaat	tgtgagcggg	taacaatttc	acacaggaaa	240
cagctatgac	catgattacg	ccaagctatt	taggtgacat	tatagaataa	ctcaagttat	300
gcatacaagt	tggtaccgag	ttcggatcca	ctagtaacgg	ccgccagtgt	gtggaattcg	360
gcttagtagt	tgccgaccat	ggagtgtctac	ctaggctaga	atacctgagy	tcctccctag	420
cctcactcac	attaaattgt	atcttttcta	cattagatgt	cctcagcgcc	ttattttctgc	480
tggacwatcg	ataaattaat	cctgatagga	tgatagcagc	agattaatta	ctgagagtat	540
gttaatgtgt	catccctcct	atataacgta	tttgcathtt	aatggagcaa	ttctggagat	600
aatccctgaa	ggcaaaggaa	tgaatcttga	gggtgagaaa	gccagaatca	gtgtccagct	660
gcagttgtgg	gagaaggtga	tattatgtat	gtctcagaag	tgacaccata	tgggcaacta	720
ctaagcccga	attccagcac	actggcgggc	gttactaatg	gatccgagct	cggtaccaag	780
cttgatgcat	agcttgagta	tctatagtgt	cactaaatag	cctggcggtta	tcattggtcat	840
agctgtttcc	tgtgtgaaat	tggtatccgc	tcccaattcc	ccccaccata	cgagccggaa	900
cataaagt						908

<210> 47
<211> 480
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(480)
<223> n = A,T,C or G

<400> 47

tgccaacaag	gaaagtttta	aatttcccct	tgaggattct	tggtgatcat	caaattcagt	60
ggtttttaag	gttgttttct	gtcaaataac	tctaacttta	agccaaacag	tatatggaag	120
cacagataka	atattacaca	gataaaagag	gagttgatct	aaagtaraga	tagttggggg	180
ctttaatttc	tggaacctag	gtctcccat	cttcttctgt	gctgaggaac	ttcttggaag	240
cggggattct	aaagttcttt	ggaagacagt	ttgaaaacca	ccatgttgtt	ctcagtacct	300
ttatttttaa	aaagtaggtg	aacatthttga	gagagaaaag	ggcttggttg	agatgaagtc	360
ccccccccc	cttttttttt	ttttagctga	aatagatacc	ctatgttnaa	rgaarggatt	420
attatttacc	atgccaytar	scacatgctc	tttgatgggc	nyctccstac	cctccttaag	480

<210> 48
<211> 591
<212> DNA
<213> Homo sapien

<400> 48

aagagggtac	cgagtggaaat	ttccgcttca	ctagtctggg	gtggctagtc	ggtttcgtgg	60
tggccaacat	tacgaacttc	caactcaacc	gttcttggtg	gttcaagcgg	gagtaccggc	120

gaggatggtg	gcgtgaattc	tggcctttct	ttgccgtggg	atcggtagcc	gccatcatcg	180
gtatgtttat	caagatcttc	tttactaacc	cgacctctcc	gatttacctg	cccagagccgt	240
ggtttaacga	ggggaggggg	atccagtcac	gcgagtactg	gtcccagatc	ttcgccatcg	300
tcgtgacaat	gcctatcaac	ttcgtcgtca	ataagttgtg	gaccttccga	acggtgaagc	360
actccgaaaa	cgtccgggtg	ctgctgtgcg	gtgactccca	aaatcttgat	aacaacaagg	420
taaccgaatc	gcgctaagga	accccgcat	ctcgggtact	ctgcatatgc	gtaccacctta	480
agccgaattc	cagcacactg	gcggccgtta	ctaattggat	ccgaactccg	taaccaagcc	540
tgatgcgtaa	cttgagttat	tctatagtgt	ccctaaaata	acctggcggt	a	591

<210> 49

<211> 454

<212> DNA

<213> Homo sapien

<400> 49

aagagggtag	ctgccttgaa	atttaaattgt	ctaaggaaar	tgggagatga	ttaagagttg	60
gtgtggcyta	gtcacaccaa	aatgtattta	ttacatcctg	ctcctttcta	gttgacagga	120
aagaaaagctg	ctgtggggaa	aggagggata	aatactgaag	ggatttacta	aacaaatgtc	180
catcacagag	ttttcctttt	tttttttttg	agacagagtc	ttgctctgtc	accagggctg	240
gaatgaagwg	gtatgatctc	agttgaatgc	aacctctacc	tcctaggttc	aagcgattct	300
catgcctcag	cctcctgagc	agctgggact	ataggcgcat	gctaccatgc	caggctaatt	360
tttatatttt	tattagagac	ggggtgttgc	catgttggcc	aggcaggctc	cgaactcctg	420
ggcctcagat	gatctgcccc	accgtaccct	ctta			454

<210> 50

<211> 463

<212> DNA

<213> Homo sapien

<400> 50

aagagggtag	caaaaaaaag	aaaaaggaaa	aaaagaaaaa	caacttgtag	aaggctttct	60
gctgcataca	gctttttttt	tttaaataaa	tggtgccaac	aaatgttttt	gcattcacac	120
caattgctgg	ttttgaaatc	gtactcttca	aaggtatttg	tgcagatcaa	tccaatagtg	180
atgccccgta	ggttttgtgg	actgcccacg	ttgtctacct	tctcatgtag	gagccattga	240
gagactgttt	ggacatgcct	gtgttcatgt	agccgtgatg	tccggggggc	gtgtacatca	300
tgttaccgtg	gggtgggggc	tgcattggct	gctgggcata	tggctgggtg	cccatcatgc	360
ccatctgcat	ctgcataggg	tattggggcg	tttgatccat	atagccatga	ttgctgtggt	420
agccactggt	catcattggc	tgggacatgc	tgttaccctc	tta		463

<210> 51

<211> 399

<212> DNA

<213> Homo sapien

<400> 51

cttcaacctc	ccaaagtgtc	gggattacag	gactgagcca	ccacgctcag	cctaagcctc	60
tttttcacta	ccctctaagc	gatctaccac	agtgatgagg	ggctaaagag	cagtgcaatt	120
tgattacaat	aatggaactt	agatttatta	attaacaatt	tttccttagc	atgttggttc	180
cataattatt	aagagtatgg	acttacttag	aaatgagctt	tcatttttaag	aatttcatct	240
ttgaccttct	ctattagtct	gagcagtatg	acactatacg	tatttttatt	aactaaccta	300
ccttgagcta	ttacttttta	aaaggctata	tacatgaatg	tgtattgtca	actgtaaagc	360
cccacagtat	ttaattatat	catgatgtct	ttgagggttg			399

<210> 52

<211> 392
 <212> DNA
 <213> Homo sapien

<400> 52
 cttcaacctc aatcaacctt ggtaattgat aaaatcatca cttaactttc tgatataatg 60
 gcaataatta tctgagaaaa aaaagtgggtg aaagattaaa cttgcatttc tctcagaatc 120
 ttgaaggata tttgaataat tcaaaagcgg aatcagtagt atcagccgaa gaaactcact 180
 tagctagaac gttggaccca tggatctaag tccctgccct tccactaacc agctgattgg 240
 ttttgtgtaa acctcctaca cgcttgggct tggtcgcctc atttgtcaaa gtaaaggctg 300
 aaataggaag ataatgaacc gtgtcttttt ggtctctttt ccatccatta ctctgatttt 360
 acaaagaggc ctgtattccc ctggtgaggt tg 392

<210> 53
 <211> 179
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(179)
 <223> n = A,T,C or G

<400> 53
 ttccgggtgat gcctcctcag gctacagtga agactggatt acagaaaggt gccagcgaga 60
 tttcagattc ctgtaaacct cttaaagaaaa ggagtcgcgc ctcaactgat gtagaaatga 120
 ctagttcagc atacngagac acntctgact ccgattctag aggactgagt gacctgcan 179

<210> 54
 <211> 112
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(112)
 <223> n = A,T,C or G

<400> 54
 ttccgggtgat gcctcctcag gctacatcat natagaagca aagtagaana atcnngtttg 60
 tgcattttcc cacanacaaa attcaaatga ntggaagaaa ttggganagt at 112

<210> 55
 <211> 225
 <212> DNA
 <213> Homo sapien

<400> 55
 tgagcttccg cttctgacaa ctcaatagat aatcaaagga caactttaac agggattcac 60
 aaaggagtat atccaaatgc caataaacat ataaaaagga attcagcttc atcatcatca 120
 gaagwatgca aattaaaacc ataatgagaa accactatgt cccactagaa tagataaaat 180
 cttaaaagac tggtaaaacc aagtgttggg aaggcaagag gagca 225

<210> 56

<211> 175
 <212> DNA
 <213> Homo sapien

<400> 56
 gctcctcttg ccttaccaac acattctcaa aaacctgtta gagtcctaag cattctcctg 60
 ttagtattgg gattttaccc ctgtcctata aagatgttat gtaccaaaaa tgaagtggag 120
 ggccataccc tgagggaggg gagggatctc tagtgttgtc agaagcggaa gctca 175

<210> 57
 <211> 223
 <212> DNA
 <213> Homo sapien

<400> 57
 agccatttac caccatgga tgaatggatt ttgtaattct agctgttgta ttttgtgaat 60
 ttgttaattt tggtgttttt ctgtgaaaca catacattgg atatgggagg taaaggagtg 120
 tccagttgc tctgtgtcac tccctttata gccattactg tcttgtttct tgtaactcag 180
 gttaggtttt ggtctctctt gctccactgc aaaaaaaaaa aaa 223

<210> 58
 <211> 211
 <212> DNA
 <213> Homo sapien

<400> 58
 gttcgaaggt gaacgtgtag gtagcggatc tcacaactgg ggaactgtca aagacgaatt 60
 aactgacttg gatcaatcaa atgtgactga ggaaacacct gaaggtgaag aacatcatcc 120
 agtggcagac actgaaaata aggagaatga agttgaagag gtaaaagagg aggggtccaaa 180
 agagatgact ttggatgggt ggtaaatggc t 211

<210> 59
 <211> 208
 <212> DNA
 <213> Homo sapien

<400> 59
 gctcctcttg ccttaccaac tttgcacca tcatcaacca tgtggccagg tttgcagccc 60
 aggtgcaca tcaggggact gcctcgcaat acttcatgct gttgctgctg actgatggtg 120
 ctgtgacgga tgtggaagcc acacgtgagg ctgtgggtgcg tgcctcgaac ctgccccatgt 180
 cagtgatcat tatgggtggt aaatggct 208

<210> 60
 <211> 171
 <212> DNA
 <213> Homo sapien

<400> 60
 agccatttac caccatact aaattctagt tcaaactcca acttcttcca taaaacatct 60
 aaccactgac accagttggc aatagcttct tccttcttta acctcttaga gtatttatgg 120
 tcaatgccac acatttctgc aactgaataa agttggtgaag gcaagaggag c 171

<210> 61
 <211> 134

<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)..(134)
<223> n = A,T,C or G

<400> 61
cgggtgatgc ctccctcaggc tttggtgtgt ccactcnact cactggcctc ttctccagca 60
actggtgaan atgtcctcan gaaaancncc acacgcngct caggggtggg tggaancat 120
canaatcatc nggc 134

<210> 62
<211> 145
<212> DNA
<213> Homo sapien

<400> 62
agaggggtaca tatgcaacag tatataaagg aagaagtgca ctgagaggaa cttcatcaag 60
gccatttaat caataagtga tagagtcaag gctcaaccca ggtgtgacgg attccaggtc 120
ccaagctcct tactggtacc ctctt 145

<210> 63
<211> 297
<212> DNA
<213> Homo sapien

<400> 63
tgcactgaga ggaattcaaa gggtttatgc caaagaacaa accagtcctc tgcagcctaa 60
ctcatttggt tttgggctgc gaagccatgt agagggcgat caggcagtag atggccctc 120
ccacagtcag cgccatggtg gtccggtaaa gcatttggtc aggcaggcct cgtttcagggt 180
agacggggcac acatcagctt tctggaaaaa cttttgtagc tctggagctt tgtttttccc 240
agcataatca tacactgtgg aatcggaggt cagtttagtt ggtaaggcaa gaggagc 297

<210> 64
<211> 300
<212> DNA
<213> Homo sapien

<400> 64
gcactgagag gaacttccaa tactatgttg aataggagtg gtgagagagg gcateccttgt 60
cttgtgccgg ttttcaaagg gaatgcttcc agcttttgcc cattcagtat aatattaaag 120
aatgttttac ctttttctgt cttgctgttt tttctgtgtt tttgttggtc tcttcattct 180
ccatttttag gcctttacat gttaggaata tatttctttt aatgatactt cacctttggt 240
atcttttgtg agactctact catagtgtga taagcactgg gttggtgaagg caagaggagc 300

<210> 65
<211> 203
<212> DNA
<213> Homo sapien

<400> 65
gctcctcttg ccttaccaac tcacccagta tgtcagcaat tttatcrgct ttacctacga 60

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<210> 66
<211> 344
<212> DNA
<213> Homo sapien
```

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<210> 67
<211> 157
<212> DNA
<213> Homo sapien
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```
<210> 68
<211> 137
<212> DNA
<213> Homo sapien
```

```
<210> 69
<211> 137
<212> DNA
<213> Homo sapien
```

```
<210> 70
<211> 220
<212> DNA
<213> Homo sapien
```

```
<220>
<221> misc feature
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<222> (1)...(220)

<223> n = A,T,C or G

<400> 70

agcatgttga	gcccagacac	gcaatctgaa	tgagtgtgca	cctcaagtaa	atgtctacac	60
gctgcctggg	ctgacatggc	acaccatcnc	gtggagggca	casctctgct	cngcctacwa	120
cgagggcant	ctcatwgaca	ggttccaccc	accaaactgc	aagaggetca	nnaagtactr	180
ccaggggmya	sggacmasgg	tgggaytyca	ycacwcatct			220

<210> 71

<211> 353

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(353)

<223> n = A,T,C or G

<400> 71

cgttagggtc	tctatccact	gctaaacat	acacctgggt	aaacagggac	catttaacat	60
tcccanctaa	atatgccaa	tgacttcaca	tgtttatctt	aaagatgtcc	aaaacgcaac	120
tgattttctc	ccctaaacct	gtgatgggtg	gatgattaan	cctgagtggg	ctacagcaag	180
ttaagtgcaa	gggtgctaaat	gaangtgacc	tgagatacag	catctacaag	gcagtacctc	240
tcaacncagg	gcaactttgc	ttctcanagg	gcatttagca	gtgtctgaag	taattttctgt	300
attacaactc	acggggcggg	gggtgaatat	ctantggana	gnagacccta	acg	353

<210> 72

<211> 343

<212> DNA

<213> Homo sapien

<400> 72

gcactgagag	gaacttccaa	tacyatkatc	agagtgaaca	rgcarccyac	agaacaggag	60
aaaatgttyg	caatctctcc	atctgacaaa	aggctaatat	ccagawtcta	awaggaactt	120
aaacaaatth	atgagaaaag	aacaracaac	ctcawcaaaa	agtgggtgaa	ggawatgcts	180
aaargaagac	atytattcag	ccagtaaaca	yatgaaaaaa	aggctcatsa	tcactgawca	240
ttagagaaat	gcaaatcaaa	accacaatga	gataccatct	yayrccagtt	agaaygggtga	300
tcattaaaaa	stcaggaaac	aacagatgct	ggacaagggtg	tca		343

<210> 73

<211> 321

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(321)

<223> n = A,T,C or G

<400> 73

gcactgagag	gaacttcaga	gagagagaga	gagttccacc	ctgtacttgg	ggagagaaac	60
agaaggtgag	aaagtctttg	gttctgaagc	agcttctaag	atcttttcat	ttgcttcatt	120
tcaaagttcc	catgctgcca	aagtgccatc	ctttggggta	ctgttttctg	agctccagtg	180

```

ataactcatt tatacaaggg agatacccag aaaaaaagtg agcaaattctt aaaaaggtgg      240
cttgagttca gccttaaata ccatcttgaa atgacacaga gaaagaanga tgttggtgg      300
gagtggatag agaccctaac g                                           321

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<210> 74
<211> 321
<212> DNA
<213> Homo sapien

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```

<400> 74
gcactgagag gaacttcaga gagagagaga gagttccacc ctgtacttgg ggagagaaac      60
agaaggtgag aaagtctttg gttctgaagc agcttctaag atcttttcat ttgcttcatt      120
tcaaagttcc catgctgcca aagtgccatc ctttggggta ctgttttctg agtccagtg      180
ataactcatt tatacaaggg agatacccag aaaaaaagtg agcaaattctt aaaaaggtgg      240
cttgagttca gycctaaata ccatcttgaa atgamacaga gaaagaagga tgttggtgg      300
gagtggatag agaccctaac g                                           321

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<210> 75
<211> 317
<212> DNA
<213> Homo sapien

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```

<400> 75
gcactgagag gaacttcac atgcactgag aaatgcatgt tcacaaggac tgaagtctgg      60
aactcagttt ctcagttcca atcctgattc aggtgtttac cagctacaca accttaagca      120
agtcagataa ccttagcttc ctcatatgca aaatgagaat gaaaagtact catcgtgaa      180
ttgttttgag gattagaaaa acatctggca tgcagtagaa attcaattag tattcatttt      240
cattcttcta aattaaacaa ataggatttt tagtggtgga acttcagaca ccagaaatgg      300
gagtggatag agacctt                                           317

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<210> 76
<211> 244
<212> DNA
<213> Homo sapien

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<400> 76
cgttagggtc totatccact cccactactg atcaaactct atttatttaa ttatttttat      60
catactttta gttctgggat acacgtgcag catgctcagg tttgttgcag aggtatacac      120
ttgccatggt ggtttgctgc acccatcagt ccatcatcta cattaggtat ttctcctaatt      180
gctatccctc ccctagcccc ttacaccccc aacaggctct agtgtgtgaa gttcctctca      240
gtgc                                           244

```

```

<210> 77
<211> 254
<212> DNA
<213> Homo sapien

```

```

<400> 77
cgttagggtc totatccact gaaatctgaa gcacaggagg aagagaagca gtyctagtga      60
gatggcaagt tcwtttacca cactctttta catttygttt agttttaacc tttattttatg      120
gataataaag gttaatatta ataatgattt attttaaggc attcccraat ttgcataatt      180
ctccttttgg agataccctt ttatctccag tgcaagtctg gatcaaagtg atasamagaa      240
gttcctctca gtgc                                           254

```

1000
 900
 800
 700
 600
 500
 400
 300
 200
 100
 0

<210> 78
 <211> 355
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(355)
 <223> n = A,T,C or G

<400> 78
 ttcgatacag gcaaacaatga actgcaggag ggtgggtgacg atcatgatgt tgccgatggg 60
 ccggatggnc acgaagacgc actggancac gtgcttacgt ccttttgctc tgttgatggc 120
 cctgagggga cgcaggaccc ttatgaccct cagaatcttc acaacgggag atggcactgg 180
 attgantccc antgacacca gagacacccc aaccaccagn atatcantat attgatgtag 240
 ttccctgtaga nggccccctt gtggaggaaa gctccatnag ttggtcatct tcaacaggat 300
 ctcaacagtt tccgatggct gtgatgggca tagtcatant taacntgtn tcgaa 355

<210> 79
 <211> 406
 <212> DNA
 <213> Homo sapien

<400> 79
 taagagggta ccagcagaaa ggttagtata atcagatage atcttatacg agtaatatgc 60
 ctgctatttg aagtgtattt gagaaggaaa attttagcgt gctcactgac ctgcctgtag 120
 ccccgatgac agctaggatg tgcattctcc agccatcaag agactgagtc aagttgttcc 180
 ttaagtcaga acagcagact cagctctgac attctgattc gaatgacact gttcaggaat 240
 cggaatcctg tcgattagac tggacagctt gtggcaagtg aatttgctg taacaagcca 300
 gattttttta aatttatatt gtaaataatg tgtgtgtgtg tgtgtgtata tatatatata 360
 tgtacagtta tctaagttaa tttaaaagtt gtttggtacc ctctta 406

<210> 80
 <211> 327
 <212> DNA
 <213> Homo sapien

<400> 80
 tttttttttt ttactcggc tcagtctaata cctttttgta gtcactcata ggccagactt 60
 agggctagga tgatgattaa taagagggat gacataacta ttagtggcag gttagttggt 120
 tgtagggctc atggtagggg taaaaggagg gcaatttcta gatcaaataa taagaaggta 180
 atagctacta agaagaattt tatggagaaa gggacgcggg cgggggatat agggtcgaag 240
 ccgcaactgt aagggttgga tttttctatg tagccgttga gttgtggtag tcaaatgta 300
 ataattatta gtagtaagcc taggaga 327

<210> 81
 <211> 318
 <212> DNA
 <213> Homo sapien

<400> 81
 tagtttatgc ggttgattcg gcaatccatt atttgctgga ttttgtcatg tgttttgcca 60
 attgcattca taatttatta tgcatttatg cttgtatctc ctaagtcagtg gtatataatc 120
 catgcttttt atgttttgtc tgacataaac tcttatcaga gccctttgca cacagggatt 180

caataaatat taacacagtc tacatatttatt tgggtgaatat tgcataatctg ctgtactgaa	240
agcacattaa gtaacaaagg caagtggagaa gaatgaaaag cactactcac aacagttatc	300
atgattgcgc atagacta	318

<210> 82
 <211> 338
 <212> DNA
 <213> Homo sapien

<400> 82	
tcttcaacct ctactccac taatagcttt ttgatgactt ctagcaagcc tcgctaacct	60
cgccttacct ccactatta acctactggg agaactctct gtgctagtaa ccacgttctc	120
ctgatcaa atcactctcc tacttacagg actcaacata ctagtcacag ccctatactc	180
cctctacata tttaccacaa cacaatgggg ctcactcacc caccacatta acaacataaa	240
accctcattc acacgagaaa acacctcat gtccatacac ctatcccca ttctcctcct	300
atccctcaac cccgacatca ttaccgggtt ttctctct	338

<210> 83
 <211> 111
 <212> DNA
 <213> Homo sapien

<400> 83	
agccatttac caccatcca caaaaaaaaa aaaaaaaaaag aaaaatatca aggaataaaa	60
atagactttg acaaaaaagg aacattttgt ggccctgagga ggcacaccc g	111

<210> 84
 <211> 224
 <212> DNA
 <213> Homo sapien

<400> 84	
tcgggtgatg cctcctcagg ccaagaagat aaagcttcag acccctaaca catttccaaa	60
aaggaagaaa ggagaaaaaa gggcatcatc cccgttcoga agggtcaggg aggaggaaat	120
tgagggtgat tcacgagttg cggacaactc ctttgatgcc aagcgaggtg cagccggaga	180
ctgggggagag cgagccaatc aggttttgaa gtccctctca gtgc	224

<210> 85
 <211> 348
 <212> DNA
 <213> Homo sapien

<400> 85	
gcactgagag gaacttcggt ggaaacgggt ttttttcatg taaggctaga cagaagaatt	60
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gagcagactt gtaacactct twttgtggaa tttgcaagtg gagatttcag scgctttgaa	180
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<210> 142

<211> 419

<212> DNA

<213> Homo sapien

<400> 142

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<211> 402

<212> DNA

<213> Homo sapien

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<210> 144
<211> 224
<212> DNA
<213> Homo sapien

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<210> 145
<211> 111
<212> DNA
<213> Homo sapien

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<210> 146
<211> 585
<212> DNA
<213> Homo sapien

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<400> 146
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<210> 147
<211> 579
<212> DNA
<213> Homo sapien

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<222> (1)...(579)
<223> n = A,T,C or G

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<400> 147

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<210> 148

<211> 249

<212> DNA

<213> Homo sapien

<400> 148

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<210> 149

<211> 255

<212> DNA

<213> Homo sapien

<400> 149

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<210> 150

<211> 318

<212> DNA

<213> Homo sapien

<400> 150

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<210> 151

<211> 323

<212> DNA

<213> Homo sapien

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 <222> (1)...(323)
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<210> 152
 <211> 311
 <212> DNA
 <213> Homo sapien

<400> 152
 tcaagattcc ataggctgac cagtccaagg agagttgaaa tcatgaagga gagtctatct 60
 ggagagagct gtagttttga ggggttgcaaa gacttaggat ggagttgggtg ggtgtgggta 120
 gtctctaagg ttgattttgt tcataaattt catgccttga atgccttgct tgcctcacc 180
 tgggtccaagc cttagtgaac acctaaaagt ctctgtcttc ttgctctcca aacttctcct 240
 gaggatttcc tcagattgtc tacattcaga togaagccag ttggcaaaca agatgcagtc 300
 cagagggtca g 311

<210> 153
 <211> 332
 <212> DNA
 <213> Homo sapien

<400> 153
 caagattcca taggctgacc aggaggctat tcaagatctc tggcagttga ggaagtctct 60
 ttaagaaaat agtttaaaaca atttggtaaa atttttctgt cttacttcat ttctgtagca 120
 gttgatatct ggctgtcctt tttataatgc agagtgggaa ctttccctac catgtttgat 180
 aaatgttgtc caggtcccat tgccaataat gtgttggtcca aaatgcctgt ttagttttta 240
 aagacggaac tccacccttt gcttggtctt aagtatgtat ggaatgttat gataggacat 300
 agtagtagcg gtggtcagcc tatggaatct tg 332

<210> 154
 <211> 345
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(345)
 <223> n = A,T,C or G

<400> 154
 tcaagattcc ataggctgac ctggacagag atctcctggg totggcccag gacagcaggc 60
 tcaagctcag tggagaaggt ttccatgacc ctcagattcc cccaaacctt ggattgggtg 120
 acattgcac tcctcagaga gggaggagat gtangtctgg gcttccacag ggacctggta 180


```

ttttaggatac aggggtaccgc tggcctgagg cttggatcat tcanagcctg ggggtggaat 240
ggctggcagc ctgtggcccc attgaaatag gctctggggc actccctctg ttcctanttg 300
aacttgggta aggaacagga atgtggtcan cctatggaat cttga 345

```

```

<210> 155
<211> 295
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(295)
<223> n = A,T,C or G

```

```

<400> 155
gacgcttggc cacttgacac attaaacagt tttgcataat cactancatg tattttctagt 60
ttgctgtctg ctgtgatgcc ctgccctgat tctctggcgt taatgatggc aagcataatc 120
aaacgctgtt ctgttaattc caagttataa ctggcattga ttaaagcatt atctttcaca 180
actaaactgt tcttcatana acagcccata ttattatcaa attaagagac aatgtattcc 240
aatatccttt anggccaata tatttnatgt cccttaatta agagctactg tccgt 295

```

```

<210> 156
<211> 406
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(406)
<223> n = A,T,C or G

```

```

<400> 156
gacgcttggc cacttgacac tgcagtggga aaaccagcat gagccgctgc cccaaggaa 60
cctcgaagcc caggcagagg accagccatc ccagcctgca ggtaaagtgt gtcacctgtc 120
aggtgggctt ggggtgagtg ggtgggggaa gtgtgtgtgc aaagggggtg tnaatgtnta 180
tgcgtgtgag catgagtgat ggctagtgtg actgcatgtc agggagtgtg aacaagcgtg 240
cgggggtgtg tgtgcaagtg cgtatgcata tgagaatatg tgtctgtgga tgagtgcatt 300
tgaaagtctg tgtgtgtgcg tgtggtcatg anggtaantt antgactgcg caggatgtgt 360
gagtgtgcat ggaacactca ntgtgtgtgt caagtggccn ancgtc 406

```

```

<210> 157
<211> 208
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(208)
<223> n = A,T,C or G

```

```

<400> 157
tgacgcttgg ccacttgaca cactaaaggg tgttactcat cactttcttc tctcctcggt 60
ggcatgtgag tgcatttatt cacttggcac tcatttgttt ggcagtgact gtaanccana 120
tctgatgcat acaccagctt gtaaattgaa taaatgtctc taatactatg tgctcacaat 180

```

anggtanggg tgaggagaag gggagaga

208

<210> 158
 <211> 547
 <212> DNA
 <213> Homo sapien

 <220>
 <221> misc_feature
 <222> (1)...(547)
 <223> n = A,T,C or G

<400> 158
 cttcaacctc cttcaacctc ctggattcaa acaatcatcc cacctcagac 60
 tccttagtag ctgagactac agactcacgc cactacatct ggctaaattt ttgtagagat 120
 agggtttcat catgttgccc tggctgggtct caaactcctg acctcaagca atgtgcccac 180
 ctcagcctcc caaagtgtctg ggattacagg cataagccac catgcccagt ccatntttaa 240
 tctttcctac cacattctta ccacactttc ttttatgttt agatacataa atgcttacca 300
 ttatgataca attgccaca gtattaagac agtaacatgc tgcacagggt tgtagcctag 360
 gaacagtagg caataccaca tagcttaggt gtgtggtaga ctataccatc taggtttgtg 420
 taagttacac tttatgctgt ttacacaatg acaaaacat ctaatgatgc atttctcaga 480
 atgtatcctt gtcagtaagc tatgatgtac agggaaact gcccaaggac acagatattg 540
 tacctgt 547

<210> 159
 <211> 203
 <212> DNA
 <213> Homo sapien

<400> 159
 gctcctcttg ccttaccac tcacccagta tgtcagcaat tttatcrgct ttacctacga 60
 aacagcctgt atccaaacac ttaacacact cacctgaaaa gttcaggcaa caatcgctt 120
 ctcatgggtc tctctgctcc agttctgaac ctttctcttt tcctagaaca tgcatttarg 180
 tcgatagaag ttcctctcag tgc 203

<210> 160
 <211> 402
 <212> DNA
 <213> Homo sapien

<400> 160
 tgtaagtcga gcagtgtgat ggggtggaaca gggttgtaag cagtaattgc aaactgtatt 60
 taaacaataa taataatatt tagcatttat agagcacttt atatcttcaa agtacttgca 120
 aacattayct aattaaatac cctctctgat tataatctgg atacaaatgc acttaaaactc 180
 aggacagggt catgagaraa gtatgcattt gaaagtgggt gctagctatg ctttaaaaac 240
 ctatacaatg atgggraagt tagagttcag attctgttgg actgtttttg tgcatttcag 300
 ttcagcctga tggcagaatt agatcatatc tgcactcgat gactygtctt gataacttat 360
 cactgaaatc tgagtgttga tcatcacact gctcgactta ca 402

<210> 161
 <211> 193
 <212> DNA
 <213> Homo sapien

<400> 161
agcatgttga gccagacac tgaccaggag aaaaaccaac caatagaaac acgcccagac 60
actgaccagg agaaaaacca accaataaaa acaggcccg acataagaca aataataaaa 120
ttagcggaca aggacatgaa aacagctatt gtaagagcgg atatagtgg gtgtgtctgg 180
gctcaacatg cta 193

<210> 162
<211> 147
<212> DNA
<213> Homo sapien

<400> 162
tggtgagccc agacactgac caggagaaaa accaaccaat aaaaacaggc cgggacataa 60
gacaaaataat aaaattagcg gacaaggaca tgaaaacagc tattgtaaga gcggatatag 120
tggtgtgtgt ctgggctcaa catgcta 147

<210> 163
<211> 294
<212> DNA
<213> Homo sapien

<400> 163
tagcatgttg agcccagaca caaatctttc cttaagcaat aaatcatttc tgcatatgtt 60
tttaaaacca cagctaagcc atgattattc aaaaggacta ttgtattggg tattttgatt 120
tggtgtctta tctccctcac attatcttca tttctatcat tgacctctta tcccagagac 180
tctcaaactt ttatgtttata caaatcacat tctgtctcaa aaaatatctc acccaacttct 240
cttctgtttc tgcgtgtgta tgtgtgtgtg tgtgtgtctg ggctcaacat gcta 294

<210> 164
<211> 412
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(412)
<223> n = A,T,C or G

<400> 164
cgggattggc tttgagctgc agatgctgcc tgtgaccgca cccggcgtgg aacagaaaagc 60
cacctggctg caagtgcgcc agagccgccc tgactacgtg ctgctgtggg gctggggcgt 120
gatgaactcc accgccctga aggaagccca ggccaccgga taccgccg acagatgta 180
cggcgtgtgg tgggccggtg cggagcccgga tgtgctgtac gtgggcgaag gcgccaaggg 240
ctacaacgcg ctggtctctga acggtacgg cagcgagtcc aaggtgatcc angacatcct 300
gaaacacgtg cagcacaagg gccagggcac ggggcccaaa gacgaagtgg gctcgtgtgt 360
gtacaccgcg ggcgtgatca tccagatgct ggacaaggtg tcaatcacta at 412

<210> 165
<211> 361
<212> DNA
<213> Homo sapien

<400> 165
ttgacacctt gtccagcatc tgcattctgat gagagcctca gatggctacc actaatggca 60

```

gaaggcaaag gagaacaggc attgtatggc aagaaaggaa gaaagagaga ggggagaaag 120
gtgctaggtt cttttcaaca accagttctt gatggaactg agagtaagag ctcaaggcca 180
ggtgtggtga ctccaaccag taatcccaac attttaggag gctgaggcag gcagatgtct 240
tgaccccatg agtttgtgac cagcctgaac aacatcatga gactccatct ctacaataat 300
tacaaaaatt aatcaggcat tgtggtatgc cctgtagtcc cagatgctgg acaagggtgc 360
a 361

```

```

<210> 166
<211> 427
<212> DNA
<213> Homo sapien

```

```

<400> 166
twgactgact catgtccctt acacccaact atcttctcca ggtggccagg catgatagaa 60
tctgatcctg acttagggga atattttctt tttacttccc atcttgattc cctgccggtg 120
agtttcttgg ttccagggtaa gaaaggagct caggccaaag taatgaacaa atccatcctc 180
acagacgtac agaataagag aacwtggacw tagccagcag aacmcaaktg aaamcagaac 240
mcttamctag gatracaamc merraratar ktgcycmcmc wtataataga aaccaaactt 300
gtatctaatt aaatatattt ccacygtcag ggcatttagt gttttgataa atacgctttg 360
gctaggattc ctgagggttag aatggaaraa caattgcamc gagggtaggg gacatgagtc 420
aktctaa 427

```

```

<210> 167
<211> 500
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(500)
<223> n = A,T,C or G

```

```

<400> 167
aacgtcgcat gctcccggcc gccatggccg cgggatagac tgactcatgt cccctaagat 60
agaggagaca cctgctaggt gtaaggagaa gatgggttagg tctacggagg ctccagggtg 120
ggagtagttc cctgctaagg gagggtagac tgttcaacct gttcctgctc cggcctccac 180
tatagcagat gcgagcagga gtaggagaga gggaggtaag agtcagaagc ttatgttggt 240
tatgcgggga aacgcrtat cgggggcagc cragttatta ggggacantr tagwyartcw 300
agntagcatc caaagcgngg gagttntccc atatggttgg acctgcaggc ggccgcatta 360
gtgattagca tgtgagcccc agacacgcat agcaacaagg acctaaactc agatcctgtg 420
ctgattactt aacatgaatt attgtattta tttaacaact ttgagttatg aggcataatta 480
ttaggtccat attacctgga 500

```

```

<210> 168
<211> 358
<212> DNA
<213> Homo sapien

```

```

<400> 168
ttcatcgctc ggtgactcaa gcctgtaatc ccagaacttt gggaggccga ggggagcaga 60
tcacctgagg ttgggagttt gagaccagcc tggccaacat ggtgacaacc cgtctctgct 120
aaaaatacaa aaattagcca agcatggtgg catgcacttg taatcccagc tactcgggag 180
gctgaggcag gagaatcact tgaggccagg aggcagaggt tgcagtgagg cagaggttga 240
gatcatgcc a ctgactcca gcctgggcaa cagagtaaga ctccatctca aaaaaaaaaa 300

```

aaaaaaagaa tgatcagagc cacaaatata gaaaaccttg agtcaccgag cgatgaaa 358

<210> 169
<211> 1265
<212> DNA
<213> Homo sapien

<400> 169

ttctgtccac	accaatotta	gagctctgaa	agaatttgct	tttaaataatc	ttttaaatagt	60
aacatgtatt	ttatggacca	aattgacatt	ttcgactatt	ttttcccaaa	aaaagtcagg	120
tgaatttcag	cacactgagt	tgggaatttc	ttatcccaga	agwccgcacg	agcaatttca	180
tatttattta	agattgattc	catactccgt	tttcaaggag	aatccctgca	gtctccttaa	240
aggtagaaca	aatactttct	atTTTTTTTT	caccattgtg	ggattggact	ttaagagggtg	300
actctaaaaa	aacagagaac	aaatatgtct	cagttgtatt	aagcacggac	ccatattatc	360
atatccactt	aaaaaaatga	tttcctgtgc	accttttggc	aacttctctt	ttcaatgtag	420
ggaaaaactt	agtcaccctg	aaaaccacaca	aaataaataa	aacttgtaga	tgtgggcaga	480
argtttgggg	gtggacattg	tatgtgttta	aattaaaccc	tgtatcactg	agaagctggt	540
gtatgggtca	gagaaaatga	atgcttagaa	gctgttcaca	tcttcaagag	cagaagcaaa	600
ccacatgtct	cagctatatt	attattttatt	ttttatgcat	aaagtgaatc	atttcttctg	660
tattaatttc	caaagggttt	taccctctat	ttaaatgctt	tgaaaaacag	tgcattgaca	720
atgggttgat	atTTTTCTTT	aaaagaaaaa	tataattatg	aaagccaaga	taatctgaag	780
cctgttttat	tttaaaactt	tttatgttct	gtgggtgatg	ttgtttgttt	gtttgtttct	840
atTTTgttgg	TTTTTTactt	tgtTTTTTgt	TTTgtTTTgt	TTTgtTTtdg	catactacat	900
gcagtttctt	taaccaatgt	ctgttttggt	aatgtaatta	aagttgttaa	tttatatgag	960
tgcatttcaa	ctatgtcaat	ggtttcttaa	tatttattgt	gtagaagtac	tggtaatttt	1020
tttatttaca	atatgtttta	agagataaca	gtttgatatg	ttttcatgtg	tttatagcag	1080
aagttattta	tttctatggc	attccagcgg	atattttggt	gtttgcgagg	catgcagtca	1140
atattttgta	cagttagtgg	acagtattca	gcaacgcctg	atagcttctt	tggccttatg	1200
ttaaataaaa	agacctgttt	gggatgtaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	1260
aaaaa						1265

<210> 170
<211> 383
<212> DNA
<213> Homo sapien

<400> 170

tgtaagtcga	gcagtgtgat	gacgatattc	ttcttattaa	tgtggtaatt	gaacaaatga	60
tctgtgatac	tgatcctgag	ctaggaggcg	ctgttcagtt	aatgggactt	cttcgtactc	120
taattgatcc	agagaacatg	ctggctacaa	ctaataaaac	cgaaaaaagt	gaattttctaa	180
atTTTTtcta	caaccattgt	atgcatgttc	tcacagcacc	acttttgacc	aatacttcag	240
aagacaaatg	tgaaaaggat	aatatagttg	gatcaaacaa	aaacaacaca	atttgtcccg	300
ataattatca	aacagcacag	ctacttgcct	taatttttaga	gttactcaca	ttttgtgtgg	360
aacatcacac	tgctcgactt	aca				383

<210> 171
<211> 383
<212> DNA
<213> Homo sapien

<400> 171

tgggcacctt	caatatcgca	agttaaaaat	aatgttgagt	ttattatact	tttgacctgt	60
ttagctcaac	agggtgaagg	catgtaaaga	atgtggactt	ctgaggaatt	ttctttttaa	120
aagaacataa	tgaagtaaca	ttttaattac	tcaaggacta	cttttggttg	aagtttataa	180

```

tctagataacc tctacttttt gtttttgctg ttcgacagtt cacaaagacc ttcagcaatt 240
tacagggtaa aatcgttgaa gtagtggagg tgaaactgaa atttaaaatt attctgtaaa 300
tactataggg aaagaggctg agcttagaat cttttggtg ttcattgtgt ctgtgctctt 360
atcatcacac tgctcgactt aca 383

```

```

<210> 172
<211> 699
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(699)
<223> n = A,T,C or G

```

```

<400> 172
tcgggtgatg cctcctcagg cttgtcggtta gtgtacacag agctgctcat gaagcgacag 60
cggtcgcccc tggcaacttca gaacctcttc ctctacactt ttggtgcgct totgaatcta 120
ggtctgcatg ctggcgggcg ctctggccca ggctcctctg aaagtttctc aggatgggca 180
gcactcgctg tgctgagcca ggcactaaat ggactgctca tgtctgctgt catggagcat 240
ggcagcagca tcacacgcct ctttgtggtg tctgctcgc tgggtgtcaa cgccgtgctc 300
tcagcagtc tgctacggct gcagctcaca gccgccttct tcctggccac attgctcatt 360
ggcctggcca tgcgcctgta ctatggcagc cgctagctcc tgacaacttc caccctgatt 420
ccggaccctg tagattgggc gccaccacca gatccccctc ccaggccttc ctccctctcc 480
catcagcggc cctgtaacaa gtgccttctg agaaaagctg gagaagtgag ggcagccagg 540
ttattctctg gaggttgggt gatgaagggg tacccttagg agatgtgaag tgtgggtttg 600
gttaaggaaa tgcttaccat cccccacccc caaccaagtt nttccagact aaagaattaa 660
ggtaacatca atacctaggc ctgaggaggc atcacccga 699

```

```

<210> 173
<211> 701
<212> DNA
<213> Homo sapien

```

```

<400> 173
tcgggtgatg cctcctcagg ccagatcaaa cttgggggtg aaaactgtgc aaagaaatca 60
atgtcggaga aagaattttg caaaagaaaa atgcctaata agtactaatt taataggtca 120
cattagcagt ggaagaagaa atgttgatat tttatgtcag ctattttata atcaccagag 180
tgcttagctt catgtaagcc atctcgtatt cattagaaat aagaacaatt ttattcgtcg 240
gaaagaactt ttcaatttat agcatcttaa ttgctcagga ttttaaattt tgataaagaa 300
agctccactt ttggcaggag tagggggcag ggagagagga ggctccatcc acaaggacag 360
agacaccagg gccagtaggg tagctgggtg ctggatcagt cacaacggac tgacttatgc 420
catgagaaga aacaacctcc aaatctcagt tgcttaatac aacacaagct catttcttgc 480
tcacgttaca tgtcctatgt agatcaacag caggtgactc agggacccag gctccatctc 540
catatgagct tccatagtca ccaggacacg ggctctgaaa gtgtcctcca tgcagggaca 600
catgcctctt cctttcattg ggcagagcaa gtcacttatg gccagaagtc aactgcagg 660
gcagtgccat cctgctgtat gcctgaggag gcatcacccg a 701

```

```

<210> 174
<211> 700
<212> DNA
<213> Homo sapien

```

```

<220>

```

```

<221> misc_feature
<222> (1)...(700)
<223> n = A,T,C or G

```

```
<400> 174
```

```

tcgggtgatg cctcctcang cccctaaatc agagtccagg gtcagagcca caggagacag      60
ggaaagacat agattttaac cggccccctt caggagattc tgaggctcag ttcactttgt      120
tgcagtttga acagaggcag caaggetagt ggtaggggc acggtctcta aagctgcaact      180
gcctggatct gcctcccagc tctgccagga accagctgcg tggccttgag ctgctgacac      240
gcagaaagcc cctgtgggac ccagtctcct cgtctgtaag atgaggacag gactctagga      300
accttttccc ttggtttggc ctcactttca caggctccca tcttgaactc tatctactct      360
tttctgaaa ccttgtaaaa gaaaaaagtg ctagcctggg caacatggca aaacctgtc      420
tctacaaaa atacaaaaat tagttgggtg tggtagcatg tgcctgtagt ccagccact      480
tgggaggtgc tgaggtggga ggatcacttg agcccgagg gtggaggttg cagtgaagcca      540
agatcatgcc actgcactcc agcctgagta atagagtaag actctgtctc aaaaacaaca      600
acaacaacag tgagtgtgcc tctgtttccg ggtaggatgg ggcaccacat ttatgcatct      660
ctcagatttg gacgctgcag cctgaggagg catcaccgca      700

```

```

<210> 175
<211> 484
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(484)
<223> n = A,T,C or G

```

```
<400> 175
```

```

tatagggcga attgggcccg agttgcatgn tcccgccgc catggccgcg ggattcgggt      60
gatgcttctt caggcttgtc tgccacaagc tacttctctg agctcagaaa gtgccccttg      120
atgagggaaa atgtcctact gcactgcgaa tttctcagtt ccattttacc tcccagtcct      180
ccttctaacc cagttaataa attcattcca caagtattta ctgattacct gcttgtgccca      240
gggactattc tcaggctgaa gaaggtggga ggggagggcg gaacctgagg agccacctga      300
gccagcttta tatttcaacc atggctggcc catctgagag catctcccca ctctcgccaa      360
cctatcgggg catagcccag ggatgcccc aggcggccca ggtagatgc gtccctttgg      420
cttgtcagtg atgacataca ccttagctgc ttagctgggt ctggcctgag gaggcatcac      480
ccga      484

```

```

<210> 176
<211> 432
<212> DNA
<213> Homo sapien

```

```
<400> 176
```

```

tcgggtgatg cctcctcagg gctcaaggga tgagaagtga cttctttctg gagggaccgt      60
tcatgccacc caggatgaaa atggataggg acccacttgg aggacttgct gatatgttg      120
gacaaatgcc aggtagcggg attggtactg gtccaggagt tatccaggat agattttcac      180
ccaccatggg acgtcatcgt tcaaatcaac tcttcaatgg ccatggggga cacatcatgc      240
ctccacacac atcgcatgtt ggagagatgg gaggaagtt tatgaaaagc caggggctaa      300
gccagctcta ccataaccag agtcagggac tcttatccca gctgcaagga cagtcgaagg      360
atatgccacc tcggttttct aagaaaaggac agcttaatgc agatgagatt agcctgagga      420
ggcatcacc ga      432

```

<210> 177
 <211> 788
 <212> DNA
 <213> Homo sapien

<400> 177
 tagcatgttg agcccagaca cagtagcatt tgtgcccaatt tctggttgga atggtgacaa 60
 catgctggag ccaagtgcta acatgccttg gttcaaggga tggaaagtca cccgtaagga 120
 tggcaatgcc agtggaaacca cgtgcttgga ggctctggac tgcatectac caccaactcg 180
 cccaactgac aagcccttgc gcctgcctct ccaggatgtc tacaaaattg gtggtattgg 240
 tactgttccct gttggccgag tggagactgg tgttctcaaa cccggtatgg tggcacctt 300
 tgctccagtc aacgttacaa cggaaagtaa atctgtcgaa atgcaccatg aagctttgag 360
 tgaagctctt cctggggaca atgtgggctt caatgtcaag aatgtgtctg tcaaggatgt 420
 tcgtcgtggc aacgttgctg gtgacagcaa aaatgaccca ccaatggaag cagctggctt 480
 cactgctcag gtgattatcc tgaaccatcc aggccaaata agtgccggct atgccctgt 540
 attggattgc cacacggctc acattgcatg caagtttgct gagctgaagg aaaagattga 600
 tcgccgttct ggtaaaaagc tggaaagatgg coctaaattc ttgaagtctg gtgatgctgc 660
 cattgttgat atggttctcg gcaagcccat gtgtgttgag agcttctcag actatccacc 720
 tttgggtcgc tttgctgttc gtgatatgag acagacagtt gcgggtgggtg tctgggctca 780
 acatgcta 788

<210> 178
 <211> 786
 <212> DNA
 <213> Homo sapien

<400> 178
 tagcatgttg agcccagaca cctgtgtttc tgggagctct ggcagtggcg gattcatagg 60
 cacttgggct gcactttgaa tgacacactt ggctttatta gattcactag tttttaaaaa 120
 attgttgctt gtttcttttc attaaagggt taatcagaca gatcagacag cataattttg 180
 tatttaatga cagaaacggt ggtacatttc ttcatgaatg agcttgcatc ctgaagcaag 240
 agcctacaaa aggcacttgt tataaatgaa agttctggct ctgagggcca gtactctgga 300
 gtttcagagc agccagtgtg tgttccagtc agtgatgcct agttatatag aggaggagta 360
 cactgtgcac tcttctaggt gtaagggtat gcaacttttg atcttaaaat tctgtacaca 420
 tacacacttt atatatatgt atgtatgtat gaaaacatga aattagtttg tcaaatatgt 480
 gtgtgttttag tatttttagct tagtgcaact atttccacat tattttattaa attgatctaa 540
 gacactttct tgttgacacc ttgaatatta atgttcaagg gtgcaatgtg tattccttta 600
 gattgttaaa gcttaattac tatgatttgt agtaaattaa cttttaaaat gtatttgagc 660
 ccttctgtag tgtcgtaggg ctcttacagg gtgggaaaga ttttaatttt ccagttgcta 720
 attgaacagt atggcctcat tatatatattt gatttatagg agtttgtgtc tgggctcaac 780
 atgcta 786

<210> 179
 <211> 796
 <212> DNA
 <213> Homo sapien

<400> 179
 tagcatgttg agcccagaca ctggttacaa gaccagacct gcttcctcca tatgtaaaca 60
 gcttttaaaa agccagtga cttttttaat actttggcaa ccttctttca caggcaaaga 120
 acacccccat ccgcccttg tttggagtgc agagtttggc tttggttctt tgccttgctt 180
 ggagtatact tctaattcct gttgtcctgc acaagctgaa taccgagcta cccaccgcca 240
 ccaggccag gtttccactc atttattact ttatgtttct gttccattgc tgggtccacag 300
 aaataagttt tcctttggag gaatgtgatt ataccctttt aatttctctc ttttgctttt 360


```

tittaatatc attggtatgt gtttggccca gaggaactg aaattcacca tcatcttgac 420
tggcaatccc attaccatgc tttttttaaa aaacgtaatt tttcttgctt tacattggca 480
gagtagccct tcctggctac tggcttaatg tagtcaactca gtttctaggt ggcattaggc 540
atgagacctg aagcacagac tgtcttacca caaaagggtga caagatctca aaccttagcc 600
aaagggctat gtcagggttc aatgctatct gcttctgttc ctgctcaactg ttctggattt 660
tgtccttctt catccctagc accagaattt cccagtctcc ctccctacct tcccttggtt 720
taattctaatt ctatcagcaa aataactttt caaatgtttt aaccggtatc tccatgtgtc 780
tgggctcaac atgcta 796

```

```

<210> 180
<211> 488
<212> DNA
<213> Homo sapien

```

```

<400> 180
ggatgtgctg caaggcgatt aagttgggta acgccagggt tttcccagtc acgacgttgt 60
aaaacgacgg ccagtgaatt gtaatacgac tcaactatagg gcgaattggg cccgacgtcg 120
catgctcccg gcgcgatgg cgcggggata gcatgttgag cccagacacc tgcaggatcat 180
ttggagagat ttttcacgtt accagcttga tgggtctttt caggaggaga gacactgagc 240
actcccaagg tgagggttgaa gatttctctt agatagccgg ataagaagac taggagggat 300
gcctagaaaa tgattagcat gcaaatttct acctgccatt tcagaactgt gtgtcagccc 360
acattcagct gcttcttggt aactgaaaag agagagggtat tgagactttt ctgatggccg 420
ctctaacatt gtaacacagt aatctgtgtg tgtgtgggtg tgtgtgtgtg tctgggctca 480
acatgcta 488

```

```

<210> 181
<211> 317
<212> DNA
<213> Homo sapien

```

```

<400> 181
tagcatgttg agcccagaca cggcgacggt acctgatgag tgggggtgatg gcacctgtga 60
aaaggaggaa cgtcatcccc catgatattg gggaccagga tgatgaacca tggctccgcg 120
tcaatgcata ttaatccat gatactgctg attggaagga cctgaacctg aagtttgtgc 180
tgcaggttta tcgggactat tacctcacgg gtgatcaaaa cttcctgaag gacatgtggc 240
ctgtgtgtct agtaagggtat gcacatgcag tggccagtgt gccaggggta tggttgggtg 300
ctgggctcaa catgcta 317

```

```

<210> 182
<211> 507
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(507)
<223> n = A,T,C or G

```

```

<400> 182
tagcatgttg agcccagaca ctggctgtta gccaaatcct ctctcagctg ctccctgtgg 60
tttggtgact caggattaca gaggcatact gtttcaggga acaaaaagat tttagctgcc 120
agcagagagc accacatata ttagaatggg aaggactgcc acctccttca agaacaggag 180
tgagggtggg ggtgaatggg aatggaagcc tgcattccct gatgcatttg tgctctctca 240
aatcctgtct tagtcttagg aaaggaagta aagtttcaag gacggttccg aactgctttt 300

```

tgtgtctggg	ctcaacatgc	tatcccgcgg	ccatggcggc	cgggagcatg	cgacgtcggg	360
cccaattcgc	cctatagtga	gtcgtattac	aattcactgg	ccgtcgtttt	acaacgtcgt	420
gactgggaaa	accctggcgt	tacccaactt	aatcgccttg	cagcacaatcc	ccctttccca	480
gctggcgtaa	tancgaaaag	gcccgcga				507

<210> 183
 <211> 227
 <212> DNA
 <213> Homo sapien

<400> 183						
gattttacgct	gcaacactgt	ggaggtagcc	ctggagcaag	gcaggcatgg	atgcttctgc	60
aatccccaaa	tggagcctgg	tatttcagcc	aggaatctga	gcagagcccc	ctctaattgt	120
agcaatgata	agttattctc	tttgttcttc	aaccttccaa	tagccttgag	cttcagggg	180
agtgtcgtaa	atcattacag	cctggtctcc	acagtgttgc	agcgtaa		227

<210> 184
 <211> 225
 <212> DNA
 <213> Homo sapien

<400> 184						
ttacgctgca	acactgtgga	gcagattaac	atcagacttt	tctatcaaca	tgactggggg	60
tactaaaaag	acaacaaatc	aatggcttca	aaagtctaag	gaataatttc	gatacttcaa	120
ctttataaaa	cctgacaaaa	ctatcaatca	agcataaaga	cagatgaaga	acatttccag	180
atatttgcca	atcagatatt	ttacctccac	agtgttgcag	cgtaa		225

<210> 185
 <211> 597
 <212> DNA
 <213> Homo sapien

<400> 185						
ggccccgacgt	cgcattgctcc	cggccgcccat	ggccgcggga	ttcgttaggg	tctctatcca	60
ctgggaccca	taggctagtc	agagtattta	gagttgagtt	cctttctgct	tcccagaatt	120
tgaaagaaaa	ggagttaggt	gatagagctg	agagatcaga	tttgectctg	aagcctgttc	180
aagatgtatg	tgctcagacc	ccaccactgg	ggcctgtggg	tgaggctctg	ggcatctatt	240
tgaatgaatt	gctgaagggg	agcactatgc	caaggaaggg	gaacccatcc	tggcactggc	300
acaggggtca	ccttatccag	tgctcagtg	ttctttgctg	ctacctggtt	ttctctcata	360
tgtgaggggc	aggtagaag	aagtgccrg	tgttgtgcga	gttttagaac	atctaccagt	420
aagtggggaa	gtttcacaaa	gcagcagctt	tgttttgtgt	attttcacct	tcagttagaa	480
gaggaaggct	gtgagatgaa	tgtagttga	gtggaaaaga	cgggtaagct	tagtggatag	540
agaccctaac	gaatcactag	tgcggccgcc	ttgcaggtcg	accatatggg	agagctc	597

<210> 186
 <211> 597
 <212> DNA
 <213> Homo sapien

<400> 186						
ggccccgaagt	tgcatgttcc	cggccgcccat	ggccgcggga	ttcgttaggg	tctctatcca	60
ctacctaaaa	aatccccaac	atataactga	actcctcaca	cccaattgga	ccaatccatc	120
accccagagg	cctacagatc	ctcctttgat	acataagaaa	atttcccaa	actacctaac	180
tatatcattt	tgcaagattt	gttttaccaa	attttgatgg	cctttctgag	cttgtcagtg	240

```

tgaaccacta ttacgaacga tcggatatta actgcccctc accgtccagg tgtagctggc 300
aacatcaagt gcagtaaata ttcattaagt ttccacctac taagggtgctt aaacacccta 360
gggtgccatg tcggtagcag atcttttgat ttgtttttat ttcccataag ggtcctgttc 420
aaggtcaatc atacatgtag tgtgagcagc tagtcactat cgcattgactt ggagggtgat 480
aatagaggcc tcctttgctg ttaaagaact cttgtcccag cctgtcaaag tggatagaga 540
ccctaacgaa tcactagtgc ggccgcctgc aggtcgacca tatgggagag ctcccaa 597

```

```

<210> 187
<211> 324
<212> DNA
<213> Homo sapien

```

```

<400> 187
tcgttagggg ctctatccac ttgcaggtaa aatccaatcc tgtgtatatc ttatagtctt 60
ccatatgtag tggttcaaga gactgcagtt ccagaaagac tagccgagcc catccatgtc 120
ttccacttaa ccctgccttg ggttacacat cttaactttt ctgttcaagt ttctctgtgt 180
agtttatagc atgagtattg ggawaatgcc ctgaaacctg acatgagatc tgggaaacac 240
aaacttactc aataagaatt tctcccatat ttttatgatg gaaaaatttc acatgcacag 300
aggagtggat agagacccta acga 324

```

```

<210> 188
<211> 178
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(178)
<223> n = A,T,C or G

```

```

<400> 188
gcgcggggat tcgggggtgat acctcctcat gccaaaatac aacgtntaat ttcacaactt 60
gccttccaat ttacgcattt tcaatttgct ctccccattt gttgagtcac aacaaacacc 120
attgcccgaga aacatgtatt acctaacatg cacatactct taaaactact catccctt 178

```

```

<210> 189
<211> 367
<212> DNA
<213> Homo sapien

```

```

<400> 189
tgacaccttg tccagcatct gacacagtct tggctcttgg aaaatatttg ataaatgaaa 60
atgaatttct ttagcaagtg gtataagctg agaatatacg tatcacatat cctcatttcta 120
agacacattc agtgtccctg aaattagaat aggacttaca ataagtgtgt tcactttctc 180
aatagctggt attcaattga tggtaggcct taaaagtcaa agaaatgaga gggcatgtga 240
aaaaaagctc aacatcactg atcattagaa aacttccatt caaaccccca atgagatacc 300
atctcatacc agtcagaatg gctattatta aaaagtcaaa aaataacaga tgctggacaa 360
ggtgtca 367

```

```

<210> 190
<211> 369
<212> DNA
<213> Homo sapien

```

<220>
 <221> misc_feature
 <222> (1)...(369)
 <223> n = A,T,C or G

<400> 190
 gacaccttgt ccagcatctg acaacgctaa cagcctgagg agatctttat ttatttattt 60
 agtttttact ctggctaggc agatggtggc taaaacattc atttaccat ttattcattt 120
 aattgttctt gcaaggccta tggatagagt attgtccagc actgctctgg aagctaggag 180
 catggggatg aacaagatag gctacatcct gttccacag aacttccact ttagtctggg 240
 aaacagatga tatatacaaa tatataaatg aattcaggta gttttaagta cgaaaagaat 300
 aagaaagcag agtcatgatt tanaatgctg gaaacagggg ctattgcttg agatattgaa 360
 ggtgcccaa 369

<210> 191
 <211> 369
 <212> DNA
 <213> Homo sapien

<400> 191
 tgacaccttg tccagcatct gcacagggaa aagaaactat tatcagagtg aacaggcaac 60
 ctacagaatg ggagaaaatt tttgcaatct atccatctga caaagggcta atatccagaa 120
 tctacaaaga acttatacaa atttacaaga aacaaacaaa caaacaactc ctcaaaaagt 180
 ggggtgaagga tgtgaacaga cactttcaca aagaagacat ttatggggcc acaaaacata 240
 tgaaaaaaag ctcacatca ctggtcacta gataaatgca aatcaaaacc acaatgagat 300
 accatctcat tccagttaga atggcaatca ttaaaaagtc aggaacaac agatgctgga 360
 caaggtgtc 369

<210> 192
 <211> 449
 <212> DNA
 <213> Homo sapien

<400> 192
 tgacgcttgg ccacttgaca cttcatcttt gcacagaaaa acttctttac agatttaatt 60
 caagactggt ctagtacag tctccagac attttttcat ttgttccata tacgtggaat 120
 tttaaaatca tgtttcatca gtttgaaatg atttgggctg ctaatcaaca caattggatc 180
 gactgttcta ctaaacaaca ggaaaatgtg tatctggcag cctgtggaga aacactaaac 240
 attgattttt ctttgccttt tacggacttt gttccagcta catgtaatac caagttctct 300
 ttaagaggag aagatgttga tcttcatttg tttctaccag actgccacco tagtaaatat 360
 tctttattta tgctggtaaa aaattgccat ccaaataaga tgattcatga tactggtatt 420
 cctgctgagt gtcaagtggc caagcgtca 449

<210> 193
 <211> 372
 <212> DNA
 <213> Homo sapien

<400> 193
 tgacgcttgg ccacttgaca ccagggatgt akcagttgaa tataatcctg caattgtaca 60
 tattggcaat tcccatcaa acattctaga aagagacaac caggattgct aggccataaa 120
 agctgcaata aataactggg aattgcagta atcatttcag gccattcaa tccagtttgg 180
 ctacagagtg cctttggctg agagaagagg tgagatataa tgtgttttct tgcaacttct 240
 tggaagaata actccacaat agtctgagga ctagatacaa acctatttgc cattaaagca 300

```
ccagagtctg ttaattccag tactgataag tgttggagat tagactccag tgtgtcaagt 360
ggccaagcgt ca 372
```

```
<210> 194
<211> 309
<212> DNA
<213> Homo sapien
```

```
<220>
<221> misc_feature
<222> (1)...(309)
<223> n = A,T,C or G
```

```
<400> 194
tgacgcttgg ccacttgaca cttatgtaga atccatcgtg ggetgatgca agccctttat 60
ttaggcttag tgttgtgggc accttcaata tcacactaga gacaaacgcc acaagatctg 120
cagaaacatt cagttctgan cactcgaatg gcaggataac tttttgtgtt gtaatccttc 180
acatatataa aaacaaactc tgcantctca cgttacaaaa aaacgtactg ctgtaaaata 240
ttaagaaggg gtaaaggata ccatctataa caaagtaact tacaactagt gtcaagtggc 300
caagcgtca 309
```

```
<210> 195
<211> 312
<212> DNA
<213> Homo sapien
```

```
<220>
<221> misc_feature
<222> (1)...(312)
<223> n = A,T,C or G
```

```
<400> 195
tgacgcttgg ccacttgaca cccaatctcg cacttcatcc tcccagcacc tgatgaagta 60
ggactgcaac tatccccact tcccagatga ggggaccaan gtacacatta ggaccggat 120
gggagcacag atttgtccga tcccagactc caagcactca gcgtcactcc aggacagcgg 180
ctttcagata aggtcacaaa catgaatggc tccgacaacc ggagtcagtc cgtgctgagt 240
taaggcaatg gtgacacgga tgcacgtgtn acctgtaatg gttcatcgta agtgtcaagt 300
ggccaagcgt ca 312
```

```
<210> 196
<211> 288
<212> DNA
<213> Homo sapien
```

```
<400> 196
tgtatgaag tagtgggtctc ctcagccatg cagaactgtg actcaattaa acctctttcc 60
tttatgaatt acccaatctc gggtagtgtc tttatagtag tgtgagaatg gactaataca 120
agtacatttt acttagtaat aataataaac aaatatatta catttttgtg tatttactac 180
accatatttt ttattgttat tgtagtgtac accttctact tattaaaaga aataggcccg 240
aggcggggcag atcacgaggt caggagatgg agaccactac gtcgatac 288
```

```
<210> 197
<211> 289
<212> DNA
```

<213> Homo sapien

<400> 197

ttgggcacct	tcaatatcat	gacaggtgat	gtgataacca	agaaggctac	taagtgatta	60
atgggtgggt	aatgtataca	gagtaggtac	actggacaga	ggggtaattc	atagccaagg	120
caggagaagc	agaatggcaa	aacatttcat	cacactactc	aggatagcat	gcagtttaaa	180
acctataagt	agttttat	tggaattttc	cacttaatat	tttcagactg	caggtaacta	240
aactgtggaa	cacaagaaca	tagataaggg	gagaccacta	cgtcgatac		289

<210> 198

<211> 288

<212> DNA

<213> Homo sapien

<400> 198

gtatcgacgt	agtggctctc	caagcagtg	gaagaaaacg	tgaaccaatt	aaaatgtatc	60
agatacccca	aagaaaggcg	cttgagtaaa	gattccaagt	gggtcacaa	ctcagatctt	120
aaaattcagg	ctgtcaaaga	gatttgctat	gaggttgctc	tcaatgactt	caggcacagt	180
cggcaggaga	ttgaagccct	ggccattgtc	aagatgaagg	agctttgtgc	catgtatggc	240
aagaaagacc	ccaatgagcg	ggactcctgg	agaccactac	gtcgatac		288

<210> 199

<211> 1027

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1027)

<223> n = A,T,C or G

<400> 199

gcttttttggg	aaaaacncaa	ntgggggaaa	gggggnttnn	tngcaagggg	ataaaggggg	60
aancccgagg	tttccccatt	cagggagggtg	taaaaagncg	gccaggggat	tgtaanagga	120
ttcaataata	gggggaatgg	gccnngaagt	tgcaagggtc	cngcccgcca	tgncgcggg	180
atttagtgac	attacgacgs	tggttaataaa	gtgggsccaa	waaatatttg	tgatgtgatt	240
tttsgaccag	tgaaccatt	gwacaggacc	tcatttctcty	tgagatgrta	gccataatca	300
gataaaagrt	tagaagtytt	tctgcacgtt	aacagcatca	ttaaattggag	tggtatcacc	360
aatttcaccc	tttgttagcc	gataccttcc	cottgaaggc	attcaattaa	gtgaccaatc	420
gtcatacgag	aggggatggc	atggggattg	atgatgatat	caggggtgat	accttcacag	480
gtgaaaggca	tatcctcttg	tctatactga	ataccacaag	tacccttttg	accatgtcga	540
ctagcaaatt	tgtctccaat	ctgtgtwatc	cctaacagag	cgtaccctta	ttttacaaaa	600
tttatatcct	tcctgattga	gagttacat	aacctgatcc	acaatgcccg	tctcgctwtg	660
tctgagaaaa	gtgctacagt	ctctcttggt	atagcgtcta	ttggtgctct	ccaattcatc	720
ttcatttttc	aggcaagggtg	aactgttttg	cctataataa	cmtcatctcc	tgatacmcga	780
aacccckgga	rctatcaaac	catcatcatc	cagcgttckt	watgtymcta	aatccctatt	840
ggggccgcct	gcaggtcaac	atatnggaaa	acccccacc	ccttnggagc	ntaccttgaa	900
ttttccatat	gtcccntaaa	ttanctngnc	ttanccctggc	cntaacctnt	tccggtttta	960
attgtttcgc	ccccnttcc	cnccttnna	accggaaacc	ttaatttttna	accnggggtt	1020
cctatcc						1027

<210> 200

<211> 207

<212> DNA

<213> Homo sapien

<400> 200

agtgacatta	cgacgctggc	catcttgaat	cctagggcat	gaagttgcc	caaagttcag	60
cacttggtta	agcctgatcc	ctctggttta	tcacaaagaa	taggatggga	taaagaaagt	120
ggacacttaa	ataagctata	aattatatgg	tccttgtcta	gcaggagaca	actgcacagg	180
tatactacca	gcgtogtaat	gtcacta				207

<210> 201

<211> 209

<212> DNA

<213> Homo sapien

<400> 201

tgggcacctt	caatatctat	taaaagcaca	aatactgaag	aacacaccaa	gactatcaat	60
gaggttacat	ctggagtcct	cgatatatca	ggaaaaaatg	aagtgaacat	tcacagagtt	120
ttacttcttt	gggaactcaa	atgctagaaa	agaaaagggg	gcctcttttc	tctggcttcc	180
tggtcctatc	cagcgtcgta	atgtcacta				209

<210> 202

<211> 349

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(349)

<223> n = A,T,C or G

<400> 202

ntacgtgca	acactgtgga	gccactgggt	tttattcccg	gcagggttatc	cagcaaacag	60
tcactgaaca	caccgaagac	cgtggtatgg	taaccgttca	cagtaatcgt	tccagtcgtc	120
tgcgggaccc	cgacgagcgt	cactgggtac	agaccagatt	cagccggaag	agaaagcgcc	180
gcaggagag	actcgaactc	cactccgctg	gtgagcagcc	ccatgttttc	aactcgaagt	240
tcaaacggca	ttgggttata	taccatcagc	tgaacttcac	acacatctcc	ttgaaccac	300
tggaaatcta	ttttcttggt	ccgtctttct	ccacagtgtt	gcagcgtaa		349

<210> 203

<211> 241

<212> DNA

<213> Homo sapien

<400> 203

tgctcctctt	gccttaccaa	cccaaagccc	actgtgaaat	atgaagtga	tgacaaaatt	60
cagttttcaa	cgcaatatag	tatagtttat	ctgattcttt	tgatctccag	gacactttta	120
acaactgcta	ccaccaccac	caacctaggg	atttaggatt	ctccacagac	cagaaattat	180
ttctcctttg	agtttcaggc	tcctctggga	ctcctgttca	tcaatgggtg	gtaaatggct	240
a						241

<210> 204

<211> 248

<212> DNA

<213> Homo sapien

```

<400> 204
tagccatttta ccacccatct gcaaaccswg acmwwcargr cywgwackya ggcgatttga      60
agtactggta atgctctgat catgttagtt acataagtgt ggtcagttta caaaaattca      120
cagaactaaa tactcaatgc tatgtgttca tgtctgtgtt tatgtgtgtg taatgtttca      180
attaagtttt tttaaaaaaa agagatgatt tccaaataag aaagccgtgt tggttaaggca      240
agaggagc                                         248

```

```

<210> 205
<211> 505
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(505)
<223> n = A,T,C or G

```

```

<400> 205
tacgctgcaa cactgtggag ccattcatac aggtccctaa ttaaggaaca agtgattatg      60
ctacctttgc acggttaggg taccgcggcc gttaaacaatg tgtcactggg caggcgggtgc      120
ctctaatact ggtgatgcta gaggtgatgt ttttggtaaa caggcgggggt aagatttgcc      180
gagttccctt tacttttttt aacctttcct tatgagcatg cctgtgttgg gttgacagtg      240
ggggtaataa tgacttggtg gttgattgta gatattgggc tgttaattgt cagttcagtg      300
ttttaatctg acgcaggctt atgcggagga gaatgttttc atgttactta tactaacatt      360
agttcttcta tagggtgata gattggtcca attgggtgtg aggagttcag ttatatgttt      420
gggatttttt aggtagtggg tgttgancct gaacgctttc ttaattgggtg gctgctttta      480
rgcctactat ggggtggtaaa tggct                                         505

```

```

<210> 206
<211> 179
<212> DNA
<213> Homo sapien

```

```

<400> 206
tagactgact catgtcccct accaaagccc atgtaaggag ctgagttcct aaagactgaa      60
gacagactat tctctggaga aaaataaaat ggaaattgta ctttaaaaaa aaaaaaatc      120
ggccgggcat ggtagcacac acctgtaatc ccagctacta ggggacatga gtcagtcta      179

```

```

<210> 207
<211> 176
<212> DNA
<213> Homo sapien

```

```

<400> 207
agactgactc atgtccccta cccaccttc tgctgtgctg cctgtttcct aacaggtcac      60
agactggtac tggtcagtgg cctgggggtt ggggacctct attatatggg atacaaattt      120
aggagttgga attgacacga ttagtgact gatgggatat gggtggtaaa tggcta          176

```

```

<210> 208
<211> 196
<212> DNA
<213> Homo sapien

```

```

<400> 208

```

1000
 900
 800
 700
 600
 500
 400
 300
 200
 100
 0


```

agactgactc atgtccccta tttaacaggg tctctagtgc tgtgaaaaaa aaaaatgctg      60
aacattgcat ataacttata ttgtaagaaa tactgtacaa tgactttatt gcatctgggt      120
agctgtaagg catgaaggat gccagaagat ttaaggaata tgggtggtaa atggctaggg      180
gacatgagtc agtcta                                     196

```

```

<210> 209
<211> 345
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(345)
<223> n = A,T,C or G

```

```

<400> 209
gacgcttggc cacttgacac cttttatattt ttaaggattc ttaagtcatt tangtnactt      60
tgtaagtatt tctgtgccc ccataagaat gatagcttta aaaattatgc tggggtagca      120
aagaagatac ttctagcttt agaatgtgta ggtatagcca ggattcttgt gaggaggggt      180
gatttagagc aaattttctta ttctccttgc ctcatctgta acatggggat aataatagaa      240
ctggcttgac aagggttgaa ttagtattac atggtaaata catgtaaaat gtttagaatg      300
gtgccaaagta tctaggaagt acttgggcat ggggtggtaa tggct                                     345

```

```

<210> 210
<211> 178
<212> DNA
<213> Homo sapien

```

```

<400> 210
gacgcttggc cacttgacac tagagtaggg tttggccaac tttttctata aaggaccaga      60
gagtaaatat ttcaggcttt gtgggttggt cagtctctct tgcaactact cagctctgcc      120
attgtagcat agaaatcagc catagacagg acagaaatga atgggtggta aatggcta       178

```

```

<210> 211
<211> 454
<212> DNA
<213> Homo sapien

```

```

<400> 211
tgggcacctt caatatctat ccagcgcac taaattcgct tttttcttga ttaaaaaattt      60
caccacttgc tgtttttgct catgtatacc aagtagcagt ggtgtgaggc catgcttggt      120
ttttgattcg atatcagcac cgtataagag cagtgccttg gccattaatt tatcttcatt      180
gtagacagca tagtgtagag tggatatccc ataactcatc ggaatatttg gatcagtgcc      240
atgttccagc aacattaacg cacattcacc ttcttggcat tgtacggcct ttgtcagagc      300
tgtcctcttt ttgttgtcaa ggacattaag ttgacatcgt ctgtccagca cgagttttac      360
tactttctgaa ttccatttgg cagaggccag atgtagagca gtccctcttt gcttgtccct      420
cttgttcaca tcagtgtccc tgagcataac ggaa                                     454

```

```

<210> 212
<211> 337
<212> DNA
<213> Homo sapien

```

```

<400> 212

```

```

tccgttatgc caccagaaaa acctactgga gttacttatt aacatcaagg ctggaaccta      60
tttgcctcag tcctatctga ttcacgagca catgggttatt actgatcgca ttgaaaacat      120
tgatcacctg gggttcttta tttatcgact gtgtcatgac aaggaaactt acaaaactgca      180
acgcagagaa actattaaag gtattcagaa acgtgaagcc agcaattgtt tcgcaattcg      240
gcattttgaa aacaaatttg ccgtggaaac ttttaatttgt tcttgaacag tcaagaaaaa      300
cattattgag gaaaattaat atcacagcat aacggaa                                337

```

```

<210> 213
<211> 715
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(715)
<223> n = A,T,C or G

```

```

<400> 213
tcgggtgatg cctcctcagg catcttccat ccatctcttc aagattagct gtcccaaagt      60
tttttccctc tcttctttac tgataaattt ggactccttc ttgacactga tgacagcttt      120
agtatccttc ttgtcacctt gcagacttta aacataaaaa tactcattgg ttttaaaagg      180
aaaaaagtat acattagcac tattaagctt ggccctgaaa ctttttctat cttttattaa      240
atgtcgggta gctgaacaga attcatttta caatgcagag tgagaaaaga agggagctat      300
atgcatttga gaatgcaagc attgtcaaat aaacatttta aatgctttct taaagtgagc      360
acatacagaa atacattaag atattagaaa gtgtttttgc ttgtgtaact ctaattaggg      420
aagcaccttg tatagttcct cttctaaaat tgaagtagat tttaaaaacc catgtaattt      480
aattgagctc tcagttcaga ttttaggaga attttaacag ggatttggtt ttgtctaaat      540
tttgtcaatt tnttttagtta atctgtataa ttttataaat gtcaaactgt atttagtccg      600
ttttcatgct gctatgaaag aaatacccan gacagggtta tttataaang gaaagangtt      660
aatttgactc ccagttcaca ggccctgagga ngnatcnccc gaaatcctta ttgcg                                715

```

```

<210> 214
<211> 345
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(345)
<223> n = A,T,C or G

```

```

<400> 214
ggtaangngc ataentcggg gctccggccg ccggagtcgg gggattcggg tgatgcctcc      60
tcaggcccac ttgggcctgc ttttcccaaa tggcagctcc tctggacatg ccattccttc      120
tcccacctgc ctgattcttc atatgttggg tgccctgtt tttctggtgc tatttctga      180
ctgctgttca gctgccactg tcctgcaaag cctgcctttt taaatgcctc accattcctt      240
catttgtttc ttaaataatg gaagtgaag tgccacctga ggccgggcac agtggtcac      300
gcctgtaatc ccagcacttt gggagcctga ggaggcatca cccga                                345

```

```

<210> 215
<211> 429
<212> DNA
<213> Homo sapien

```

<400> 215

ggtgatgcct	cctcaggcga	agctcaggga	ggacagaaac	ctcccgtgga	gcagaagggc	60
aaaagctcgc	ttgatcttga	ttttcagtac	gaatacagac	cgtgaaagcg	gggcctcacg	120
atccttctga	ccttttgggt	tttaagcagg	aggtgtcaga	aaagttacca	cagggataac	180
tggcttgtgg	cggccaagcg	ttcatagcga	cgtcgctttt	tgatccttcg	atgtcggttc	240
ttcctatcat	tgtgaagcag	aattcaccaa	gcgttggttt	gttcacccac	taatagggaa	300
cgtgagctgg	gttttagaccg	tcgtgagaca	ggtttagttt	accctactga	tgatgtgtkg	360
ttgccatggt	aatcctgtct	agtacgagag	gaaccgcagg	ttcasacatt	tgggtgtatgt	420
gcttgccctt						429

<210> 216

<211> 593

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(593)

<223> n = A,T,C or G

<400> 216

tgacacctat	gtcngcatc	tgttcacagt	ttccacaaat	agccagcctt	tggccacctc	60
tctgtcctga	ggtatacaag	tatatcagga	ggtgtatacc	ttctcttctc	ttccccacca	120
aagagaacat	gcaggctctg	gaagctgtct	taggagcctt	tgggctcaga	atttcagagt	180
cttgggtacc	ttggatgtgg	tctggaagga	gaaacatttg	ctctggataa	ggagtacagc	240
cggaggaggg	tcacagagcc	ctcagctcaa	gcccctgtgc	cttagtctaa	aagcagcttt	300
ggatgaggaa	gcaggttaag	taacatacgt	aagcgtacac	aggtagaaaag	tgctgggagt	360
cagaattgca	cagtgtgtag	gagtagtacc	tcaatcaatg	agggcaaata	aactgaaaaga	420
agaagaccna	ttaatgaatt	gcttangggg	aaggatcaag	gctatcatgg	agatctttct	480
aggaagatta	ttgtttanaa	ttatgaaagg	antagggcag	ggacagggcc	agaagtanaa	540
ganaacattg	cctatanccc	ttgtcttgca	cccagatgct	ggacaagggtg	tca	593

<210> 217

<211> 335

<212> DNA

<213> Homo sapien

<400> 217

tgacaccttg	tccagcatct	gacgtgaaga	tgagcagctc	agaggaggtg	tcctggattt	60
cctgggttctg	tgggctccgt	ggcaatgaat	tcttctgtga	agtggatgaa	gactacatcc	120
aggacaaatt	taatcttact	ggactcaatg	agcagggtccc	tcactatcga	caagctctag	180
acatgatctt	ggacctggag	cctgatgaag	aactggaaga	caaccccaac	cagagtgacc	240
tgattgagca	ggcagccgag	atgcttttatg	gattgatcca	cgcccgtctac	atccttacca	300
accgtggcat	cgcccagatg	ctggacaagg	tgtca			335

<210> 218

<211> 248

<212> DNA

<213> Homo sapien

<400> 218

taagtaactgg	tcttgaaggt	cttaggtaga	gaaaaaatgt	gaatatttaa	tcaaagacta	60
tgtatgaaat	gggactgtaa	gtacagaggg	aagggtggcc	cttatcgcca	gaagttggta	120
gatgcgtccc	cgtcatgaaa	tggtgtgtca	ctgcccgcga	tttgccgaat	tactgaaatt	180

```
ccgtagaatt agtgcaaatt ctaacgttgt tcatactaaga ttatgggtcc atgtttctag 240
tactttta 248
```

```
<210> 219
<211> 530
<212> DNA
<213> Homo sapien
```

```
<220>
<221> misc_feature
<222> (1)...(530)
<223> n = A,T,C or G
```

```
<400> 219
tgacgcttgg ccacttgaca caagtagggg ataaggacaa agacccatna ggtggcctgt 60
cagccttttg ttactgttgc ttccctgtca ccacggcccc ctctgtaggg gtgtgctgtg 120
ctctgtggac attggtgcat tttcacacat accattctct ttctgcttca cagcagtctt 180
gaggcgggag cacacaggac taccttgtca gatgangata atgatgtctg gccaaactcac 240
cccccaacct tctcactagt tatangaaga gccangccta naaccttcta tcttgncccc 300
ttgcectatg acctcatccc tgttccatgc cctattctga tttctggtga actttggagc 360
agcctggttt ntccctctca ctccagcctc tctccatacc atggtanggg ggtgctgttc 420
cacncaaang gtcaggtgtg tctggggaat cctnananct gccnggagtt tccnangcat 480
tcttaaaaac cttcttgccct aatcanatng tgtccagtgg ccaaccntcn 530
```

```
<210> 220
<211> 531
<212> DNA
<213> Homo sapien
```

```
<400> 220
tgacgcttgg ccacttgaca ctaaataagca tcttctaaag gcctgattca gagttgtgga 60
aaattctccc agtgtcaggg attgtcagga acagggtgct tctgtgtctc actttacctg 120
ctgtgtttct gctggaaaag gaggggaagag gaatggctga tttttacctc atgtctccca 180
gtttttcata ttcttcttgg atcctcttct ctgacaactg ttcccttttg gtcttcttct 240
tcttgctcag agagcagggtc tctttaaaaac tgagaaggga gaatgagcaa atgattaaag 300
aaaacacact tctgaggccc agagatcaaa tattaggtaa atactaaacc gcttgccctg 360
tgtgtgtcact tttctcctct ttccatgct ctatccctct atccccacc tattcatatg 420
gcttttatct gccaaagttat cgggcctctc atcaaccttc tcccctagcc tactggggga 480
tatccatctg ggtctgtctc tgggtgtattg gtgtcaagtg gccaaagcgtc a 531
```

```
<210> 221
<211> 530
<212> DNA
<213> Homo sapien
```

```
<400> 221
attgacgctt ggccacttga caccgcctg cctgcaatac tggggcaagg gccttcactg 60
ctttctctgc accagctgcc actgcacaca gagatcagaa atgctacca ccaagactgt 120
ttgtctctcag cctctctgag gagaaagagc agaagcctgg aagtcagaag agaagctaga 180
tcggctacgg ccttggcagc cagcttcccc acctgtggca ataaagtcgt gcatggctta 240
acaatggggg cacctcctga gaaacacatt gtaggcaat tcggcgtgtg ttcacagag 300
catatttaca caaacctcga tagtgagcc tactatccac tattgctcct acgctgcaaa 360
cctgaacagc atgggactgt actgaatact ggaagcagct ggtgatggta cttatttgtg 420
tatctaaaca cagagaaggt acagtaagaa tatggtatca taaacttaca gggaccgcca 480
```

tcctatatgc agtctgttgt gaccaaaatg tgtcaagtgg ccaagcgtca

530

<210> 222
 <211> 578
 <212> DNA
 <213> Homo sapien

 <220>
 <221> misc_feature
 <222> (1)...(578)
 <223> n = A,T,C or G

<400> 222
 tgtatcgacg tagtgggtctc cgggctacta ggccgttgtg tgctggtagt acctgggttca 60
 ctgaaaggcg catctccctc cccgcgtcgc cctgaagcag ggggaggact tcgcccagcc 120
 aaggcagttg tatgagtttt agctgcggca cttcgagacc tctgagccca cctccttcag 180
 gagccttccc cgattaagga agccagggtg aggatccctt cctccccccag acaccacgaa 240
 caaaccacca cccccctat tctggcagcc catatacatc agaacgaaac aaaaataaca 300
 aataaacnaa aaccaaaaaa aaaagagaag gggaaatgta tatgtctgtc catcctgttg 360
 ctttagcctg tcagctccta nagggcaggg accgtgtctt ccgaatggtc tgtgcagcgc 420
 cgactgctgg aagtatcgga ggaggaagca gagtcagcag aagttgaacg gtgggcccgg 480
 cggctcttgg gggctgggtg tgtacttcga gaccgcttcc gctttttgtc ttagatttac 540
 gtttgctctt tggagtggga naccactacn tcnataca 578

<210> 223
 <211> 578
 <212> DNA
 <213> Homo sapien

<400> 223
 tgtatcgacg tagtgggtctc ctcttgcaaa ggactggctg gtgaatgggt tccctgaatt 60
 atggactttac cctaaacata tcttatcatc attaccagtt gcaaaatatt agaatgtgtt 120
 gtcactgttt catttgattc ctagaagggt agtccttagat atgttaacttt aacctgtatg 180
 ctgtagtgct ttgaatgcat tttttgtttg catttttgtt tgcccaacct gtcaattata 240
 gctgcttagg tctggactgt cctggataaa gctgttaaaa tattcaccag tccagccatc 300
 ttacaageta attaagtcaa ctaaagtctt ccttgttttg ccagacttgt tatgtcaatc 360
 ctcaattttc ggggttcattt tgggtgccct aaatcttagg gtgtgacttt cttagcatcc 420
 tgtaacatcc attcccaagc aagcacaact tcacataata ctttcagaa gttcattgct 480
 gaagcctttc cttcaccag cggagcaact tgattttcta caacttccct catcagagcc 540
 acaagagtat gggatatgga gaccactacg tcgataca 578

<210> 224
 <211> 345
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(345)
 <223> n = A,T,C or G

<400> 224
 tgtatcgacg tantgggtctc ccaaggtgct gggattgcag gcatgagcca ccaactcccag 60
 gtggatcttt ttctttatac ttacttcatt aggtttctgt tattcaagaa gtgtagtggt 120

```

aaaagtcttt tcaatctaca tggttaaata atgatagcct gggaaataaa tagaaatfff 180
ttctttcatc tttaggttga ataaagaaac agaaaaata gaacatactg aaaataatct 240
aagttccaac catagaagaa ctgcagaaga aatgaagaaa gtgatgatga tttagatfff 300
gatattgatt tagaagacac aggaggagac cactacgtcg ataca 345

```

```

<210> 225
<211> 347
<212> DNA
<213> Homo sapien

```

```

<400> 225
tgtatcgacg tagtgggtctc caaactgagg tatgtgtgcc actagcacac aaagccttcc 60
aacagggacg caggcacagg cagtttaaa ggaatctgtt tctaaattaa tttccacctt 120
ctctaagtat tctttcctaa aactgatcaa ggtgtgaagc ctgtgctctt tcccaactcc 180
cctttgacaa cagccttcaa ctaacacaag aaaaggcatg tctgacactc ttcttgagtc 240
tgactctgat acgttgttct gatgtctaaa gagctccaga acaccaaagg gacaattcag 300
aatgctgggtg tataacagac tccaatggag accactacgt cgataca 347

```

```

<210> 226
<211> 281
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(281)
<223> n = A,T,C or G

```

```

<400> 226
aggngnggga ntgtatcgac gtagtgggtct cccaacagtc tgtcattcag tctgcagggtg 60
tcagtgtttt ggacaatgag gcaccattgt cacttattga ctctcagct ctaaattgctg 120
aaattaaatc ttgtcatgac aagtctggaa ttcttgatga gggttttataa agtatttttg 180
atcaatactc caacaaatca gaaagccaga aagaggatcc tttcaatatt gcagaaccac 240
gagtggattt acacacctca ggagaccact acgtcgatac a 281

```

```

<210> 227
<211> 3646
<212> DNA
<213> Homo sapien

```

```

<400> 227
gggaaacact tcctcccagc cttgtaaggg ttggagccct ctccagtata tgetgcagaa 60
ttttttctctc gggtttctcag aggattatgg agtccgcctt aaaaaaggca agctctggac 120
actctgcaaa gtagaatggc caaagtttgg agttgagtgg ccccttgaag ggtcactgaa 180
cctcacaatt gttcaagctg tgtggcgggt tggtactgaa actcccggcc tccctgatca 240
gtttccctac attgatcaat ggctgagttt ggtcaggagc accccttcog tggctccact 300
catgcaccat tcataatttt acctccaagg tcctcctgag ccagaccgtg ttttcgcctc 360
gaccctcagc cggttcggct cgccctgtac tgctctctc tgaagaagag gagagtctcc 420
ctcaccagat cccaccgcct taaaaccagc ctactccctt agggtcaccc catgtctcct 480
cggctatgtc cctgttaggc tcatcaccca ttgcctcttg gttgcaaccg tgggtgggagg 540
aagtagcccc tctactacca ctgagagagg cacaagtcct tctgggtgat gagtgtctca 600
cccccttcct gggtttatgtc ccttctttct acttctgact tgtataattg gaaaacccat 660
aatcctccct tctctgaaaa gccccaggct ttgacctcac tgatggagtc tgtactctgg 720
acacattggc ccacctggga tgactgtcaa cagctccttt tgaccctttt cacctctgaa 780

```

gagagggaaa	gtatccaaag	agaggccaaa	aagtacaacc	tcacatcaac	caataggccg	840
gaggaggaag	ctagaggaat	agtgattaga	gacccaattg	ggacctaatt	gggacccaaa	900
tttctcaagt	ggagggagaa	cttttgacga	tttccaccgg	tatctcctcg	tgggtattca	960
gggagctgct	cagaaaccta	taaacttgtc	taaggcgact	gaagtcgtcc	aggggcatga	1020
tgagtcacca	ggagtgtttt	tagagcacct	ccaggaggct	tatcagattt	acaccctttt	1080
tgacctggca	gccccgaaa	atagccatgc	tcttaatttg	gcatttgtgg	ctcaggcagc	1140
cccagatagt	aaaaggaaac	tccaaaaact	agagggattt	tgctggaatg	aataccagtc	1200
agctttttaga	gatagcctaa	aagggtttttg	acagtcaaga	ggttgaaaaa	caaaaacaag	1260
cagctcaggc	agctgaaaaa	agccactgat	aaagcatcct	ggagtatcag	agtttactgt	1320
tagatcagcc	tcatttgact	tcccctccca	catggtgttt	aaatccagct	acactacttc	1380
ctgactcaaa	ctccactatt	cctgttcatg	actgtcagga	actgttgga	actactgaaa	1440
ctggccgacc	tgatcttcaa	aatgtgcccc	taggaaaggt	ggatgccacc	atgttcacag	1500
acagtagcag	cttcctcgag	aagggactac	gaaaggccgg	tgcagctgtt	accatggaga	1560
cagatgtggt	gtgggctcag	gctttaccag	caaacacctc	agcacaaaag	gctgaattga	1620
tcgccctcac	tcaggctctc	cgatggggta	aggatattaa	cgttaacact	gacagcaggt	1680
acgcctttgc	tactgtgcat	gtacgtggag	ccatctacca	ggagcgtggg	ctactcacct	1740
cagcaggtgg	ctgtaatcca	ctgtaaagga	catcaaaagg	aaaacacggc	tggtgcccgt	1800
ggtaaccaga	aagctgattc	agcagctcaa	gatgcagtgt	gactttcagt	cacgcctcta	1860
aacttgctgc	ccacagtctc	ctttccacag	ccagatctgc	ctgacaatcc	cgcatactca	1920
acagaagaag	aaaactggcc	tcagaactca	gagccaataa	aaatcaggaa	ggttggtgga	1980
ttcttctga	ctctagaatc	ttcatacccc	gaactcttgg	gaaaacttta	atcagtcacc	2040
tacagtctac	caccatttta	ggaggagcaa	agctacctca	gctcctccgg	agccgtttta	2100
agatccccc	tcttcaaagc	ctaacagatc	aagcagctct	ccggtgcaca	acctgcgccc	2160
aggtaaattgc	caaaaaaggt	cctaaaacca	gcccaggcca	ccgtctccaa	gaaaactcac	2220
caggagaaaa	gtgggaaatt	gactttacag	aagtaaaacc	acaccgggct	gggtacaaat	2280
accttctagt	actggtagac	accttctctg	gatggactga	agcatttgct	acccaaaacg	2340
aaactgtcaa	tatggtagtt	aagtttttac	tcaatgaaat	catccctcga	catgggctgc	2400
ctgtttgcc	tagggctctga	taatggaccg	gccttcgcct	tgtctatagt	ttagtcagtc	2460
agtaaggcgt	taaacattca	atggaagctc	cattgtgcct	atcgacccca	gagctctggg	2520
caagtagaac	gcatgaactg	caccctaaaa	aacactctta	caaaattaat	cttagaaaacc	2580
ggtgtaaatt	gtgtaagtct	ccttccttta	gcctacttta	gagtaagggtg	cacccttac	2640
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ctaagagatg	cccaattggc	aaaaatatca	caaactaatt	tattacagta	cctacagtct	2760
ccccaacagg	tacaagatat	catcctgcc	cttgctcgag	gaacccatcc	caatccaatt	2820
cctgaacaga	cagggccctg	ccattcattc	ccgccagggtg	acctgttggt	tggtaaaaag	2880
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acggctctga	aggtggatgg	cattcctgcg	tggattcatc	actccgcgat	caaaaaggcc	3000
aacagagccc	aactagaaac	atgggtcccc	agggtgggtg	caggccctt	aaaactgcac	3060
ctaagttggg	tgaagccatt	agattaattc	tttttcttaa	ttttgtaaaa	caatgcatag	3120
cttctgtcaa	acttatgtat	cttaagactc	aatataaccc	ccttggtata	actgaggaat	3180
caatgatttg	attcccccaa	aaacacaagt	ggggaatgta	gtgtccaaac	tggtttttac	3240
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gataagatac	tgtggcaagc	tatatccgca	gttcccagga	attcgtccaa	ttgatcacag	3480
cccctctacc	cttcagcaac	caccacctg	atcagtcagc	agccatcagc	accgaggcaa	3540
ggccctccac	cagcaaaaag	attctgactc	actgaagact	tggatgatca	ttagtatttt	3600
tagcagtaaa	gttttttttt	ctttttcttt	ctttttttct	cgtgcc		3646

<210> 228

<211> 419

<212> DNA

<213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(419)
 <223> n = A,T,C or G

<400> 228
 taagagggta caagatctaa gcacagccgt caatgcagaa cacagaacgt agcctggtaa 60
 gtgtgttaag agtgggaatt tttggagtac agagtaaggc acctaaccct agctggggtt 120
 tggtgacggt cccagatggc ttacagaaga aagtgtcctg agatgagttt ttaagaatga 180
 ataaggatag acacaagtga ggactgactt ggcagtgggt aatgggtggg ggcaaaaaac 240
 ttcgcatgta tggaaactgc acgtacagga atgaagaatg agactgtgtg gtgtttaatg 300
 agctgcaaact actaatTTTA tcctgaaagt tttgaagagt taactaaaaa gtatTTTTTA 360
 gtaaggaaat aaccctacat ttcaggggta ttgtttgttt anatattgaa ggtgccccaa 419

<210> 229
 <211> 148
 <212> DNA
 <213> Homo sapien

<400> 229
 aagagggtag ctgtatgtag ccatgggtggc aatgagagac tgattactac ctgctggaga 60
 ttgtttaagt gagttaatat attaaggata aagggagcca ggtTTTTTga ctgttggaga 120
 aggaaattac agatattgaa ggtcccaa 148

<210> 230
 <211> 257
 <212> DNA
 <213> Homo sapien

<400> 230
 taagagggta cmaaaaaaaaa aaaatagaac gaatgagtaa gacctactat ttgatagtag 60
 aacaggggtga ctatagtcaa tgataactta attatacatt taacatagag tgtaattgga 120
 ttgtttgttaa ctcgaaggat aaatgcttga gaggatggat accccattct ccatgatgta 180
 cttatttcac attacatgcc tgtatcaaag catctcatat accctataaa tatgtacacc 240
 tactatgtac cctctta 257

<210> 231
 <211> 260
 <212> DNA
 <213> Homo sapien

<400> 231
 taagagggta cgggtatttg ctgatgggat ttttttttct ttctttttct ttggaaaaca 60
 aaatgaaagc cagaacaaaa ttattgaaca aaagacaggg actaaatctg gagaaatgaa 120
 gtcccctcac ctgactgcca tttcattcta tctgaccttc cagtctaggt taggagaata 180
 gggggtggag gggattaatc tgatacaggt atattttaaag caactctgca tgtgtgccag 240
 aagtccatgg taccctctta 260

<210> 232
 <211> 596
 <212> DNA
 <213> Homo sapien

<220>

<221> misc_feature
 <222> (1)...(596)
 <223> n = A,T,C or G

<400> 232
 tgctcctctt gccttaccaa ccacaaatta gaaccataat gagatgtcac ctcatacctg 60
 gtgggattaa cattatttaa aaaatcagaa gtattgacaa ggatgtgaag aaattagaac 120
 atctgtgcac tgttggtggg aatgtaaaaa aggtgtggcc actatgggta acagcatgaa 180
 ggttcctcaa aaaaaatfff ttttaatcta ctctatgac gatcttgagg ttgtttatgc 240
 aaaagaactg aaatcaggat tttgaggaaa tattcacatt cccacatcca tttctgcttt 300
 attcataata ctcaagagat ggaaacaacc taaatgtcca tcccgggatg aatggataaa 360
 cacagtgtgg tatatgcata caatggaata ttatttagtc tttaaaaaga aaaattctat 420
 catatactac aacttanatn aaccttgagg acacaatgct nagtgaaata agccacggaa 480
 ggacgaatac tgcattattc ccttatatga agtatctaaa gtggtcaaac tcttanagca 540
 naaagtaaaa atgggtgggt gccanacagt tggtaggcn agaaganaan cctant 596

<210> 233
 <211> 96
 <212> DNA
 <213> Homo sapien

<400> 233
 tcttctgaag accttttcgag actcttaagc tcgtgggttg taaggcaaga ggagcgttgg 60
 taaggcaaga ggagcgttgg taaggcaaga ggagca 96

<210> 234
 <211> 313
 <212> DNA
 <213> Homo sapien

<400> 234
 tgtaagtcga gcagtgtgat gataaaactt gaatggatca atagttgctt cttatggatg 60
 agcaaagaaa gtagtttctt gtgatggaat ctgctcctgg caaaaatgct gtgaacgttg 120
 ttgaaaagac aacaaagagt ttagagtagt acataaatff agaatagtag ataaacttag 180
 aatagtagat aaacttagta cataaataat gcacgaagca ggggcagggc ttgagagaat 240
 tgacttcaat ttggaaagag tatctactgt aggttagatg ctctcaaaca gcatcacact 300
 gctcgactta caa 313

<210> 235
 <211> 550
 <212> DNA
 <213> Homo sapien

<400> 235
 aacgaggaca gatccttaaa aagaatgttg agtgaaaaaa gtagaaaata agataatctc 60
 caaagtccag tagcattatt taaacatfff taaaaaatat actgataaaa attttgtaca 120
 tttcccaaaa atacatatgg aagcacagca gcatgaatgc ctatgggrtt gaggatagg 180
 gttgggagta gggatgggga taaaggggga aaataaaacc agagaggagt cttacacatt 240
 tcatgaacca aggagtataa ttatttcaac tatttgtaac wgaagtccag aaagagtggga 300
 ggcagaaggg ggagaagagg gcgaagaaac gtttttgagg gaggggtccc asaagagaga 360
 ttttcgcat gtggcgctac atacgttttt ccaggatgcc ttaagctctg caccctatff 420
 ttctcatcac taatattaga ttaaaccctt tgaagacagc gtctgtgggt tctctacttc 480
 agctttccct ccgtgtcttg cacacagtag ctgttttaca agggttgaac tgactgaagt 540
 gagattatfc 550

<210> 236
 <211> 325
 <212> DNA
 <213> Homo sapien

<400> 236
 tagactgact catgtcccct accagagtag ctagaattaa tagcacaagc ctctacaccc 60
 aggaactcac tattgaatac ataaatggaa tttattcagc cttaaaaagt ttggaaggaa 120
 attctgacat atgctaaaaac atggatgaac cttgaagact ttatgataag taaaagaagc 180
 cagtcataaa aggaaaaata ttgcatgatt ccacttatat gaggtaccta gagtagtcaa 240
 tttcatagaa acacaaaata gaatggtgtt tgccagggtt tttgaggaaa agggaatgac 300
 aagttagggg acatgagtca gtcta 325

<210> 237
 <211> 373
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(373)
 <223> n = A,T,C or G

<400> 237
 tagactgact catgtcccct atctactcaa catttccact tgaagtctga taggcatctc 60
 agacttatct tgtcccaaag caaactcttt atttcttttc atcctagtct ttatttcttg 120
 tgctgtctta cccatctcaa aagagtgcc aatccacca agttgctgaa acagaaatct 180
 aagaaatctc cttgattctt ctttttccca tctacttcac ttctaattca ttagtaaata 240
 atctgtttca gaaaaccaa cacctcatgt tctactcat aagggggagt tgaacaatga 300
 gaacacacag acacaggag ggaacatca cacaccacgg ccgctcagg agtangggac 360
 atgagtcagt cta 373

<210> 238
 <211> 492
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(492)
 <223> n = A,T,C or G

<400> 238
 tagactgact catgtcccct ataatgctcc caggcatcag aaagcatctc aaactggagc 60
 tgacaccatg gcagagggtt caggtaagtc acaaaagggg tcctaaagaa tttgccctca 120
 atatcagagt gattagaaga agtgacaga gctacccaag ttaaacatat gcgagataaa 180
 aaaaatatgg cacttgatgaa cacacactac aggaggaaaa taaggaacat aatagcatat 240
 tgtgctatta tgatgatgaa gaacctctct anaagaaaac ataaccaaag aaacaaagaa 300
 aattcctgcn aatgtttaat gctatagaag aaattaacaa aaacatatat tcaatgaatt 360
 cagaaaagtt agcagggtcan aagaaaacaa atcaaagacc agaataatcc catttttagat 420
 tgtcgagtaa actanaacag aaagaatacc actggaaatt gaattcctac gtangggaca 480
 tgantcanc ta 492

<210> 239
 <211> 482
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(482)
 <223> n = A,T,C or G

<400> 239
 tggaaagtat ttaatgatgg gcaacttgct gtttacttcc tacatatccc atcatcttct 60
 gtatTTTTTT aaataacttt tttttggatt tttaaagtaa cttattctg agaggtaaca 120
 tggattacat acttctaagc cattaggaga ctctatgtta aacccaaaagg aaatgttact 180
 agatcttcat ttgatcaata ggatgtgata atcatcatct ttctgctcta atggaaaagt 240
 actanaaaca tggaaccata atcttagatg aacaacgtta gaatttgac taattctacg 300
 gaatttcagt aattcggcaa atgtcgggca gtgacacaac atttcatgac ggggacgcat 360
 ctaccaactt ctggcgataa gggccaccct tccctctgta cttacagtcc catttcatac 420
 acagtctttg attaaatatt cacatTTTTT ctctacctaa agaccttcaa gaccagtacg 480
 ta 482

<210> 240
 <211> 519
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(519)
 <223> n = A,T,C or G

<400> 240
 tgtatcgacg tagtgggtctc cccatgtgat agtctgaaat atagcctcat gggatgagag 60
 gctgtgcccc agcccgacac ccgtaaaggg tctgtgctga ggtggattag taaaagagga 120
 aagccttgca gttgagatag aggaagggca ctgtctcctg cctgcccctg ggaactgaat 180
 gtctcgggtat aaaacccgat tgtacatttg ttcaattctg agataggaga aaaaccaccc 240
 tatggcgagg ggcgagacat gttggcagca atgctgcctt gttatgcttt actccacaga 300
 tgtttgggcg gagggaaaca taaatctggc ctacgtgcac atccaggcat agtacctccc 360
 tttgaactta attatgacac agattccttt gctcacatgt ttttttgctg accttctcct 420
 tattatcacc ctgctctcct accgcattcc ttgtgctgag ataatgaaaa taatatcaat 480
 aaaaacttga nggaactcgg agaccactac gtcgatata 519

<210> 241
 <211> 771
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(771)
 <223> n = A,T,C or G

<400> 241
 tgtatcgacg tagtgggtctc cactcccgcc ttgacggggc tgctatctgc cttccaggcc 60

actgtcacgg	ctccccgggta	gaagtcactt	atgagacaca	ccagtgtggc	cttgttggct	120
tgaagctcct	cagaggaggg	tgggaacaga	gtgaccgagg	gggcagcctt	gggctgacct	180
aggacggtea	gcttgggtccc	tccgccaaac	acgagagtgc	tgctgcttgt	atatgagctg	240
cagtaataat	cagcctcgtc	ctcagcctgg	agcccagaga	tggtcagggg	ggccgtgttg	300
ccanacttgg	agccagagaa	gcgattagaa	acccctgagg	gccgattaac	gacctcataa	360
atcatgaatt	tgggggcttt	gcctgggtgc	tgttggtagc	angagacatt	attataacca	420
ccaacgtcac	tgctgggtcc	antgcaggga	aaatggttga	tcnaactgtc	caagaaaacc	480
actacgtcca	taccaatcca	ctaattgccc	gccgcctgca	ggttcaacca	tattggggaa	540
naactccccc	ccgcgcgttg	ggattgncat	naacctttga	aattttttcc	tattanttgt	600
ccccctaaaa	taaacnnttg	ggccttaatc	cattgggtcc	atancttntt	tncccggttt	660
ttaaaanttg	tttatcccg	cncnctttt	cccccccaac	tttccaaaac	ccgaaacctt	720
tnaaatttnt	tnaaaccttg	gggggttccc	nnaattnnan	ttnaancctc	c	771

<210> 242

<211> 167

<212> DNA

<213> Homo sapien

<400> 242

tgggcacctt	caatatcggg	ctcatcgata	acatcacgct	gctgatgctg	ctgttgcctg	60
tcctctctag	gaacctctgg	atcttcaa	ttcttgagga	attcatccaa	attatctgcc	120
tctctctctt	tctctctttt	tctaaggtct	tctggtacaa	gcggtca		167

<210> 243

<211> 338

<212> DNA

<213> Homo sapien

<400> 243

ttgggcacct	tcaatatcta	ctgatctaaa	tagtggtggt	tgaggcctct	tgttcctggc	60
taaaaatcct	tggcaagagt	caatctccac	tttacaatag	aggtaaaaaat	cttacaatgg	120
atattcttga	caaagctagc	atagagacag	caattttaca	caaggatttt	ttcacctggt	180
taataacagt	ggttttctta	cacccatagg	gtgccaccaa	gggaggagtg	cacagttgca	240
gaaacaaaatt	aagatactga	agacaacact	aettaccatt	tcccgtatag	ctaaccacca	300
gttcaactgt	acatgtatgt	tcttatgggc	aatcaaga			338

<210> 244

<211> 346

<212> DNA

<213> Homo sapien

<400> 244

tttttggctc	ccatacagca	cactctcatg	ggaaatgtct	gttctaaggt	caaccataa	60
tgcaaaaaatc	atcaatatac	ttgaagatcc	ccgtgtaagg	tacaatgtat	ttaatattat	120
cactgataca	attgatccaa	taccagtttt	agtctggcat	tgaatcaaat	cactgttttt	180
gttgataaaa	aagagaaata	tttagcttat	atttaagtac	catattgtaa	gaaaaaagat	240
gcttatcttt	acatgctaaa	atcatgatct	gtacattggt	gcagtgaata	ttactgtaaa	300
agggagaag	gaatgaagac	gagctaagga	tattgaaggt	gcccaa		346

<210> 245

<211> 521

<212> DNA

<213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(521)
 <223> n = A,T,C or G

<400> 245
 accaatccca cacggatact gagggacaag tatatcatcc catttcatcc ctacagcagc 60
 aacttcatga ggcaggagtt attagtccca ttttacagaa gaggaaactg agacttaggg 120
 agatcaagta atttgcccag gtcgcacaat tagtgataga gccagggtt gaagcgacgt 180
 ctgtcttaag ccaatgaccc ctgcagatta ttagagcaac tgttctccac aacagtgtaa 240
 gcctcttget anaagctcag gtccacaagg gcagagattt ttgtctgttt tgctcattgc 300
 tccttcccca ttgcttagag cagggtctgc cacgaancag gttctcaatg catagttatt 360
 aaatgtatat aagagcaaac atatgttaca gagaactttc tgtatgcttg tcacttacat 420
 gaatcacctg tganatgggt atgcttggtc cccantgttg cagatnaaga tattgaangt 480
 gcccaaataca ctanttgagg gcgcctgcan gtccancata t 521

<210> 246
 <211> 482
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(482)
 <223> n = A,T,C or G

<400> 246
 tggaaccaat ccaaatatccc atcaatgata gactggataa agaaaatttg gcacatgttc 60
 accatgaaat actatgcagc cataaaaaag gatgagttca tatcctttgc agggacatgg 120
 atgaagctgg agaccatcat tctcagcaaa ctaacaaggg aacagaaaac caaacactgc 180
 atgttctcac tcttaagtgg gagctgaaca atgagaacac atggacacag ggaggggaac 240
 atcacacagt ggggcctgct ggtgggtagg ggtctagggg agggatagca ttaggagaaa 300
 tacctaattg agatgacggg ttgatgggtg cagcaaacca ccatgacacg tgtataccta 360
 tgtaacaaac ctgcatgttc tgcacatgta cccagaact taaagtgtta ataaaaaaat 420
 taagaaaaaa gttaagtatg tcatagatac ataaaatatt gtanatattg aaggtgccca 480
 aa 482

<210> 247
 <211> 474
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(474)
 <223> n = A,T,C or G

<400> 247
 ttcgatacag gcacagagta agcagaaaaa tggtctgtgtt ttaaccaagt gagtacagtt 60
 aagtgaagaga ggggcagaga agacaagggc atatgcaggg ggtgattata acaggtgggt 120
 gtgctgggaa gtgagggtac tcggggatga ggaacagtga aaaagtggca aaaagtggta 180
 agatcagtga attgtacttc tccagaattt gatttctggn ggagtcaa atactatccag 240
 tttggggat catanggcaa cagttgaggt ataggaggta gaagtcncag tgggataatt 300
 gaggttatga anggtttggt actgactggt actgacaang tctgggttat gaccatggga 360

```

atgaatgact gtanaagcgt anaggatgaa actattccac ganaaagggg tccnaaaact 420
aaaaannnaa gnnnnngggg aatattattt atgtggatat tgaangtgcc caaa 474

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<210> 248
<211> 355
<212> DNA
<213> Homo sapien

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<220>
<221> misc_feature
<222> (1)...(355)
<223> n = A,T,C or G

```

```

<400> 248
ttcgatacag gcaaacatga actgcaggag ggtggtgacg atcatgatgt tgccgatggt 60
ccgatggnc acgaagacgc actgganac gtgcttacgt ccttttgctc tgttgatggc 120
cctgagggga cgcaggaccc ttatgaccct cagaatcttc acaacgggag atggcactgg 180
attgantccc antgacacca gagacacccc aaccaccagn atatcantat attgatgtag 240
ttcctgtaga nggccccctt gtggaggaaa gctccatnag ttggtcatct tcaacaggat 300
ctcaacagtt tccgatggct gtgatgggca tagtcatant taaccntgtn tcgaa 355

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<210> 249
<211> 434
<212> DNA
<213> Homo sapien

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<400> 249
ttggattggt cctccaggag aacaagggga aaaagggtgac cgagggtccc ctggaactca 60
aggatctcca ggagcaaaaag gggatggggg aattcctggt cctgctggct ccttaggtcc 120
acctggctct ccaggcttac caggctcctca aggcccaaag ggtaacaaaag gctctactgg 180
acctgctggc cagaaaagggt acagtgggtct tccagggcct cctgggcctc caggtccacc 240
tggatgaagtc attcagcctt taccaatctt gtctctccaa aaaacgagaa gacatactga 300
aggcatgcaa gcagatgcag atgataatat tcttgattac tcggatggaa tgggaagaaat 360
atcttggttc ctcaattccc tgaaacaaga catcgagcat atgaaatttc caatgggtac 420
tcagaccaat ccaa 434

```

```

<210> 250
<211> 430
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(430)
<223> n = A,T,C or G

```

```

<400> 250
tggattggct acatggcaga gacaggattc caaggcagtg agaggaggat acaatgcttc 60
tcactagtta ttattattta ttttattttt gagatgaagt ctgctttgt ctcccaggct 120
ggagagcggg ggtgcgatct tggctctctg caacccccgc ctcaagcaat tctcctgtct 180
tagcctcgcg ggtagatgga attacaggcg cccaccgccca tgcccaacta atttttttgt 240
gtcttcagta gagacagggt ttcgccatgt tgggcaggct ggtcttgaac tctgacctc 300
nagtgatctg cctcctcgg cctcaciaag tgctggaatt acaggcatgg gctgctgcac 360
ccagtcaact tctcactagt tatggcctta tcattttcac cacattctat tggcccaaaa 420

```

aaaaaaaaan

430

<210> 251
 <211> 329
 <212> DNA
 <213> Homo sapien

<400> 251
 tgggtactcca ccatyatggg gtcaaccgcc atcctcgcgc tctcctcggc tgttctccaa 60
 ggagtctgtg ccgaggtgca gctgrtgag tctggagcag aggtgaaaaa gtccggggag 120
 tctctgaaga tctcctgtaa gggttctgga tacaccttta agatctactg gatcgctgg 180
 gtgcgccagt tgcccgggaa aggcctggag tggatggggc tcctctttcc tgatgactct 240
 gataccagat acagcccgtc cttccaaggc caggtcacca tctcagtcga taagtccatc 300
 agcaccgcct atctgcagtg gactaccaa 329

<210> 252
 <211> 536
 <212> DNA
 <213> Homo sapien

<400> 252
 tgggtactcca ctcagcccaa ccttaattaa gaattaagag ggaacctatt actattctcc 60
 caggctcctc tgctctaacc aggcctctgg gacagtatta gaaaaggatg tctcaacaag 120
 tatgtagatc ctgtactggc ctaagaagtt aaactgagaa tagcataaat cagaccaaac 180
 ttaatggtcg ttgagacttg tgctcctggag cagctgggat aggaaaactt ttgggcagca 240
 agaggaagaa ctgcctggaa gggggcatca tgtaaaaaat tacaagggga acccacacca 300
 ggcccccttc ccagctctca gcctagagta ttagcatttc tcagctagag actcacaact 360
 tccttgctta gaatgtgcca ccggggggag tcctgtgtgg tgatgaggct ctcaagagt 420
 agagtggcat cctatcttct gtgtgcccac aggagcctgg cccgagactt agcaggtgaa 480
 gtttctggtc caggctttgc ccttgactca ctatgtgacc tctggtggag taccaa 536

<210> 253
 <211> 507
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(507)
 <223> n = A,T,C or G

<400> 253
 ntgttgcat cccagtaact cgggaagctg aggcgggagg atcacctgag ctcaggaggt 60
 tgaggccgca gtgagccggg accacgccac tacactccag cctggggcat agagtgagac 120
 cctccaagac agaaaagaaa agaaaggaag ggaaagggaa agggaaaagg aaaaggaaaa 180
 ggaaaaggaa aaggaaaaga caagacaaaa caagacttga atttggatct cctgacttca 240
 attttatggt ctttctacac cacaattcct ctgcttacta agatgataat ttagaaaccc 300
 ctogttccat tctttacagc aagctggaag tttggtcaag taattacaat aatagtaaca 360
 aatttgaata ttatatgcca ggtgttttct attcctgctc tcaacttaatt ctaccactc 420
 tgatataaat acaattgctg ccgggtgtgg tggctcatgc ctgtaatccc ggcactttgg 480
 gagaccgagg tgggcgggats gcaacaa 507

<210> 254
 <211> 222

<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(222)
<223> n = A,T,C or G

<400> 254
ttggattggt cactgtgagg aagccaaatc ggatccgaga gtctttttct aaaggccagt 60
actggccaca cttttctcctg ccgccttcct caaagctgaa gacacacaga gcaaggcgct 120
tctgttttac tccccaatgg taactccaaa ccatagatgg ttagctnccc tgctcatctt 180
tccacatccc tgctattcag tatagtcctg ggaccaatcc aa 222

<210> 255
<211> 463
<212> DNA
<213> Homo sapien

<400> 255
tggtgcgatc cataaatgct gaaatggaaa taaacaacat gatgagggag gattaagttg 60
gggaggggagc acattaaggt ggccatgaag tttgttggaa gaagtgactt ttgaacaagg 120
ccttggtggt aagagctgat gagagtgtcc cagacagagg ggccactggg acaatagacg 180
agatgggaga gggcttggaa ggtgtgcgaa ataggaagga gtttgttctg gtatgagtct 240
agtgaacaca gaggcgagag gccctgggtg gtgcagctgg agagtattgc agaataacat 300
taggccctgt gggggactgt agactgtcag caataatcca cagtttggat tttattctaa 360
gagtgatggg aagccgtgga aaggggggta agcaaggagt gaaattatca gatttacagt 420
gataaaaata aattgggtctg gctactgggg aaaaaaaaaa aaa 463

<210> 256
<211> 262
<212> DNA
<213> Homo sapien

<400> 256
ttggattggt caacctgctc aactctacyt ttcctccttc ttcttaaaaa attaatgaat 60
ccaatacatt aatgccaaaa cccttgggtt ttatcaatat ttctgttaaa aagtattatc 120
cagaactgga cataatacta cataataata cataacaacc ccttcatctg gatgcaaaca 180
tctattaata tagcttaaga tcactttcac tttacagaag caacatcctg ttgatgttat 240
tttgatgttt ggaccaatcc aa 262

<210> 257
<211> 461
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(461)
<223> n = A,T,C or G

<400> 257
gnggnnnnnn nnncaattcg actcngttcc cntggtancc ggtcgacatg gccgcgggat 60
taccgcttgt nnetgggggt gtatggggga ctatgaccgc ttgtagctgg ggggtgatgg 120


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<210> 258
<211> 332
<212> DNA
<213> Homo sapien
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[illegible]

<400>	259						
ctttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	60
ctttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	gaccgcttgt	120
ctttgt	gaccgcttgt	nacngggggg	gtctggggga	ctatgannga	ntgtactgg		180
ctctgg	gggncatga	nngantgtna	cnggggtgt	ctgggggact	atganngact		240
ncctg	ggggatcnga	ggagantngn	ggntagngat	ggttngggan	a		291

[illegible]

<211> 746
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(746)
 <223> n = A,T,C or G

<400> 261

ttgggcacct	tcaatatcaa	tagctaacat	ttattgagtg	tttatcgat	cataaaacac	60
tggttctaagc	ctttaaacgt	actaattcat	ttaatgctca	taatcacttt	agaagggtggg	120
tactagtatt	agtctcattt	acagatgcaa	catgcaggca	cagagagggt	aattaacttg	180
cccaaggtaa	cacagctaag	aaatagaaaa	aatattgaat	ctggaaagtt	gggtttcttg	240
gtaaccacac	gagttctcaa	tgagcctggg	gcctcactca	gtttgctttt	acaaagcgaa	300
tgagtaacat	cacttaattc	agtgagtagg	ccaaatggag	gtcagctacg	agttttctgct	360
gttcttgcag	tggactgaca	gatgtttaca	acgtctggcc	atcagtwaat	ggactgatta	420
tcattgggaw	gtgggtgggc	tgaatgttgg	ccagtgaagt	ttattcawgc	catattttta	480
tgtttaggat	gacttttggc	tggtcctagg	gcaagctctg	tctgscacgg	aacacagaat	540
wacacaggga	ccccctcaat	ttctgggtgtg	gctagaacca	tgaaccactg	gttgggggaa	600
caagcgggtca	aaacctcaat	gcggccggct	ggcagggtcc	acccatatgg	ggaaaactcc	660
cnacgcgttt	ggaatgcctn	agctngaatt	attctaanag	ttgtccnct	aaaattagcc	720
tgggcggttaa	tcangggctn	naagcc				746

<210> 262
 <211> 588
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(588)
 <223> n = A,T,C or G

<400> 262

tgaccgcttg	tcatttcaca	tgggggtcctg	cacgcttttg	cctttgtagg	aaacctgaca	60
tttgtctgtt	tcttctttct	cttttccttc	ccatatactc	ctaatttacg	tttgacttgt	120
ttgctgagga	ggcaggagct	agagactgct	gtgagctcat	aggggtggga	agtttatcct	180
tcaagtcccg	cccactcatc	actgcttctc	accttccct	gaccaggctt	acaagtgggt	240
tcttgcttgc	tttccctttg	gacccaacaa	gcccctgtaa	tgagtgtgca	tgactctgac	300
agctgtggac	tcagggtcct	tggctacagc	tgccatgtaa	aatatctcat	ccagttctcg	360
caaattgtta	aaataaccac	atttcttaga	ttccagtaac	caaatacatg	ctttacgaac	420
tgctcctcac	acccagaagt	ggcacaaata	ttcttgggga	attattactt	ttttttttct	480
ctctntttnn	gnnnngnnng	gnnnngnccag	gaattaccac	nttggaagac	ctggccngaa	540
tttattatan	aggggagccg	attntttttc	ctaacacaaa	gcgggtca		588

<210> 263
 <211> 730
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(730)

<223> n = A,T,C or G

<400> 263

tttttttttt	tttggcctga	gcaactgaaa	ttatgaaatt	tccatatact	caaaagagta	60
agactgcaaa	aagattaaat	gtaaaagttg	tcttgtatac	agtaatgttt	aagataccta	120
ttanatttat	aaatggaaaa	ttagggcatt	tgatataca	agttgaaaat	tcaggagtga	180
ggttgggctg	gctgggtata	tactgaaaac	tgtcagtaca	cagatgacat	ctaaaaccac	240
aaatctgggt	ttatttttagc	agtgatatgt	gtcactccca	caaaagcctt	cccaattggc	300
ctcagcatac	acaacaagtc	acctccccac	agccctctac	acataaaca	attccttagt	360
ttagttcagg	aggaaatgcg	cccttttctt	tccgctctag	gtgaccgcaa	ggcccagttc	420
tcgtcaccaa	gatgttaagg	gaagtctgcc	aaagaggcat	ctgaaaggaa	ataaggggaa	480
tgggagtga	cacaaaggaa	agccaaggan	aaactttgga	gaccgtttct	agancctgg	540
catttcacaa	caaaactcng	gaacaaacct	tgtctcatca	atcatttaag	cccttcgttt	600
ggannagact	ttctgaactg	ggcgtgaac	ataancctca	ttgaatgtct	tcacagtctc	660
ccagctgaag	gcacaccttg	ggccagaagg	ggaatcttcc	aggtcctcaa	nacagggctc	720
gccctttgnc						730

<210> 264

<211> 715

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(715)

<223> n = A,T,C or G

<400> 264

tttttttttt	tttggccagt	atgatagtct	ctaccactat	attgaagctc	ttaggtcatt	60
tacacttaat	gtggttatag	atgctgttga	gcttacttct	accaccttgc	tatttctccc	120
gtctcttttt	tgttcttttt	ctcttctttt	cctcccttat	tttataattg	aatttttttag	180
gattctattt	tatatagatt	tatcagctat	aacactttgt	attcttttgt	tttgtggttc	240
ttctgtcatt	tcaatgtgca	tcttaaaact	atcacaaact	atcttcaaat	aatatcatat	300
aacctttacat	ataatgtaag	aatctaccac	catatatctt	catttctccc	ttccatccta	360
tgtntgtcat	atcttttctt	ttatatatgt	tttaaagaca	taatagtata	tgaggaggtt	420
ttgcttaaaa	tgtgatcaat	attccttcaa	ngaaacgtaa	aaattcaaaa	taaatntctg	480
tttattctca	aatnnacct	atatttctta	ccatntctna	taentttcaa	gaatctgaag	540
gcattgggtt	tttccggctt	aagaacctcc	tctaaagcac	tctaagcaga	attaagtctt	600
ctgggagagg	aattctccca	agcttgggcc	ttnanntgta	ctcentnang	gttaaanttt	660
ggccgggaaa	tagaaattcc	aagttaacag	gntanttttt	ntttntnttn	tcncc	715

<210> 265

<211> 152

<212> DNA

<213> Homo sapien

<400> 265

tttttttttt	tttcccaaca	caaagcacca	ttatctttcc	tcacaatttt	caacatagtt	60
tgattcccat	gaagagggtta	tgatttctaa	agaaaacatg	gctactatac	tatcaatcag	120
ggttaaatct	tttttttttg	agacggagtt	ta			152

<210> 266

<211> 193

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(193)

<223> n = A,T,C or G

<400> 266

taaactccgt	ccccttctta	atcaatatgg	aggctaccca	ctccacatta	ccttcttttc	60
aagggactgt	ttccgtaact	gttgtgggta	ttcacgacca	ggcttctaaa	cctcttaaaa	120
ctccccaatt	ctggtgccaa	cttggaacaac	atgctttttt	tttttttttt	tttttttttt	180
gagacggagt	tta					193

<210> 267

<211> 460

<212> DNA

<213> Homo sapien

<400> 267

tgttgcgatc	ccttaagcat	gggtgctatt	aaaaaaatgg	tggagaagaa	aatacctgga	60
atttacgtct	tatctttaga	gattgggaag	accctgatgg	aggacgtgga	gaacagcttc	120
ttcttgaatg	tcaattccca	agtaacaaca	gtgtgtcagg	cacttgctaa	ggatcctaaa	180
ttgcagcaag	gctacaatgc	tatgggatcc	tcccaggagg	gccaatttct	gagggcagtg	240
gctcagagat	gcccttcacc	tcccatgatc	aatctgatct	cggttggggg	acaacatcaa	300
ggtgtttttg	gactccctcg	atgcccagga	gagagctctc	acatctgtga	cttcacccga	360
aaaacactga	atgctggggc	gtactccaaa	gttgttcagg	aacgcctcgt	gcaagccgaa	420
tactggcatg	accataaaaa	ggaggatgtg	gatcgcaaca			460

<210> 268

<211> 533

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(533)

<223> n = A,T,C or G

<400> 268

tgttgcgatc	cgttgataga	atagcgacgt	ggtaatgagt	gcatggcacg	cctccgactt	60
accttcgccc	gtggggaccc	cgagtagctc	tacggcgctc	tcaacttagag	tacctcttgg	120
acgcccgggc	gcgttcgatt	taccggaagc	gcgagctgca	gtgggcttgc	gcccccgccc	180
aaattctttg	gggggtttta	ggccgcgggg	aatttgaggt	atctctatca	gtatgtagcc	240
aagttggaac	agtcgccatt	cccgaaatcg	ctttctttga	atccgcaccg	cctccagcat	300
tgcctcattc	atcaacctga	aggcacgcgt	aagtgcgggt	tgtgtcttca	gcagctccac	360
tccataacta	gcgcgctcga	cctcgtcttc	gtacgcgccca	ggtccgtgog	tgcgaaattcc	420
caactccggt	gagttgcgca	tttcaagttt	cgaaactggt	cgccctccacn	atttggcatg	480
ttcacgcatg	acacggaata	aactcgteca	gtaccgggaa	tgggatcgca	aca	533

<210> 269

<211> 50

<212> DNA

<213> Homo sapien

```
<210> 270
<211> 519
<212> DNA
<213> Homo sapien
```

```
<210> 271
<211> 457
<212> DNA
<213> Homo sapien
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```
<210> 272
<211> 102
<212> DNA
<213> Homo sapien
```

```
<210> 273
<211> 455
<212> DNA
<213> Homo sapien
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```
<220>  
<221> misc_feature  
<222> (1)...(455)  
<223> n = A,T,C or G
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tttttttttt ttggcaatca acaggtttta gtcttcggcc gaagttaatc tcgtgttttt 60
ggcaatcaac aggttttaagt cttcggccga agttaatctc gtgttttttg caatcaacag 120
gtttaagtct tcggccgaag ttaatctcgt gtttttggca atcaacagggt ttaagtcttc 180
ggccgaagtt aatctcgtgt ttttggaat caacagggtt aagtcttcgg ccgaagttaa 240
tctcgtgttt ttggcaatca acaggtttta gtcttcggcc gaagttaatc tcgtgttttt 300
ggcaatcaag aggttttaagt cttcggccga agttaatctc gtgttttttg caatcaacag 360
gtttaagtct tcggccgaan ttaatctcgt gtttttggca atcaacagggt ttaantcttc 420
ggccgaagtt aatctcgtgt ttttggaat caana 455

```

```

<210> 274
<211> 461
<212> DNA
<213> Homo sapien

```

```

<400> 274
tttttttttt ttggccaata cccttgatga acatcaatgt gaaaatcctc ggtaaaatac 60
tggcaaacca aatccagcag cacatcaaaa agcttatcca ccatgatcaa gtgggcttca 120
tccctgggat gcaaggctgg ttcaacataa gaaaatcaat aaatgtaatc catcacataa 180
acagaacca agacaaaaac cacatgatta tctcaataga tgcagaaaag gccttgaca 240
aattcaacag cccttcatgc taaacactct taataaacta gatattgatg gaatgtatct 300
caaaataata agagctatct atgacaaacc cacagccaat atcatactga atgggcaaag 360
actggaagca ttccctttga aaactggcac aagacaagga tgccctctct caccgctcct 420
attcaacata gtattggaag ttctggccag ggcaatcaag a 461

```

```

<210> 275
<211> 729
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(729)
<223> n = A,T,C or G

```

```

<400> 275
tttttttttt ttggccaaca ccaagtcttc cacgtgggag gttttattat gttttacaac 60
catgaaaaca taggaagggt gctgttacag caaacatttc agatagacga atcggccaag 120
ctccccaaac cccaccttca cagcctcttc cacacgtctc ccanagattg ttgtccttca 180
cttgcaaatt canggatgtt ggaagtngac atttnnagtn gcnggaaccc catcagtga 240
ncantaagca gaantacgat gactttgana nacanctgat gaagaacacn ctacnganaa 300
ccctttctnt cgtgttanga tctcnngtcc ntcactaatg cggcccccctg cnggtccacc 360
at ttggggaga actccccccn cg ttggatcc ccccttgagt ntccattct ngtecccan 420
accngncttg ngngncantn cncctcnca cntgtttcc ctgngtnaa aatnngtttt 480
nccgcncccc naattccac cnaatcaca gcgaancng aaggccttcn naagtgttta 540
angccngng gtttctctnt ntantgag cctaccctcc cncctnnnt tncngttgg 600
tcgcgccctg gncncgctn gttcctcttt nnggnacaa cctngntcnn nggcnctcn 660
nnctnttcc tnnnactagc tngcctntcc ncnccnggn ncanngcaca ttncncnnac 720
tntgtnncc 729

```

```

<210> 276
<211> 339
<212> DNA
<213> Homo sapien

```

<400> 276

tgacctgaca	tgtagtagat	acttaataaa	tatttggtga	atgaatggat	gaagtggagt	60
tacagagaaa	aatagaaaag	tacaaattgt	tgtcagtgtt	ttgaaggaaa	attatgatct	120
ttcccaaagt	tctgacttca	ttotaagaca	gggttagtat	ctccatacat	aattttactt	180
gcttttgaaa	atcaaagag	ataatctatt	tagattgata	atttatttag	actggctata	240
aactattaag	tgctagcaaa	tatacatttt	aatctcattt	tccacctctt	gtgatatagc	300
tatgtagggtg	ttgactttta	tggtatgtcag	gtcaatccc			339

<210> 277

<211> 664

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(664)

<223> n = A,T,C or G

<400> 277

tgacctgaca	tccataacaa	aatctttctc	catttatattc	ttctagggga	atttcttgaa	60
aagcatccaa	aggaaacaaa	tgatggtaag	accgtgccaa	gtggggagca	gacaccaaag	120
taagaccaca	gattttacat	tcaacaggta	gtcacagta	ctttgccga	cactgtgggc	180
agaaatagcc	tcctaagtga	agccctggct	cagtattgcc	atccaaatgc	gccatgctga	240
aagaggggtt	tgcatcctgg	tcagatnaag	aagcaatggt	gtgctgagga	aatcccatc	300
gaataagtga	gcattcagaa	cttgagctag	caggaggagg	actaagatga	tgtgtgagca	360
actctttgta	atggctttca	tctaaaataa	catggtacgt	gccaccagtt	tcacgagcaa	420
gtacagtga	aacgcgaact	tctgcagaca	atccaataac	agatactcta	atttttagctg	480
cctttagggt	cttgattaaa	tcataaatat	tagatggatc	gcaagttgta	aggntgctaa	540
aagatgatta	gtacttctcg	acttgtatgt	ccaggcatgt	tgttttaaan	tctgccttag	600
nccctgctta	ggggaatttt	ttaaagaagat	ggctctccat	gttcanggtc	aatcacnaat	660
tgcc						664

<210> 278

<211> 452

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(452)

<223> n = A,T,C or G

<400> 278

tgacctgaca	ttgaggaaga	gcacacacct	ctgaaattcc	ttaggttcag	aagggcattt	60
gacacagagt	gggcctctga	taattcatga	aatgcattct	gaagtcaccc	agaatggagg	120
ctgcaatctg	ctgtgctttg	ggggttgcc	cactgtgctc	ctggatatca	cacaaaagct	180
gcaatccttc	ttcttcaact	aacattttgc	agtatttgct	gggattttta	ctgcagacat	240
gatacatagc	ccatagtgcc	cagagctgaa	cctctggttg	agagaagttg	ccaaggagcg	300
ggaaaaatgt	cttgaaagat	ctataggtca	ccaatgctgt	catcttacia	cttgaacttg	360
gccaatctctg	tatggttgca	tgcagatctt	ggagaagagt	acgcctctgg	aagtcacggg	420
atatccaaan	ctgtctgtca	gatgtcaggt	ca			452

<210> 279

<211> 274

<212> DNA
<213> Homo sapien

<400> 279

tttttttttt	ttcggcaagg	caaatttact	tctgcaaaag	ggtgctgctt	gcacttttgg	60
ccactgcgag	agcacaccaa	acaaagtagg	gaaggggttt	ttatccctaa	cgcggttatt	120
ccctggttct	gtgtcgtgtc	cccattggct	ggagtcagac	tgcacaatct	acactgaccc	180
aactggctac	tgtttaaaat	tgaatatgaa	taattaggta	ggaaggggga	ggctgtttgt	240
tacggtacaa	gacgtgtttg	ggcatgtcag	gtca			274

<210> 280
<211> 272
<212> DNA
<213> Homo sapien

<400> 280

tacctgacat	ggagaaataa	cttgtagtat	tttgcggtgca	atggaatact	atatgagggt	60
gaaaatgaat	gaactagcaa	tgcgtgtatc	aacatgaata	aatcccaaaa	acataataat	120
gttgaatgga	aaaggtgagt	ttcagaagga	tatatatgcc	ctctaaatcc	atttatgtaa	180
acctttaaaa	aactacatta	tttatgggtca	taagtccatc	cagaaaatat	ttaaaaacct	240
acatgggatt	gataactact	gatgtcaggt	ca			272

<210> 281
<211> 431
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(431)
<223> n = A,T,C or G

<400> 281

tttttttttt	ttggccaata	gcatgattta	aacattggaa	aaagtcaaat	gagcaatgcg	60
aatttttatg	ttctcttgaa	taatcaaaag	agtaggcaac	attggttcct	cattcttgaa	120
tagcattaat	cagaaaatat	tgcatagcct	ctagcctcct	tagagtaggt	gtgctctctc	180
aaatatatca	tagtcccaca	gtttatttca	tgtatatattt	ctgcctgaat	cacatagaca	240
tttgaatttg	caacgcctga	tgtaaatata	taaattctta	ccaatcagaa	acatagcaag	300
aaattcaggg	acttgggtcat	yatcagggtg	tgacagcana	tcctgtara	aacactgata	360
cacactcaca	cacgtatgca	acgtggagat	gtcgcyttww	kkktwywcm	rmrycrwcn	420
aatcacttan	n					431

<210> 282
<211> 98
<212> DNA
<213> Homo sapien

<400> 282

attcgattcg	atgcttgagc	ccaggagtgc	aagactgcag	tgagccactg	cacttcaggc	60
tggacaacag	agcgagtccc	tgtgccaaaa	aaaaaaaa			98

<210> 283
<211> 764
<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(764)

<223> n = A,T,C or G

<400> 283

tttttttttt	ttcgcaagca	cgtgcacttt	attgaatgac	actgtagaca	ggtgtgtggg	60
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cccnnaacttt	atttaccttt	ncaccctagc	ncctctacttn	acccancnc	tcctacctcc	720
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<210> 284

<211> 157

<212> DNA

<213> Homo sapien

<400> 284

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<210> 285

<211> 150

<212> DNA

<213> Homo sapien

<400> 285

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<210> 286

<211> 219

<212> DNA

<213> Homo sapien

<400> 286

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gcaaccttgg	ttaggatcaa	tccaatattc	accatctggg	aagtcaggat	ggctgagttg	180
caggtcttta	caagttcggg	ctggattggg	ctgagtaca			219

<210> 287

<211> 196
 <212> DNA
 <213> Homo sapien

<400> 287
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 actgtgagag agtacatttc tcttggttta agccaagaga atctgtcttt tggctactta 180
 tatcatagcc tcaaga 196

<210> 288
 <211> 199
 <212> DNA
 <213> Homo sapien

<400> 288
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 taggtactga agattcaagt gaccgagatg ctagcccttg ggttcaagtg atccctctcc 180
 cagagtgcac tggactgaa 199

<210> 289
 <211> 182
 <212> DNA
 <213> Homo sapien

<400> 289
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 gtattataat cttagggacc accattatat atgtgggtcca tcattggcca aaaaaaaaaa 180
 aa 182

<210> 290
 <211> 1646
 <212> DNA
 <213> Homo sapien

<400> 290
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<210> 291

<211> 1851

<212> DNA

<213> Homo sapien

<400> 291

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<210> 292

<211> 1851

<212> DNA

<213> Homo sapien

<400> 292

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<210> 293

<211> 668

<212> DNA

<213> Homo sapien

<400> 293

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accrtataag	agcagtgtct	tggccattaa	tttatctttc	attttagaca	gcrtagtgya	180
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cgtctgtcca	gcaggagttt	tactacttct	gaattcccat	tggcagaggg	cagatgtaga	540
gcagtcctat	gagagtgaga	agacttttta	ggaaattgta	gtgcactagc	tacagccata	600
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aaaaaaaa						668

<210> 294
 <211> 1512
 <212> DNA
 <213> Homo sapien

<400> 294

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tccatgccgg	ctgcttcttc	tgtgaagaag	ccatttggtc	tcaggagcaa	gatgggcaag	300
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<210> 295
 <211> 1853
 <212> DNA
 <213> Homo sapien

<400> 295

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<210> 296

<211> 2184

<212> DNA

<213> Homo sapien

<400> 296

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<210> 297
 <211> 1855
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1855)
 <223> n = A,T,C or G

<400> 297						
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<210> 298
 <211> 1059
 <212> DNA
 <213> Homo sapien

<400> 298

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gcgcttgrgg agactmcgat gacagygcct tcatggagcc caggtaccac gtccgtggag      180
aagatctgga caagctccac agagctgccc tgggtggggta aagtccccag aaaggatctc      240
atcgatcatgc tcagggacac tgaygtgaac aagarggaca agcaaaagag gactgctcta      300
catctggcct ctgccaatgg gaattcagaa gtagtaaaac tcstgctgga cagacgatgt      360
caacttaatg tccttgacaa caaaaagagg acagctctga yaaaggccgt acaatgccag      420
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cttcaaaata ctgaaatgca ttcattttaa cattgacgtg tgtaagggcc agtcttccgt      660
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<210> 299

<211> 329

<212> PRT

<213> Homo sapien

<400> 299

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Leu His Leu Ala Gly Ser Asp Leu Leu Ser Arg Ser Leu Met Ala Glu
 20          25          30
Glu Tyr Thr Ile Val His Ala Ser Phe Ile Ser Cys Ile Ser Ser Ser
 35          40          45
Leu Asp Gly Gln Gly Glu Arg Gln Glu Gln Arg Gly His Phe Trp Arg
 50          55          60
Pro Gln Arg Leu Leu Cys Glu Asp Ala Trp Glu Gln Glu Val Gln Val
 65          70          75          80
Val Leu Pro Leu Leu Pro Leu Leu Gln Gly Ser Gly Lys Ser Asn Val
 85          90          95
Val Ala Trp Gly Asp Tyr Asp Asp Ser Ala Phe Met Asp Pro Arg Tyr
100          105          110
His Val His Gly Glu Asp Leu Asp Lys Leu His Arg Ala Ala Trp Trp
115          120          125
Gly Lys Val Pro Arg Lys Asp Leu Ile Val Met Leu Arg Asp Thr Asp
130          135          140
Val Asn Lys Arg Asp Lys Gln Lys Arg Thr Ala Leu His Leu Ala Ser
145          150          155          160
Ala Asn Gly Asn Ser Glu Val Val Lys Leu Val Leu Asp Arg Arg Cys
165          170          175
Gln Leu Asn Val Leu Asp Asn Lys Lys Arg Thr Ala Leu Thr Lys Ala
180          185          190
Val Gln Cys Gln Glu Asp Glu Cys Ala Leu Met Leu Leu Glu His Gly
195          200          205
Thr Asp Pro Asn Ile Pro Asp Glu Tyr Gly Asn Thr Thr Leu His Tyr

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      210                      215                      220
Ala Val Tyr Asn Glu Asp Lys Leu Met Ala Lys Ala Leu Leu Leu Tyr
225                      230                      235                      240
Gly Ala Asp Ile Glu Ser Lys Asn Lys His Gly Leu Thr Pro Leu Leu
                      245                      250                      255
Leu Gly Ile His Glu Gln Lys Gln Gln Val Val Lys Phe Leu Ile Lys
                      260                      265                      270
Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg Tyr Gly Arg Thr Ala Leu
                      275                      280                      285
Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile Val Ser Pro Leu Leu
290                      295                      300
Glu Gln Asn Val Asp Val Ser Ser Gln Asp Leu Glu Arg Arg Pro Glu
305                      310                      315                      320
Ser Met Leu Phe Leu Val Ile Ile Met
                      325

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<210> 300
<211> 148
<212> PRT
<213> Homo sapien

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<220>
<221> VARIANT
<222> (1)...(148)
<223> Xaa = Any Amino Acid

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      <400> 300
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                      20                      25                      30
Asp Leu Ile Val Met Leu Arg Asp Thr Asp Val Asn Lys Xaa Asp Lys
                      35                      40                      45
Gln Lys Arg Thr Ala Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu
50                      55                      60
Val Val Lys Leu Xaa Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp
65                      70                      75                      80
Asn Lys Lys Arg Thr Ala Leu Xaa Lys Ala Val Gln Cys Gln Glu Asp
                      85                      90                      95
Glu Cys Ala Leu Met Leu Leu Glu His Gly Thr Asp Pro Asn Ile Pro
100                      105                      110
Asp Glu Tyr Gly Asn Thr Thr Leu His Tyr Ala Xaa Tyr Asn Glu Asp
115                      120                      125
Lys Leu Met Ala Lys Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser
130                      135                      140
Lys Asn Lys Val
145

```

```

<210> 301
<211> 1155
<212> DNA
<213> Homo sapien

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<400> 301

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gtcagccttc	tacttgagca	aaatattgat	gtatcttctc	aagatctatc	tggacagacg	1020
gccagagagt	atgctgtttc	tagtcatcat	catgtaattt	gccagttact	ttctgactac	1080
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accagaaata	aataa					1155

<210> 302

<211> 2000

<212> DNA

<213> Homo sapien

<400> 302

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aaaaaaaaaa	aaaaaaaaaa					2000

<210> 303
 <211> 2040
 <212> DNA
 <213> Homo sapien

<400> 303

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<210> 304
 <211> 384
 <212> PRT
 <213> Homo sapien

<400> 304
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 20 25 30
 Pro Cys Cys Arg Glu Ser Gly Lys Ser Asn Val Gly Thr Ser Gly Asp
 35 40 45
 His Asp Asp Ser Ala Met Lys Thr Leu Arg Ser Lys Met Gly Lys Trp
 50 55 60
 Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Val
 65 70 75 80
 Gly Ala Ser Gly Asp His Asp Asp Ser Ala Met Lys Thr Leu Arg Asn
 85 90 95
 Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly Ser
 100 105 110
 Gly Lys Ser Lys Val Gly Ala Trp Gly Asp Tyr Asp Asp Ser Ala Phe
 115 120 125
 Met Glu Pro Arg Tyr His Val Arg Gly Glu Asp Leu Asp Lys Leu His
 130 135 140
 Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp Leu Ile Val Met
 145 150 155 160
 Leu Arg Asp Thr Asp Val Asn Lys Lys Asp Lys Gln Lys Arg Thr Ala
 165 170 175
 Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu Val Val Lys Leu Leu
 180 185 190
 Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg Thr
 195 200 205
 Ala Leu Ile Lys Ala Val Gln Cys Gln Glu Asp Glu Cys Ala Leu Met
 210 215 220
 Leu Leu Glu His Gly Thr Asp Pro Asn Ile Pro Asp Glu Tyr Gly Asn
 225 230 235 240
 Thr Thr Leu His Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala Lys
 245 250 255
 Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys His Gly
 260 265 270
 Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln Lys Gln Gln Val Val
 275 280 285
 Lys Phe Leu Ile Lys Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg Tyr
 290 295 300
 Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile
 305 310 315 320
 Val Ser Leu Leu Leu Glu Gln Asn Ile Asp Val Ser Ser Gln Asp Leu
 325 330 335
 Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser Ser His His His Val
 340 345 350
 Ile Cys Gln Leu Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys Ile
 355 360 365
 Ser Ser Glu Asn Ser Asn Pro Glu Asn Val Ser Arg Thr Arg Asn Lys
 370 375 380

<210> 305

<211> 656

<212> PRT

<213> Homo sapien

	<400> 305														
Met 1	Val	Val	Glu	Val	Asp	Ser	Met	Pro	Ala	Ala	Ser	Ser	Val	Lys	Lys
				5					10					15	
Pro	Phe	Gly	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp	Cys	Cys	Arg	Cys	Phe
			20					25					30		
Pro	Cys	Cys	Arg	Glu	Ser	Gly	Lys	Ser	Asn	Val	Gly	Thr	Ser	Gly	Asp
		35					40					45			
His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp
	50					55					60				
Cys	Arg	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser	Gly	Lys	Ser	Asn	Val
65					70					75				80	
Gly	Ala	Ser	Gly	Asp	His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Asn
				85					90					95	
Lys	Met	Gly	Lys	Trp	Cys	Cys	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser
			100					105					110		
Gly	Lys	Ser	Lys	Val	Gly	Ala	Trp	Gly	Asp	Tyr	Asp	Asp	Ser	Ala	Phe
		115					120				125				
Met	Glu	Pro	Arg	Tyr	His	Val	Arg	Gly	Glu	Asp	Leu	Asp	Lys	Leu	His
	130					135				140					
Arg	Ala	Ala	Trp	Trp	Gly	Lys	Val	Pro	Arg	Lys	Asp	Leu	Ile	Val	Met
145					150					155				160	
Leu	Arg	Asp	Thr	Asp	Val	Asn	Lys	Lys	Asp	Lys	Gln	Lys	Arg	Thr	Ala
				165					170					175	
Leu	His	Leu	Ala	Ser	Ala	Asn	Gly	Asn	Ser	Glu	Val	Val	Lys	Leu	Leu
		180						185					190		
Leu	Asp	Arg	Cys	Gln	Leu	Asn	Val	Leu	Asp	Asn	Lys	Lys	Arg	Thr	
		195				200					205				
Ala	Leu	Ile	Lys	Ala	Val	Gln	Cys	Gln	Glu	Asp	Glu	Cys	Ala	Leu	Met
	210					215				220					
Leu	Leu	Glu	His	Gly	Thr	Asp	Pro	Asn	Ile	Pro	Asp	Glu	Tyr	Gly	Asn
225					230					235				240	
Thr	Thr	Leu	His	Tyr	Ala	Ile	Tyr	Asn	Glu	Asp	Lys	Leu	Met	Ala	Lys
			245						250					255	
Ala	Leu	Leu	Leu	Tyr	Gly	Ala	Asp	Ile	Glu	Ser	Lys	Asn	Lys	His	Gly
		260						265					270		
Leu	Thr	Pro	Leu	Leu	Leu	Gly	Val	His	Glu	Gln	Lys	Gln	Gln	Val	Val
		275					280					285			
Lys	Phe	Leu	Ile	Lys	Lys	Lys	Ala	Asn	Leu	Asn	Ala	Leu	Asp	Arg	Tyr
	290					295				300					
Gly	Arg	Thr	Ala	Leu	Ile	Leu	Ala	Val	Cys	Cys	Gly	Ser	Ala	Ser	Ile
305					310				315						320
Val	Ser	Leu	Leu	Leu	Glu	Gln	Asn	Ile	Asp	Val	Ser	Ser	Gln	Asp	Leu
			325						330					335	
Ser	Gly	Gln	Thr	Ala	Arg	Glu	Tyr	Ala	Val	Ser	Ser	His	His	His	Val
			340					345					350		

Glu Glu Met Lys Lys His Glu Ser Asn Asn Val Gly Leu Leu Glu Asn
 420 425 430
 Leu Thr Asn Gly Val Thr Ala Gly Asn Gly Asp Asn Gly Leu Ile Pro
 435 440 445
 Gln Arg Lys Ser Arg Thr Pro Glu Asn Gln Gln Phe Pro Asp Asn Glu
 450 455 460
 Ser Glu Glu Tyr His Arg Ile Cys Glu Leu Val Ser Asp Tyr Lys Glu
 465 470 475 480
 Lys Gln Met Pro Lys Tyr Ser Ser Glu Asn Ser Asn Pro Glu Gln Asp
 485 490 495
 Leu Lys Leu Thr Ser Glu Glu Glu Ser Gln Arg Leu Glu Gly Ser Glu
 500 505 510
 Asn Gly Gln Pro Glu Leu Glu Asn Phe Met Ala Ile Glu Glu Met Lys
 515 520 525
 Lys His Gly Ser Thr His Val Gly Phe Pro Glu Asn Leu Thr Asn Gly
 530 535 540
 Ala Thr Ala Gly Asn Gly Asp Asp Gly Leu Ile Pro Pro Arg Lys Ser
 545 550 555 560
 Arg Thr Pro Glu Ser Gln Gln Phe Pro Asp Thr Glu Asn Glu Glu Tyr
 565 570 575
 His Ser Asp Glu Gln Asn Asp Thr Gln Lys Gln Phe Cys Glu Glu Gln
 580 585 590
 Asn Thr Gly Ile Leu His Asp Glu Ile Leu Ile His Glu Glu Lys Gln
 595 600 605
 Ile Glu Val Val Glu Lys Met Asn Ser Glu Leu Ser Leu Ser Cys Lys
 610 615 620
 Lys Glu Lys Asp Ile Leu His Glu Asn Ser Thr Leu Arg Glu Glu Ile
 625 630 635 640
 Ala Met Leu Arg Leu Glu Leu Asp Thr Met Lys His Gln Ser Gln Leu
 645 650 655

<210> 306

<211> 671

<212> PRT

<213> Homo sapien

<400> 306

Met Val Val Glu Val Asp Ser Met Pro Ala Ala Ser Ser Val Lys Lys
 1 5 10 15
 Pro Phe Gly Leu Arg Ser Lys Met Gly Lys Trp Cys Cys Arg Cys Phe
 20 25 30
 Pro Cys Cys Arg Glu Ser Gly Lys Ser Asn Val Gly Thr Ser Gly Asp
 35 40 45
 His Asp Asp Ser Ala Met Lys Thr Leu Arg Ser Lys Met Gly Lys Trp
 50 55 60
 Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Val
 65 70 75 80
 Gly Ala Ser Gly Asp His Asp Asp Ser Ala Met Lys Thr Leu Arg Asn
 85 90 95
 Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly Ser
 100 105 110
 Gly Lys Ser Lys Val Gly Ala Trp Gly Asp Tyr Asp Asp Ser Ala Phe
 115 120 125
 Met Glu Pro Arg Tyr His Val Arg Gly Glu Asp Leu Asp Lys Leu His

130	135	140
Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp Leu Ile Val Met		
145	150	155
Leu Arg Asp Thr Asp Val Asn Lys Lys Asp Lys Gln Lys Arg Thr Ala		160
	165	170
Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu Val Val Lys Leu Leu		175
	180	185
Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg Thr		190
	195	200
Ala Leu Ile Lys Ala Val Gln Cys Gln Glu Asp Glu Cys Ala Leu Met		205
	210	215
Leu Leu Glu His Gly Thr Asp Pro Asn Ile Pro Asp Glu Tyr Gly Asn		220
225	230	235
Thr Thr Leu His Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala Lys		240
	245	250
Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys His Gly		255
	260	265
Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln Lys Gln Gln Val Val		270
	275	280
Lys Phe Leu Ile Lys Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg Tyr		285
	290	295
Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile		300
305	310	315
Val Ser Leu Leu Leu Glu Gln Asn Ile Asp Val Ser Ser Gln Asp Leu		320
	325	330
Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser Ser His His His Val		335
	340	345
Ile Cys Gln Leu Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys Ile		350
	355	360
Ser Ser Glu Asn Ser Asn Pro Glu Gln Asp Leu Lys Leu Thr Ser Glu		365
	370	375
Glu Glu Ser Gln Arg Phe Lys Gly Ser Glu Asn Ser Gln Pro Glu Lys		380
385	390	395
Met Ser Gln Glu Pro Glu Ile Asn Lys Asp Gly Asp Arg Glu Val Glu		400
	405	410
Glu Glu Met Lys Lys His Glu Ser Asn Asn Val Gly Leu Leu Glu Asn		415
	420	425
Leu Thr Asn Gly Val Thr Ala Gly Asn Gly Asp Asn Gly Leu Ile Pro		430
	435	440
Gln Arg Lys Ser Arg Thr Pro Glu Asn Gln Gln Phe Pro Asp Asn Glu		445
	450	455
Ser Glu Glu Tyr His Arg Ile Cys Glu Leu Val Ser Asp Tyr Lys Glu		460
465	470	475
Lys Gln Met Pro Lys Tyr Ser Ser Glu Asn Ser Asn Pro Glu Gln Asp		480
	485	490
Leu Lys Leu Thr Ser Glu Glu Glu Ser Gln Arg Leu Glu Gly Ser Glu		495
	500	505
Asn Gly Gln Pro Glu Lys Arg Ser Gln Glu Pro Glu Ile Asn Lys Asp		510
	515	520
Gly Asp Arg Glu Leu Glu Asn Phe Met Ala Ile Glu Glu Met Lys Lys		525
	530	535
His Gly Ser Thr His Val Gly Phe Pro Glu Asn Leu Thr Asn Gly Ala		540
545	550	555
Thr Ala Gly Asn Gly Asp Asp Gly Leu Ile Pro Pro Arg Lys Ser Arg		560

565 570 575
 Thr Pro Glu Ser Gln Gln Phe Pro Asp Thr Glu Asn Glu Glu Tyr His
 580 585 590
 Ser Asp Glu Gln Asn Asp Thr Gln Lys Gln Phe Cys Glu Glu Gln Asn
 595 600 605
 Thr Gly Ile Leu His Asp Glu Ile Leu Ile His Glu Glu Lys Gln Ile
 610 615 620
 Glu Val Val Glu Lys Met Asn Ser Glu Leu Ser Leu Ser Cys Lys Lys
 625 630 635 640
 Glu Lys Asp Ile Leu His Glu Asn Ser Thr Leu Arg Glu Glu Ile Ala
 645 650 655
 Met Leu Arg Leu Glu Leu Asp Thr Met Lys His Gln Ser Gln Leu
 660 665 670

<210> 307
 <211> 800
 <212> DNA
 <213> Homo sapien

<400> 307

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acttcatttt	tggtacataa	catctttata	ggacaggggt	aaaatcccaa	tactaacagg	120
agaatgctta	ggactctaac	aggtttttga	gaatgtgttg	gtaagggcca	ctcaatccaa	180
tttttcttgg	tcctccttgt	ggtctaggag	gacaggcaag	ggtgcagatt	ttcaagaatg	240
catcagtaag	ggccactaaa	tccgaccttc	ctcgttcctc	cttgtggtct	gggaggaaaa	300
ctagtgtttc	tgttgctgtg	tcagttagca	caactattcc	gatcagcagg	gtccagggac	360
cactgcaggt	tcttgggcag	ggggagaaac	aaaacaaacc	aaaaccatgg	gcrgttttgt	420
ctttcagatg	ggaaacactc	aggcatcaac	aggtcacct	ttgaaatgca	tcctaagcca	480
atgggacaaa	tttgaccac	aaacctgga	aaaagagggtg	gtcattttt	tttgactat	540
ggcttgccc	caacattctc	tctctgatgg	ggaaaaatgg	ccacctgagg	gaagtacaga	600
ttacaatact	atcctgcagc	ttgacctttt	ctgtaagagg	gaaggcaaat	ggagtgaat	660
accttatgtc	caagctttct	tttcattgaa	ggagaataca	ctatgcaaag	cttgaaattt	720
acatcccaca	ggaggacctc	tcagcttacc	cccatactct	agcctcccta	tagctcccct	780
tcctattagt	gataagcctc					800

<210> 308
 <211> 102
 <212> PRT
 <213> Homo sapien

<220>

<221> VARIANT

<222> (1)...(102)

<223> Xaa = Any Amino Acid

<400> 308

Met Gly Xaa Phe Val Phe Gln Met Gly Asn Thr Gln Ala Ser Thr Gly	
1 5 10 15	
Ser Pro Leu Lys Cys Ile Leu Ser Gln Trp Asp Lys Phe Asp Pro Gln	
20 25 30	
Thr Leu Glu Lys Glu Val Ala His Phe Phe Cys Thr Met Ala Trp Pro	
35 40 45	
Gln His Ser Leu Ser Asp Gly Glu Lys Trp Pro Pro Glu Gly Ser Thr	
50 55 60	

Asp Tyr Asn Thr Ile Leu Gln Leu Asp Leu Phe Cys Lys Arg Glu Gly
 65 70 75 80
 Lys Trp Ser Glu Ile Pro Tyr Val Gln Ala Phe Phe Ser Leu Lys Glu
 85 90 95
 Asn Thr Leu Cys Lys Ala
 100

<210> 309
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 309
 Leu Met Ala Glu Glu Tyr Thr Ile Val
 1 5

<210> 310
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 310
 Lys Leu Met Ala Lys Ala Leu Leu Leu
 1 5

<210> 311
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 311
 Gly Leu Thr Pro Leu Leu Leu Gly Ile
 1 5

<210> 312
 <211> 10
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 312
 Lys Leu Val Leu Asp Arg Arg Cys Gln Leu
 1 5 10

<210> 313
 <211> 1852
 <212> DNA
 <213> Homo sapiens

<400> 313
 ggcacgagaa ttaaaaccct cagcaaaaca ggcatagaag ggacatacct taaagtaata 60
 aaaaccacct atgacaagcc cacagccaac ataatactaa atggggaaaa gttagaagca 120
 ttctctctga gaactgcaac aataaataca aggatgctgg attttgtcaa atgccttttc 180
 tgtgtctgtt gagatgctta tgtgactttg cttttaattc tgtttatgtg attatcacat 240
 ttattgactt gcctgtgtta gaccggaaga gctgggggtg ttctcaggag ccaccgtgtg 300
 ctgcggcagc ttggggataa cttgaggctg catcactggg gaagaaacac aytccctgtcc 360
 gtggcgctga tggctgagga cagagcttca gtgtggcttc tctgcgactg gottcttcgg 420
 ggagttcttc cttcatagt ccatcatatg gctccagagg aaaattatat tattttgtta 480
 tggatgaaga gtattacgtt gtgcagatat actgcagtgt cttcatctct tgatgtgtga 540
 ttgggtaggt tccaccatgt tgccgcagat gacatgattt cagtacctgt gtctggctga 600
 aaagtgtttg tttgtgaatg gatattgtgg tttctggatc tcatcctctg tgggtggaca 660
 gctttctcca ccttgctgga agtgacctgc tgtccagaag tttgatggct gaggagtata 720
 ccatcgtgca tgcacttttc atttctgca tttcttctc cctggatgga cagggggagc 780
 ggcaagagca acgtgggcac ttctggagac cacaacgact cctctgtgaa gacgcttggg 840
 agcaagaggt gcaagtgggt ctgccactgc tccccctgct gcagggggag cggcaagagc 900
 aacgtggctg cttggggaga ctacgatgac agcgccttca tggatcccag gtaccacgtc 960
 catggagaag atctggacaa gctccacaga gctgcctggt ggggtaaagt cccagaaaag 1020
 gatctcatcg tcatgctcag ggacacggat gtgaacaaga gggacaagca aaagaggact 1080
 gctctacatc tggcctctgc caatgggaat tcagaagtag taaaactcgt gctggacaga 1140
 cgatgtcaac ttaatgtcct tgacaacaaa aagaggacag ctctgacaaa ggccgtacaa 1200
 tgccaggaag atgaatgtgc gttaatgttg ctggaacatg gcactgatcc aaatatcca 1260
 gatgagtatg gaaataccac tctacactat gctgtctaca atgaagataa attaatggcc 1320
 aaagcactgc tcttatacgg tctgatatc gaatcaaaaa acaagcatgg cctcacacca 1380
 ctgctacttg gtatacatga gcaaaaacag caagtgggtga aatttttaat caagaaaaaa 1440
 gcgaatttaa atgcgctgga tagatatgga agaactgctc tcatacttgc tgtatgttgt 1500
 ggatcagcaa gtatagtcag ccctctactt gagcaaaatg ttgatgtatc ttctcaagat 1560
 ctggaaagac ggccagagag tatgctgttt ctagtcatca tcatgtaatt tgccagttac 1620
 tttctgacta caaagaaaaa cagatgttaa aaatctcttc tgaaaacagc aatccagaac 1680
 aagacttaaa gctgacatca gaggaagagt cacaagggt taaaggagt gaaaacagcc 1740
 agccagagct agaagattta tggctattga agaagaatga agaacacgga agtactcatg 1800
 tgggattccc agaaaacctg actaacgggtg ccgctgctgg caatggtgat ga 1852

<210> 314
 <211> 879
 <212> DNA
 <213> Homo sapiens

<400> 314
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 aacgtgggca cttctggaga ccacaacgac tcctctgtga agacgcttgg gagcaagagg 120
 tgcaagtggg gctgccactg cttccctctg tgcaggggga gcggcaagag caacgtgggtc 180
 gcttggggag actacgatga cagcgccttc atggatccca ggtaccacgt ccatggagaa 240
 gatctggaca agctccacag agctgcctgg tggggtaaag tccccagaaa ggatctcatc 300
 gtcattgctca gggacacgga tgtgaacaag agggacaagc aaaagaggac tgctctacat 360
 ctggcctctg ccaatgggaa ttcagaagta gtaaaactcg tgctggacag acgatgtcaa 420
 cttaatgtcc ttgacaacaa aaagaggaca gctctgacaa aggcgtaca atgccaggaa 480
 gatgaatgtg cgttaatgtt gctggaacat ggcactgatc caaatattcc agatgagtat 540

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ggaaatacca ctctacacta tgctgtctac aatgaagata aattaatggc caaagcactg 600
ctcttatacg gtgctgatat cgaatcaaaa aacaagcatg gcctcacacc actgctactt 660
ggtatacatg agcaaaaaca gcaagtgggtg aaatttttaa tcaagaaaaa agcgaattta 720
aatgcgctgg atagatatgg aagaactgct ctcatacttg ctgtatgttg tggatcagca 780
agtatagtca gccctctact tgagcaaaat gttgatgtat cttctcaaga tctggaaaga 840
cggccagaga gtatgctgtt tctagtcatc atcatgtaa 879

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<210> 315

<211> 292

<212> PRT

<213> Homo sapiens

<400> 315

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Met His Leu Ser Phe Pro Ala Phe Leu Pro Pro Trp Met Asp Arg Gly
      5                      10                      15

Ser Gly Lys Ser Asn Val Gly Thr Ser Gly Asp His Asn Asp Ser Ser
      20                      25                      30

Val Lys Thr Leu Gly Ser Lys Arg Cys Lys Trp Cys Cys His Cys Phe
      35                      40                      45

Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Val Val Ala Trp Gly Asp
      50                      55                      60

Tyr Asp Asp Ser Ala Phe Met Asp Pro Arg Tyr His Val His Gly Glu
      65                      70                      75                      80

Asp Leu Asp Lys Leu His Arg Ala Ala Trp Trp Gly Lys Val Pro Arg
      85                      90                      95

Lys Asp Leu Ile Val Met Leu Arg Asp Thr Asp Val Asn Lys Arg Asp
      100                     105                     110

Lys Gln Lys Arg Thr Ala Leu His Leu Ala Ser Ala Asn Gly Asn Ser
      115                     120                     125

Glu Val Val Lys Leu Val Leu Asp Arg Arg Cys Gln Leu Asn Val Leu
      130                     135                     140

Asp Asn Lys Lys Arg Thr Ala Leu Thr Lys Ala Val Gln Cys Gln Glu
      145                     150                     155                     160

Asp Glu Cys Ala Leu Met Leu Leu Glu His Gly Thr Asp Pro Asn Ile
      165                     170                     175

Pro Asp Glu Tyr Gly Asn Thr Thr Leu His Tyr Ala Val Tyr Asn Glu
      180                     185                     190

Asp Lys Leu Met Ala Lys Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu
      195                     200                     205

Ser Lys Asn Lys His Gly Leu Thr Pro Leu Leu Leu Gly Ile His Glu
      210                     215                     220

```

Gln Lys Gln Gln Val Val Lys Phe Leu Ile Lys Lys Lys Ala Asn Leu
225 230 235 240

Asn Ala Leu Asp Arg Tyr Gly Arg Thr Ala Leu Ile Leu Ala Val Cys
245 250 255

Cys Gly Ser Ala Ser Ile Val Ser Pro Leu Leu Glu Gln Asn Val Asp
260 265 270

Val Ser Ser Gln Asp Leu Glu Arg Arg Pro Glu Ser Met Leu Phe Leu
275 280 285

Val Ile Ile Met
290

<210> 316

<211> 584

<212> DNA

<213> Homo sapiens

<400> 316

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agttgggcca aattecctc cccctacagc ttgaagggga cataaccaat agcctggggg 60
ttttttgtgg toctttggag atttctttgc ttattttctt ctgggtgggg gtgattagag 120
gaggcttata actaatagga aggggagcta tagggagggt aggatatggg ggtaagctga 180
gaggtcctcc tgtgggatgt aaatttcaag ctttgcatag tgtattctcc ttcaatgaaa 240
agaaagcttg gacataaggt atttcactcc atttgccttc cctcttacag aaaagggtcaa 300
gctgcaggat agtattgtaa tctgtacttc cctcagggtg ccatttttcc ccatcagaga 360
gagaatgttg gggccaagcc atagtgcaga aaaaaaatg agccacctct tttccagggt 420
tttgtgggtc aaatttgtcc cattggctta ggatgcattt caaagggtgag cctgttgatg 480
cctgagtgtt tccatctga aagacaaaac tgcccattgt tttggtttgt tttgtttctc 540
cccctgcccc agaactatca aactcctgag ccaacaacta aaaa 584
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<210> 317

<211> 829

<212> DNA

<213> Homo sapiens

<400> 317

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attagcttcc gcttctgaca acactagaga tccctcccct cctcagggt atggccctcc 60
acttcatttt tggtagataa catctttata ggacaggggt aaaatcccaa tactaacagg 120
agaatgctta ggactctaac aggtttttga gaatgtgttg gtaagggcca ctcaatccaa 180
tttttcttgg toctccttgt ggtctaggag gacaggcaag ggtgcagatt ttcaagaatg 240
catcagtaag ggcactaaa tccgaccttc ctggttctc cttgttgtct gggaggaaaa 300
ctagtgttct tgttgctgtg tcagtgcaga caactattcc gatcagcagg gtccagggac 360
cactgcagggt tcttgggcag ggggagaaac aaaacaaacc aaaaccatgg gcagttttgt 420
ctttcagatg ggaaacactc aggcataaac aggtcacct ttgaaatgca tcctaagcca 480
atgggacaaa tttgacctc aaacctgga aaaagagggt gctcattttt tttgcactat 540
ggcttggccc caacattctc tctctgatgg ggaaaaatgg ccacctgagg gaagtacaga 600
ttacaatact atcctgcagc ttgacctttt ctgtaagagg gaaggcaaag ggagtgaat 660
accttatgtc caagctttct tttcattgaa ggagaatata ctatgcaaag cttgaaattt 720
acatcccaca ggaggacctc tcagcttacc cccatatact agcctcccta tagctcccct 780
tctatttagt gataagcctc ctctaatac cccacccag aagaaaata 829
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<210> 318
 <211> 30
 <212> PRT
 <213> Homo sapien

<400> 318
 Thr Ala Ala Ser Asp Asn Phe Gln Leu Ser Gln Gly Gly Gln Gly Phe
 1 5 10 15
 Ala Ile Pro Ile Gly Gln Ala Met Ala Ile Ala Gly Gln Ile
 20 25 30

<210> 319
 <211> 41
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> PCR primer

<400> 319
 ggctctgtgcc aatgggaact cagaagtagt aaaactcctg c 41

<210> 320
 <211> 41
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> PCR primer

<400> 320
 gcaggagttt tactacttct gagttcccat tggcagaggg c 41

<210> 321
 <211> 60
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> PCR primer

<400> 321
 ggggaattcc cgctgggtgcc gcgcgggcagc cctatgggtgg ttgaggttga 50
 ttccatgccg 60

<210> 322
 <211> 42
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> PCR primer

<400> 322

cccggaattct tattttatttc tggttcttga gacattttct gg

42

<210> 323
 <211> 1590
 <212> DNA
 <213> Homo sapiens

<400> 323

atgcatcacc	atcaccatca	cacggccgcg	tccgataact	tccagctgtc	ccaggggtggg	60
cagggattcg	ccattccgat	cgggcaggcg	atggcgatcg	cgggccagat	caagcttccc	120
acogttcata	tcgggcctac	cgccttcctc	ggcttgggtg	ttgtcgacaa	caacggcaac	180
ggcgacagag	tccaacgcgt	ggtcgggagc	gctcgggcgg	caagtctcgg	catctccacc	240
ggcgacgtga	tcaccgcggt	cgacggcgct	cogatcaact	cggccaccgc	gatggcggac	300
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<210> 324
 <211> 529
 <212> PRT
 <213> Homo sapiens

<400> 324

Met His His His His His His Thr Ala Ala Ser Asp Asn Phe Gln Leu

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			20					25					30		
Ile	Ala	Gly	Gln	Ile	Lys	Leu	Pro	Thr	Val	His	Ile	Gly	Pro	Thr	Ala
		35					40					45			
Phe	Leu	Gly	Leu	Gly	Val	Val	Asp	Asn	Asn	Gly	Asn	Gly	Ala	Arg	Val
	50					55					60				
Gln	Arg	Val	Val	Gly	Ser	Ala	Pro	Ala	Ala	Ser	Leu	Gly	Ile	Ser	Thr
	65				70					75					80
Gly	Asp	Val	Ile	Thr	Ala	Val	Asp	Gly	Ala	Pro	Ile	Asn	Ser	Ala	Thr
				85					90					95	
Ala	Met	Ala	Asp	Ala	Leu	Asn	Gly	His	His	Pro	Gly	Asp	Val	Ile	Ser
			100					105					110		
Val	Thr	Trp	Gln	Thr	Lys	Ser	Gly	Gly	Thr	Arg	Thr	Gly	Asn	Val	Thr
		115					120					125			
Leu	Ala	Glu	Gly	Pro	Pro	Ala	Glu	Phe	Pro	Leu	Val	Pro	Arg	Gly	Ser
	130					135					140				
Pro	Met	Val	Val	Glu	Val	Asp	Ser	Met	Pro	Ala	Ala	Ser	Ser	Val	Lys
	145				150					155					160
Lys	Pro	Phe	Gly	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp	Cys	Cys	Arg	Cys
				165					170					175	
Phe	Pro	Cys	Cys	Arg	Glu	Ser	Gly	Lys	Ser	Asn	Val	Gly	Thr	Ser	Gly
			180					185					190		
Asp	His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Ser	Lys	Met	Gly	Lys
		195					200					205			
Trp	Cys	Arg	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser	Gly	Lys	Ser	Asn
	210					215					220				
Val	Gly	Ala	Ser	Gly	Asp	His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg
	225				230					235				240	
Asn	Lys	Met	Gly	Lys	Trp	Cys	Cys	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly
				245					250					255	
Ser	Gly	Lys	Ser	Lys	Val	Gly	Ala	Trp	Gly	Asp	Tyr	Asp	Asp	Ser	Ala
			260					265					270		
Phe	Met	Glu	Pro	Arg	Tyr	His	Val	Arg	Gly	Glu	Asp	Leu	Asp	Lys	Leu
		275					280					285			
His	Arg	Ala	Ala	Trp	Trp	Gly	Lys	Val	Pro	Arg	Lys	Asp	Leu	Ile	Val

290	295	300
Met Leu Arg Asp Thr	Asp Val Asn Lys Lys Asp	Lys Gln Lys Arg Thr
305	310	315 320
Ala Leu His Leu Ala	Ser Ala Asn Gly Asn Ser Glu Val Val Lys Leu	
	325	330 335
Leu Leu Asp Arg Arg	Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg	
	340	345 350
Thr Ala Leu Ile Lys	Ala Val Gln Cys Gln Glu Asp Glu Cys Ala Leu	
	355	360 365
Met Leu Leu Glu His	Gly Thr Asp Pro Asn Ile Pro Asp Glu Tyr Gly	
	370	375 380
Asn Thr Thr Leu His	Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala	
	385	390 395 400
Lys Ala Leu Leu Leu	Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys His	
	405	410 415
Gly Leu Thr Pro Leu	Leu Leu Gly Val His Glu Gln Lys Gln Gln Val	
	420	425 430
Val Lys Phe Leu Ile	Lys Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg	
	435	440 445
Tyr Gly Arg Thr Ala	Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser	
	450	455 460
Ile Val Ser Leu Leu	Leu Glu Gln Asn Ile Asp Val Ser Ser Gln Asp	
	465	470 475 480
Leu Ser Gly Gln Thr	Ala Arg Glu Tyr Ala Val Ser Ser His His His	
	485	490 495
Val Ile Cys Gln Leu	Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys	
	500	505 510
Ile Ser Ser Glu Asn	Ser Asn Pro Glu Asn Val Ser Arg Thr Arg Asn	
	515	520 525

Lys

<210> 325

<211> 1155

<212> DNA

<213> Homo sapiens

<400> 325

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<210> 326

<211> 384

<212> PRT

<213> Homo sapiens

<400> 326

```

Met Val Ala Glu Val Cys Ser Met Pro Thr Ala Ser Thr Val Lys Lys
          5              10              15

```

```

Pro Phe Asp Leu Arg Ser Lys Met Gly Lys Trp Cys His His Arg Phe
          20              25              30

```

```

Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Met Gly Thr Ser Gly Asp
          35              40              45

```

```

His Asp Asp Ser Phe Met Lys Met Leu Arg Ser Lys Met Gly Lys Cys
          50              55              60

```

```

Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Thr Ser Asn Val
          65              70              75              80

```

```

Gly Thr Ser Gly Asp His Glu Asn Ser Phe Met Lys Met Leu Arg Ser
          85              90              95

```

```

Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly Ser
          100              105              110

```

```

Gly Lys Ser Asn Val Gly Ala Trp Gly Asp Tyr Asp His Ser Ala Phe
          115              120              125

```

```

Met Glu Pro Arg Tyr His Ile Arg Arg Glu Asp Leu Asp Lys Leu His
          130              135              140

```

```

Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp Leu Ile Val Met

```

145		150		155		160
Leu Arg Asp Thr	Asp Met Asn Lys Arg Asp Lys Glu Lys Arg Thr Ala					
	165			170		175
Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu Val Val Gln Leu Leu						
	180			185		190
Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg Thr						
	195			200		205
Ala Leu Ile Lys Ala Ile Gln Cys Gln Glu Asp Glu Cys Val Leu Met						
	210			215		220
Leu Leu Glu His Gly Ala Asp Arg Asn Ile Pro Asp Glu Tyr Gly Asn						
	225			230		235
Thr Ala Leu His Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala Lys						
				245		250
Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys Val Gly						
	260			265		270
Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln Lys Gln Gln Val Val						
	275			280		285
Lys Phe Leu Ile Lys Lys Lys Ala Asn Leu Asn Val Leu Asp Arg Tyr						
	290			295		300
Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile						
	305			310		315
Val Asn Leu Leu Leu Glu Gln Asn Val Asp Val Ser Ser Gln Asp Leu						
	325			330		335
Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser Ser His His His Val						
	340			345		350
Ile Cys Glu Leu Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys Ile						
	355			360		365
Ser Ser Glu Asn Ser Asn Pro Glu Asn Val Ser Arg Thr Arg Asn Lys						
	370			375		380